

Review article

G. TUNNICLIFF

ACTIONS OF SODIUM VALPROATE ON THE CENTRAL NERVOUS SYSTEM

Department of Biochemistry and Molecular Biology, Indiana University School of Medicine, Evansville, Indiana, USA

The branched chain fatty acid, valproate, has a number of distinct pharmacological effects on the central nervous system. In experimental animals it showed clear anticonvulsant activity, an observation which led to its major clinical use as an antiepileptic agent, especially in petit mal seizures. More recently, valproate has shown its usefulness in treating mood disorders and migraine headaches. The basis for its clinical efficacy might be related to its ability to enhance central GABAergic neurotransmission or perhaps to its inhibition of Na⁺ channels. Whether each of the distinct therapeutic effects of valproate has the same molecular basis is not known.

Key words: valproate, epilepsy, migraine headaches, mood disorders, GABA, Na⁺ channels.

INTRODUCTION

Valproic acid (also known as 2-propylpentanoic acid or *n*-dipropylacetic acid) is a branched chain fatty acid (*Fig. 1*), and is a liquid at room temperature. Indeed, it is because of its physical properties that we now know this is an important antiepileptic drug. Meunier and colleagues (1) were studying compounds for their effects against drug-induced seizures in experimental animals. These compounds were dissolved in valproic acid before injection. Each of the drugs exhibited anticonvulsant properties and subsequently the investigators showed that it was the vehicle which provided the pharmacological effects. Sodium valproate, the sodium salt, is equally efficacious at suppressing seizures.

In addition to its inhibitory effects on epileptic seizures, sodium valproate is also a useful drug in the suppression of the development of migraine

headaches (2) and as a means of treating patients with emotional disorders such as manic depressive illness or panic disorder (3, 4).

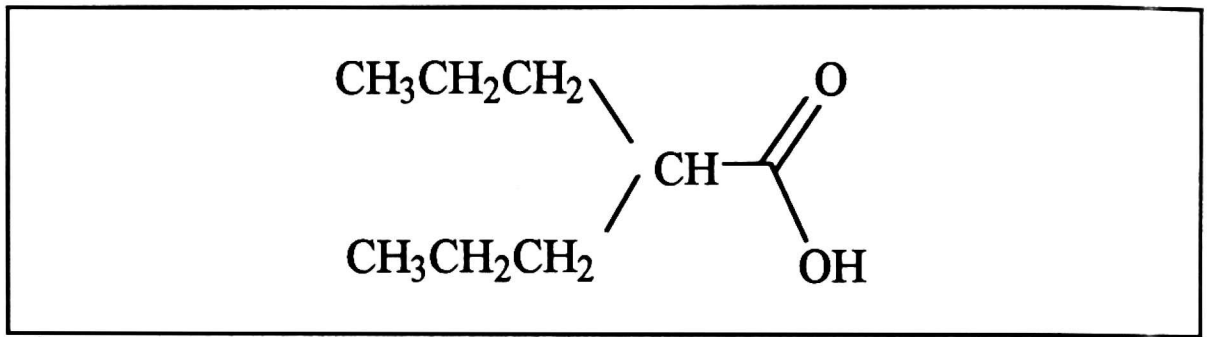


Fig. 1. Chemical structure of valproic acid

PHARMACOLOGY

Antiepileptic Activity

Valproate is unusual as an anticonvulsant since it is effective against most forms of epilepsy, particularly generalized seizures. Moreover, in experimental animals it requires relatively high doses to be effective (plasma levels: 1.4—3.5 mM) yet in humans a much lower dose is adequate (plasma concentration range: 350—700 μ M) (5). In animal models of epilepsy valproate is the most effective against pentylenetetrazol-induced seizures, a model for absence seizures, but it also provides good protection against electroshock-induced convulsions, a model of grand mal epilepsy (6).

Mechanism of Anticonvulsant Effect

The two broad types of mechanisms to be considered are (a) those involving intracellular events, e.g., inhibition of enzyme activity or effects on nucleic acid function, and (b) those involving events at the plasma membrane, e.g., alterations of ion channel or receptor function. These mechanisms are not necessarily mutually exclusive.

1. Effects on intracellular events

Since γ -aminobutyric acid (GABA) is the main inhibitory chemical transmitter in the brain (7), it is intuitive to speculate that it might be associated with seizure activity. Indeed, there is considerable evidence from animal studies that a reduction in GABA function leads to convulsive activity (8, 9). Such strong evidence, though, is lacking in human epilepsy (10). Nevertheless, in principle a drug that can enhance central GABA

neurotransmission should possess antiepileptic properties. The mechanism of action of several clinically useful therapeutic agents support this idea, e.g., phenobarbital, diazepam, and vigabatrin. Consequently, it is not surprising that studies of the potential effects of valproate on the GABA system have received much attention.

The first attempt at elucidating the mechanism of action of valproate by studying effects on GABA was reported by Godin *et al.* (11). They injected 200 mg/kg into rats and after 1 hr the GABA content of whole brain had increased approximately 30%. The rise in GABA levels was attributed to the inhibition of GABA aminotransferase, an effect they also showed to occur *in vitro*, albeit at high inhibitor concentrations. Further experiments (12) confirmed the elevated brain GABA concentrations following valproate treatment and the weak inhibitory action of valproate on GABA aminotransferase. The nature of this inhibition was found to be competitive with respect to GABA. Others observed that sub-chronic treatment of rats with valproate for 10 days led to an increase in GABA concentrations in several brain regions (13); and in human epilepsy GABA levels were elevated in cerebrospinal fluid after valproate administration (14). A detailed kinetic analysis of the inhibition of rabbit brain GABA aminotransferase by valproate supported the competitive nature of the inhibition, yielding a K_i of 42 mM and confirming the low sensitivity of the enzyme to the inhibitor (15). After administering high doses of valproate to audiogenic seizure-susceptible mice, Anlezark *et al.* (16) reported a total lack of response to auditory stimulation, together with a 57% increase in brain GABA levels and a modest reduction in GABA aminotransferase activity 45 min later. Löscher (17) measured GABA aminotransferase activity in synaptosomes isolated from various brain regions after the intraperitoneal injection of valproate, and noted that enzyme activity decreased by about 25% in substantia nigra, with smaller reductions in the pons and medulla. When whole tissue was used, however, no such decreases in enzyme activity were apparent. Interestingly, from an investigation employing neurons and astrocytes in culture, it was found that neuronal GABA aminotransferase was much more sensitive to valproate than the glial enzyme (18). Other studies, though, have failed to show an effect on GABA aminotransferase activity *in vivo* after valproate treatment (19—21).

Hearl and Churchich (22) have presented convincing evidence that GABA aminotransferase and succinic semialdehyde dehydrogenase (SSADH) form a catalytic protein complex which is responsible for the metabolic conversion of GABA to succinate, allowing the carbon skeleton of GABA to enter the citric acid cycle. Compared with its weak effect on GABA aminotransferase, valproate is much more effective at inhibiting SSADH — about 30-times, in fact (23). This has led to the suggestion that the elevation of GABA levels in the brain are the indirect result of the inhibition of SSADH. The actual mechanism

might be a slow-down in GABA aminotransferase activity caused by an accumulation of its product, succinic semialdehyde, and by an actual reversal of the reaction (24). Maitre *et al.* (25) used a series of SSADH inhibitors in order to see if GABA synthesis from succinic semialdehyde could be detected. No synthesis could be measured, but since these were *in vitro* experiments, the relevance of the observations have to be questioned. Even if a reversal of GABA aminotransferase did occur under such conditions, it would not be expected that the concentrations of succinic semialdehyde in the brain would contribute significantly to the reported increases in GABA levels. However, it has been calculated that the plasma concentrations of valproate required for it to be an effective antiepileptic agent are in the range 0.1-0.8 mM (26), and it could be anticipated that levels in the brain would be below these values. Indeed, one study claims that this concentration is about 50 μ M (27). Accordingly, it seems improbable that a direct inhibition of GABA aminotransferase occurs. Even the idea of an inhibition of SSADH *in vivo* to explain the anticonvulsant effects of valproate is difficult to sustain since relatively high K_i values have been reported (0.5 mM to 1.5 mM) (23, 24).

Some studies have indicated that another mechanism to explain the elevation of cerebral GABA concentrations by valproate might be an increase in glutamate decarboxylase activity (28—30). Other investigations have supported this idea. For example, Löscher (31) reported an increase in GABA turnover in certain brain regions after valproate administration, and experiments by Taberner *et al.* (32) demonstrated that the incorporation of 14 C from radiolabelled glucose into GABA was increased in rats receiving valproate. On the other hand, two studies in which the incorporation of 14 C from glutamate into GABA was measured clearly show that [14 C]GABA formation was *inhibited* by valproate (33, 34). Moreover, Godin *et al.* (11) noticed a weak inhibition of glutamate decarboxylase *in vitro*. Hence the notion that the observed increases in GABA levels after valproate dosing are due to an increased synthesis of the amino acid is still open to doubt.

γ -Hydroxybutyrate (GHB) is a metabolite of GABA that possesses certain attributes of an inhibitory neurotransmitter (35). It is synthesized in brain by the catalytic action of succinic semialdehyde reductase. *In vitro* studies have indicated that valproate can inhibit this biosynthesis (36). However, a recent report suggests that a cloned version of the enzyme from rat is insensitive to valproate (37). Consequently, how much of the pharmacology of valproate is related to its effects on GHB is unknown.

Valproate might have an effect on gene regulation because when incubated with rat C6 glioma cells, it was found to increase the DNA binding activity of activator protein-1 (AP-1) transcription factors by up to two-fold. The effect was time- and concentration dependent (38). Similar results have been observed with human neuroblastoma cells (39). Glycogen synthase kinase can

phosphorylate the protein *c-jun*, an effect which can inhibit AP-1 DNA binding activity. Chen *et al.* (40) found that valproate can inhibit the kinase in a concentration-dependent manner, suggesting that the observed valproate-mediated increases in AP-1 binding to DNA are the results of a reduction in the amount of phosphorylated *c-jun*. Experiments using human neuroblastoma SH-SY5Y cells supported this concept (40).

Another possible genomic site of action of valproate might be the neuroprotective protein *bcl-2*. Chronic administration of valproate to rats led to a substantial increase in the number of *bcl-2* immunoreactive cells in layers 2 and 3 of the frontal cortex (41). *Bcl-2* seems to offer protection against conditions that induce neuronal degeneration (42, 43). An earlier study had demonstrated that valproate can protect cultured cerebellar neurons in culture from degeneration (44).

Valproate has been shown to alter β -adrenergic receptor function in rat C6 glioma cells. Chronic exposure reduced both the number of receptors and the capacity of cell membranes to bind [^3H]forskolin. In addition, the ability of forskolin to stimulate cAMP production was impaired (45).

2. Membrane Effects

Sodium Channels

There is evidence that valproate can interfere with both use-dependent and voltage-dependent Na^+ channels within the nervous system. Normally there is an increase in the rate of high frequency firing of action potentials during a train, but valproate can limit these increases. Moreover, the slow-down in the firing rate is further reduced by depolarizing from hyperpolarized potentials (46). The use- and voltage-dependency of the valproate effects are consistent with the pharmacology of the anticonvulsant which is known to inhibit repetitive discharges during convulsions but to have little effect on normal nerve cell activity.

Nosek (47) measured changes in action potentials in the crayfish stretch receptor in the presence of valproate. There was a reduction in axon excitability and a decrease in Na^+ and K^+ currents. Similar results were obtained when peripheral nerve fibers of *Xenopus* were used (48). Other workers have reported that valproate can significantly inhibit fast Na^+ currents (49, 50), although Taverna *et al.* (51) failed to observe an effect of valproate up to 200 μM on fast Na^+ currents in acutely dissociated neocortical neurons. However, low concentrations of valproate (10-30 μM) markedly reduced the persistent fraction of the Na^+ current.

Adrenal chromaffin cells, which are derived from neural crest, can be grown in culture. Bovine cells contain the α - and β_1 -subunit of voltage-dependent Na^+ channels, as well as the nicotine-ion channel complex and voltage-dependent

Ca²⁺ channels. Yamamoto *et al.* (52) exposed bovine chromaffin cells to valproate for up to six days. There was a time-dependent increase in the binding of saxitoxin, a site 1 ligand, which could be abolished by the presence of cycloheximide, a protein synthesis inhibitor. The presence of valproate also potentiated the veratridine-induced influx of Na⁺ and at the same time potentiated the veratridine-induced Ca²⁺ influx. The valproate treatment also increased the nicotine-induced Na⁺ influx through the ion channel of the nicotinic receptor. The mRNA for both the α - and the β -subunits of the Na⁺ channel was substantially increased by the valproate treatment. How can these increases in Na⁺ channel expression after repeated valproate exposure be explained in light of the evidence that acute administration leads to a reduction in Na⁺ influx? If there is a decrease in Na⁺ entry into the cell via voltage-sensitive channels, the cell might respond by synthesizing more of the Na⁺ channel protein, a process that could be considered an up-regulation. The reported decreases in use- and voltage-dependent Na⁺ currents induced by valproate (46) are unlikely to be a direct effect of the drug on the Na⁺-channel protein complex since valproate was unable to inhibit the binding of either [³H]saxitoxin or [³H]BTX-B (53, 54). These ligands recognize Na⁺-channel receptor sites 1 and 2, respectively. Accordingly, the most likely explanation is that valproate, a hydrophobic fatty acid, perturbs the plasma membrane adjacent to the channel, interfering with Na⁺ fluxes.

Potentiation of GABA Responses

In cultured spinal cord neurons GABA-mediated inhibitory responses can be measured, effects which were found by Macdonald and Bergey (55) to be potentiated by valproate in a dose-dependent manner (*Fig. 2*). The effects on GABA were specific since valproate had no influence on either glycine or glutamate responses. Further, valproate alone was found to exhibit no inhibitory action. Harrison and Simmonds (56), on the other hand, saw a potentiation of the responses to GABA only at very high concentrations when they used a rat cuneate afferent fiber preparation. Similar augmentation of GABA responses have been obtained in the intact central nervous system. For instance, after an intravenous injection of valproate or after iontophoretic application, the inhibitory effects of GABA were enhanced (57). Kerwin *et al.* (58) reported similar findings. Hayashi and Negishi (59) studied the effects of GABA and valproate on carp retinal ganglion cells and observed that both suppressed spike discharges, actions that could be antagonized by bicuculline. Further, the actions of valproate and GABA were additive. Similar results were obtained by Baldino and Geller (60) after they applied valproate and GABA simultaneously to cortical neurons and recorded greater inhibitory responses than with GABA alone. However, when these investigators used hypothalamic

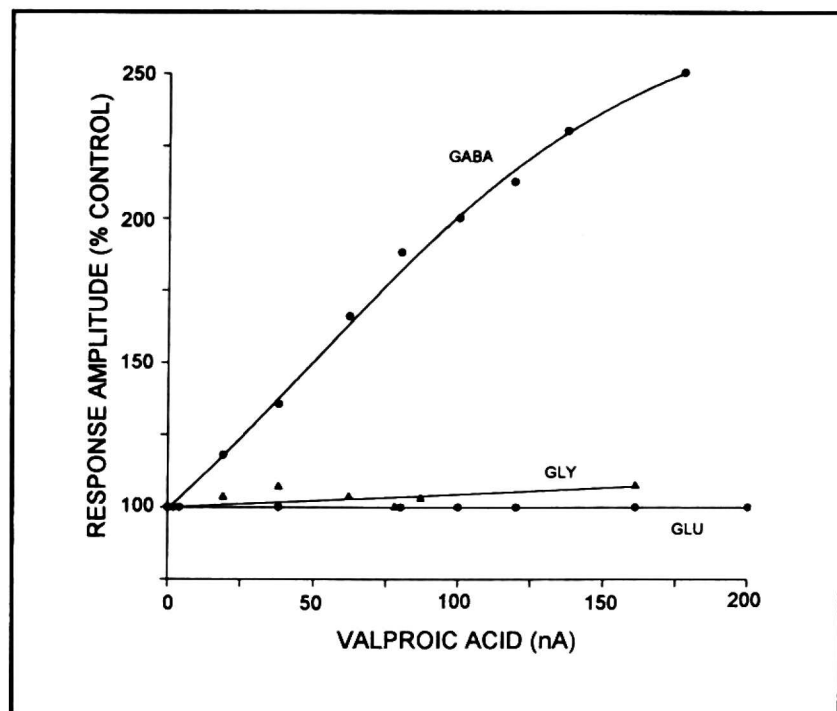
neurons, valproate exhibited inconsistent actions, leading to the suggestion that the effects of the drug are regionally specific within the brain (61). The inhibitory effects of both GABA and muscimol on locus coeruleus neurons were enhanced by valproate (62). Similarly in the preoptic area of the brain the inhibitory action of GABA was heightened by valproate (63). In addition, valproate increased the GABA-IPSP slope 54% in rat hippocampal slices, indicating that the mechanism of action of the anticonvulsant might include an augmentation of neuronal inhibition by GABA (64).

Fig. 2. Valproate selectively augmented GABA responses in cultured spinal cord neurons in a dose-dependent manner.

GLY = glycine;

GLU = glutamate.

Redrawn from Macdonald and Bergey (55).



In isolated frog primary afferent fibers valproate has been shown to increase the responsiveness to GABA and to reduce the ability of K^+ and excitatory amino acids to depolarize membranes (65). During *in vivo* experiments, Rowley *et al.* (66) observed that treatment of rats with valproate enhanced hippocampal GABA release.

Despite these reports indicating that valproate might be able to augment GABAergic mechanisms, experiments by Blume *et al.* (67) were unable to confirm this and, in fact, appeared to suggest that valproate can sometimes have excitatory actions on neurons. Extracellular recordings were made from cortical and hippocampal neurons in rats. Microiontophoretic application of valproate increased the spontaneous firing of the majority of neurons. GABA was able to inhibit the effects of valproate application.

Excitatory Amino Acid Receptors

One type of excitatory amino acid receptor in the brain is the N-methyl-D-aspartate (NMDA) receptor, the activation of which can lead to clonic convulsions (68). Antagonists at this receptor often possess

anticonvulsant properties (69). Czuczwar *et al.* (70) showed that valproate could suppress NMDA-induced seizures but that other anticonvulsants were ineffective. In rat cerebral cortical slices valproate inhibited the NMDA-stimulated release of [^3H]noradrenaline (71). NMDA can induce transient depolarizations in rat cortical pyramidal neurons which can be inhibited by valproate (72). Valproate also decreased the NMDA-epsp slope by 14% in rat hippocampal slices (64).

Another type of excitatory amino acid neurotransmitter receptor is the ionotropic AMPA receptor, whose activation leads to an influx of Na^+ ions resulting in a depolarization of the neuron. König *et al.* (73), using slices of human hippocampus incubated in a buffer containing 40 nM [^3H]AMPA, reported that concentrations of valproate of 100 μM or greater inhibited the binding of the ligand to AMPA receptors. These observations might have clinical significance since therapeutic serum levels of valproate range between 350 and 700 μM (5).

Although whole-brain levels of glutamate were unaffected by the acute administration of valproate (11, 34, 74), most brain regions showed increases in glutamate following chronic treatment (75, 13). Moreover, since aspartate might play a role in excitatory neurotransmission, the observation that valproate decreases mouse brain aspartate concentrations (76), an effect which correlates with suppression of audiogenic seizures, might contribute to an explanation of the mechanism of action of this drug. A later study found a good correlation between the dose of valproate and the reduction in brain aspartate levels (77, 78). Valproate also has been reported to (79) to inhibit the release of aspartate from rat cortical slices ($\text{IC}_{50} = 100 \mu\text{M}$).

Other Membrane Effects

Some of the effects of valproate on neuronal activity are not as clearly dependent on GABA or excitatory amino acid responses. Zhang *et al.* (80), for example, reported that the drug suppressed both simple and complex thalamocortical burst complex activity in brain slices. These activities resembled absence seizure activity and tonic-clonic activity, respectively. Low concentrations of valproate rapidly inhibited spontaneously firing cerebral cortical neurons following intraperitoneal injections (81); and Salt *et al.* (82) found that oral administration of valproate led to a reduction in the duration of afterdischarges in the rat amygdala. Another study showed that valproate could produce a modest reduction in T-type Ca^+ currents in primary afferent neurons over a concentration range 0.1—1 mM (83).

Hippocampal slices can be made to yield rhythmic, synchronous field discharges if incubated under certain conditions. Valproate reduced the occurrence of these discharges (84). Franceschetti *et al.* (85) also used

a hippocampal slice preparation and reported that valproate depressed frequency potentiation. Moreover, the drug inhibited spontaneous epileptiform activity and after discharge effected by antidromic stimulation. Because of the delayed action, these results suggest valproate does not block Na^+ currents, as phenytoin does, but perhaps activates Ca^+ -dependent K^+ flux.

Even in the invertebrate nervous system, valproate produced a hyperpolarization of *Aplysia* neurons which was accompanied by an increase in K^+ conductance (86).

Valproate has been shown to alter β -adrenergic receptor function in rat C6 glioma cells. Chronic exposure reduced both the number of receptors and the capacity of cell membranes to bind [^3H]forskolin. In addition, the ability of forskolin to stimulate cAMP production was impaired (87).

Antimigraine Activity

A major type of episodic headache is migraine. For many years the prevailing notion was that in patients undergo a migrainous attack certain cerebral blood vessels underwent dilation, resulting in the characteristic pain. However, investigators no longer believe that migraines are simply vascular headaches. The raphe nucleus of the brainstem seems to play a major role in the genesis of this type of headache. Pharmacological treatment of migraine attacks usually means treating the pain, either with analgesics or with drugs from the newer class of serotonin antagonists like sumatriptan. Yet agents which prevent the development of migraines are also important in the overall treatment available. One such drug is sodium valproate.

The first study to demonstrate that valproate might possess the ability to prophylactically treat migraine headaches was carried out by Sorensen (88). This was an open trial in which seventeen of eighteen patients obtained a benefit from the administration of valproate. The first study to employ a placebo was reported by Hering and Kuritzky (89) in which 86% of patients responded favorable to valproate treatment. In 1994 Jensen *et al.* (90) performed a triple-blind, placebo-controlled trial during which 34 patients completed the study. Seventeen patients showed a clear reduction in migraine frequency following valproate treatment. An open prospective investigation was carried out by Lenaerts *et al.* (91) in which 56 patients suffering from various types of migraine were enrolled. Over a period of six months, patients received an average of 928.5 mg sodium valproate each day which led to 80% of these patients experiencing a reduced frequency of migraine by at least one-half. A larger, multicenter study, involving 90 patients found that 48% achieved a reduction in headache frequency if valproate was administered on a daily basis (92). An even larger trial two years later involved 176 patients and confirmed the effectiveness of chronic valproate treatment on migraines (93).

The β -adrenergic receptor antagonist propranolol has been used successfully in the prophylactic treatment of migraine (94). Kaniecki (95) initiated a study to compare the relative effectiveness of valproate and propranolol as prophylactic agents in treating migraine. Thirty-two patients completed the randomized, single-blind, placebo-controlled trial which lasted 36 weeks. For valproate, most patients eventually received 1.5 g per day and for propranolol most patients eventually received 180 mg per day. Although migraine frequency was reduced in 19% of patients receiving placebo, 66% of patients receiving valproate and 63% of patients receiving propranolol reported a reduction in headache frequency. Thus valproate appears equally effective as propranolol at inhibiting the development of migraine headaches.

Valproate in Mood Disorders

Manic depression, or bipolar disorder, affects about one percent of the US population and these patients are at considerable risk for suicide. Lithium has been the drug of choice for the treatment of this disorder but serum levels have to be monitored carefully owing to its low therapeutic index. The relationship between the alleviation of bipolar symptoms and alterations in brain chemistry by lithium is far from clear. Further, many patients do not respond to lithium therapy (96), whereas others might show only partial longterm improvement (97).

A report by Lambert *et al.* (98) was one of the first to indicate valproate might be useful in treating mood disorders. Later, Emrich *et al.* (99) observed that out of five patients receiving valproate, four showed improvement in symptoms of acute mania. Since then much stronger evidence for the effective treatment of acute mania by valproate has been obtained by two clinical studies which properly employed placebo groups (94, 2, 100). The first was a randomized controlled trial in which 53% of a group of acutely manic patients experienced at least a 50% reduction in symptoms over the course of 3 weeks. Only 11% of the patients taking a placebo showed a similar improvement. The second of these randomized controlled trials yielded an improvement rate of 48% for patients on valproate compared to 25% of patients on placebo. Weaker evidence has been put forward for the alleviation of manic phase symptoms in bipolar disorder (101, 102). In a prospective clinical trial of patients with refractive bipolar disorder, valproate together with lithium was shown to have promise in treating the manic phase in one third of the eighteen patients studied (103). A subgroup of patients with bipolar disorder exhibit depression on occasion but these people are often unresponsive to lithium (104). In a parallel-group, double-blind study of acute mania certain patients who poorly responded to lithium treatment tended to exhibit symptoms of depression, whereas other patients showing signs of depression were better responders to valproate administration (105).

Recently valproate has begun to be used to treat both acute mania and manic depression in some patients. As valproate appears to enhance the GABAergic system and to interfere with Na^+ channel function, it is possible the mood stabilizing properties of the drug can be explained this way. However, glutamate function might be altered by valproate and account for its effectiveness in treating certain affective disorders. Interestingly, Dixon and Hokin (106) reported that both valproate and lithium stimulated glutamate release from cortical slices which was accompanied by an accumulation of inositol 1,4,5-triphosphate.

It has been estimated that up to 3% of the US population is susceptible panic attacks (107). Evidence points to the idea that susceptible patients might be helped with valproate treatment. In an open 7-week study by Primeau *et al.* (108), administration of valproate led to an improvement in a number of patients suffering from panic disorder. Keck *et al.* (109) reported that valproate is useful against lactate-induced panic attacks and against spontaneous attacks. Woodman and Noyes (110) have described a study involving twelve patients with panic disorder who were administered valproate over a 6-week period. Three-quarters of the patients exhibited a definite improvement in the frequency of attacks and in general anxiety levels. The remaining patients showed a moderate improvement. In a more recent trial, thirteen patients were enrolled but three eventually dropped out (111). Valproate treatment produced a significant improvement in depression, anxiety and mood instability as well as a marked reduction in the number of panic attacks.

Huntington's disease is a genetic neurological disorder in which neurons in the basal ganglia undergo a progressive degeneration. Since both GABA concentrations and the activity of glutamate decarboxylase were markedly reduced in postmortem brains from patients, a clinical trial was initiated to test the idea that valproate could alleviate neurological symptoms (112). The double-blind study involved eight patients who received placebo or 1.5 g valproate daily for one week followed by valproate and GABA (24.5 g) for a further 18 to 21 days. No improvement in chorea severity, finger dexterity, gait or speech was observed over the period of the trial.

TOXICOLOGY

Valproate is a widely used drug and is known to be reasonably safe and often effective. Nevertheless, sometimes hepatotoxicity can occur and there has been a number of fatalities, mainly at high doses (113, 114). Some of these patients exhibited microvesicular steatosis of the liver and encephalopathy. The basis of the fatalities might be explained by the effect of valproate on mitochondrial biochemical processes in hepatocytes.

In both humans and experimental animals treated with valproate, a dicarboxylic aciduria was observed, indicating an effect on mitochondrial β -oxidation of fatty acids (11). Since valproate is a branched chain fatty, it might be expected to interfere with normal metabolic oxidations of fatty acids in mitochondria. Also implicating mitochondria involvement was the presence of a persistent hyperammonemia during valproate intoxication (116—118). When isolated liver cells were studied, valproate clearly inhibited fatty acid oxidation (119, 120). Other studies using *in vitro* liver preparations were able to corroborate the inhibitory actions of valproate on β -oxidation (121—123). In addition to the effects on fatty acid oxidation, valproate inhibited fatty acid synthesis and gluconeogenesis (119, 124). Other researchers reported that valproate inhibited the urea cycle in hepatocytes (120, 125).

Some of the hepatotoxicity of valproate might be caused by metabolites of the drug. CoA esters of several valproate derivatives have been isolated (126, 127). In addition, a number of hydroxylation and dehydrogenation products have been described (128, 129) which could contribute to liver damage. Geber *et al.* (130) noticed a resemblance between the structure of 2-propylpent-4-enoate, a valproate metabolite, and the hepatotoxin methylene cyclopropylacetate.

Chronic treatment of rats with valproate led to decreased respiration in both liver and brain when either succinate or glutamate were substrates (131). Subsequent studies have shown that repeated valproate administration produces an inhibition of proton pumping ability by Complex IV of the inner membrane of liver mitochondria. This results in a reduction of oxidative phosphorylation (132).

CONCLUSIONS

Since it was shown to possess anticonvulsant properties almost forty years ago, valproate has become an important therapeutic agent to control several types of epilepsy. More recently the clinical uses of valproate have expanded to include its use in the treatment of migraine headaches and mood disorders. We are still not certain of its mechanism of action and, indeed, it is possible that its different clinical uses rely on different mechanisms. The two major targets for the action of valproate in the brain are the GABAergic neurotransmitter system and certain Na^+ channels involved in action potential propagation. It is obviously possible that the pharmacological profile of valproate depends on its action on both of these targets. At high doses valproate can result in hepatic damage. There is evidence that disturbances in mitochondrial metabolism within hepatocytes sometimes leads to irreversible alterations in liver function.

REFERENCES

1. Meunier G, Caraz G, Meunier Y, Eymard P, and Aimard M. Propriétés pharmacodynamiques de l'acide n-dipropylacétique. *Thérapie* 1963; 18: 435—438.
2. Rothrock JF. Clinical studies of valproate for migraine prophylaxis. *Cephalgia* 1997; 17: 81—83.
3. Pope HG, McElroy SL, and Keck PE. Valproate in the treatment of acute mania. *Arch Gen Psychiatr* 1991; 62—68.
4. Keck PE, Taylor VE, Tugrul KC, McElroy SK, Bennett JA. Valproate treatment of panic disorder and lactate-induced panic attacks. *Biol Psychiatr* 1993; 33: 542—546.
5. Chapman A, Keane PE, Meldrum BS, Simiand J and Verniers JC. Mechanism of anticonvulsant action of valproate. *Progr Neurobiol* 1982; 19: 315—359.
6. Fariello RG, Varasi M, Smith MC. Valproic acid: Mechanisms of action. In *Antiepileptic Drugs*, Levy RH, Mattson RH, Meldrum BS, (eds.). Raven Press, 1995, pp. 581—588.
7. Krnjevic K. Significance of GABA in brain function. In *GABA Mechanisms in Epilepsy*, Tunncliff G, Raess BU. (eds.). New York, Wiley-Liss, 1991, pp. 47—87.
8. Wood JD. The role of γ -aminobutyric acid in the mechanism of seizures. *Progr Neurobiol*, 1975; 5: 77—95.
9. Kash SF, Johnson RS, Tecott LH, Noebels JL, Mayfield RD, Hanahan D, and Baekkeskov S. Epilepsy in mice deficient in the 65-Kda isoform of glutamic acid decarboxylase. *Proc Natl Acad USA* 1997; 94: 14060—14065.
10. Tunncliff G, Raess BU. GABA neurotransmitter activity in human epileptogenic brain. In *GABA Mechanisms in Epilepsy*, Tunncliff G, Raess BU (eds.). New York, Wiley-Liss, 1991, pp. 105—119.
11. Godin Y, Heiner L, Mark J, Mandel P. Effects of di-n-propylacetate, an anticonvulsive compound, on GABA metabolism. *J Neurochem* 1969; 16: 869—873.
12. Simler S, Ciesielski L, Maitre M, Randrianarisoa H, Mandel P. Effect of sodium n-dipropylacetate on audiogenic seizures and brain γ -aminobutyric acid level. *Biochem Pharmacol* 1973; 1701—1708.
13. Patsalos PN, Lascelles PT. Changes in regional brain levels of amino acid putative neurotransmitters after prolonged treatment with the anticonvulsant drugs diphenylhydantoin, phenobarbitone, sodium valproate, ethosuximide and sulthiame in the rat. *J Neurochem* 1981; 36: 688—695.
14. Löscher W, Siemes H. Valproic acid increases gamma-aminobutyric acid in CSF of epileptic children. *Lancet* 1984; II: 225.
15. Fowler LJ, Beckford J, Johb RA. An analysis of the kinetics of the inhibition of rabbit brain γ -aminobutyrate aminotransferase by sodium-n-dipropylacetate and some other simple carboxylic acids. *Biochem Pharmacol* 1975; 24: 1267—1270.
16. Anlezark G, Horton RW, Meldrum BS, Sawaya MCB. Anticonvulsant action of ethanolamine-O-sulphate and di-n-propylacetate and the metabolism of γ -aminobutyric acid (GABA) in mice with audiogenic seizures. *Biochem Pharmacol* 1976; 25: 413—417.
17. Löscher W. *In vivo* administration of valproate reduces the nerve terminal (synaptosomal) activity of GABA aminotransferase in discrete brain areas of rats. *Neurosci Letts* 1993; 160: 177—180.
18. Larsson OM, Gram L, Schousboe I, Schousboe A. Differential effects of gamma-vinyl GABA and valproate on GABA-transaminase from cultured neurons and astrocytes. *Neuropharmacology* 1986; 25: 617—625.
19. Emson PC. Effects of chronic treatment with amino oxyacetic acid or sodium n-dipropylacetate on brain GABA levels and the development and regression of cobalt epileptic foci in rats. *J Neurochem* 1976; 27: 1498—1494.

20. Löscher W, Nau H. Valproic acid: metabolite concentrations in plasma and brain, anticonvulsant activity, and effects on GABA metabolism during subacute treatment in mice. *Arch Int Pharmacodyn Ther* 1982; 257: 20—31.
21. Nau H, Löscher W. Valproic acid: brain and plasma levels of the drug and its metabolites, anticonvulsant effects and γ -aminobutyric acid (GABA) metabolism in the mouse. *J Pharmacol Exp Ther* 1982; 220: 654—659.
22. Hearl HG, Churchich JE. Interactions between 4-aminotransferase and succinic semialdehyde dehydrogenase, two mitochondrial enzymes. *J Biol Chem* 1984; 259: 11459—11463.
23. Harvey PK, Bradford HF, Davison AN. The inhibitory effect of sodium n-dipropylacetate on the degradative enzymes of the GABA shunt. *FEBS Lett* 1975; 52: 251—254.
24. Van der Laan JW, De Boer T, Brunvels J. Di-n-propylacetate and GABA degradation: preferential inhibition of succinic semialdehyde dehydrogenase and indirect inhibition of GABA-transaminase. *J Neurochem* 1979; 32: 1769—1780.
25. Maitre M, Ossala L, Mandel P. *In vitro* studies into the effect of inhibition of rat brain succinic semialdehyde dehydrogenase on GABA synthesis and degradation. *FEBS Lett* 1976; 72: 53—57.
26. Pinder RM, Brogden RN, Speoght TM, Avery GS. Sodium valproate: a review of its pharmacological properties and therapeutic efficacy in epilepsy. *Drugs* 1977; 13: 81—123.
27. Vajda FJE, Donnan GA, Phillips T, Bladin PF. Human brain, plasma and cerebrospinal fluid concentration of sodium valproate after 72 hr of therapy. *Neurology* 1981; 31: 1364—1366.
28. Löscher W. Valproate induced changes in GABA metabolism at the subcellular level. *Biochem Pharmacol* 1981a; 30: 1364—1366.
29. Löscher W. Effect of inhibitors of GABA aminotransferase on the metabolism of GABA in brain tissue and synaptosomal fractions. *J Neurochem* 1981b; 36: 1521—1527.
30. Phillips NI, Fowler LJ. The effects of sodium valproate on γ -aminobutyrate metabolism and behaviour in naive and ethanolamine-O-sulphate pretreated rats and mice. *Biochem Pharmacol* 1982; 31: 2257—2261.
31. Löscher W. Valproate enhances GABA turnover in the substantia nigra. *Brain Res* 1987; 501: 198—203.
32. Taberner PV, Charington CB, Unwin JW. Effects of GAD and GABA-T inhibitors on GABA metabolism *in vivo*. *Brain Res Bull* 1980; 5: 621—625.
33. Cremer JE, Sarna GS, Teal HM, Cunningham VJ. Amino acid precursors: their transport into brain and initial metabolism. In *Amino Acids as Transmitters*, Fonnum F, (eds.) New York Plenum Press, 1978, pp. 669—689.
34. Chapman A, Riley K, Evans MC, Meldrum BS. Acute effects of sodium valproate and γ -vinyl GABA on regional amino acid metabolism in the rat brain: Incorporation of 2-(14 C)-glucose into amino acids. *Neurochem Res* 1982; 7: 1089—1105.
35. Tunnicliff G. Significance of γ -hydroxybutyric acid in the brain. *Gen Pharmacol* 1992; 23: 1027—1034.
36. Whittle SR, Turner AJ. Effects of the anticonvulsant sodium valproate on γ -aminobutyrate and aldehyde metabolism in ox brain. *J Neurochem* 1978; 31: 1453—1459.
37. Andriamampandry C, Siddert JC, Schmitt M, Garnier JM, Staub A, Muller C, Govaille S, Mark J, Maitre M. Cloning of a rat brain succinic semialdehyde reductase involved in the synthesis of the neuromodulator γ -hydroxybutyrate. *Biochem J* 1998; 334: 43—50.
38. Chen G, Yuan PX, Hawver DB, Potter WZ, Manji HK. Increase in AP-1 transcription factor DNA binding activity by valproic acid. *Neuropsychopharmacology* 1997; 16: 238—245.
39. Asghari V, Wang JF, Reisch JS, Young LT. Differential effects of mood stabilizers on Fas/Jun proteins and AP-1 DNA binding activity in human neuroblastoma SH—SY-5Y cells. *Mol Brain Res* 1998; 58: 95—102.

40. Chen G, Huang LD, Jiang YM, Manji HK. The mood-stabilizing agent valproate inhibits the activity of glycogen synthase kinase-3. *J Neurochem* 1999; 72: 1327—1330.
41. Chen G, Zen WZ, Yuang PX, Huang LD, Jiang YM, Zhao ZH, Manji HK. The mood-stabilizing agents lithium and valproate robustly increase the levels of the neuroprotective protein bcl-2 in the CNS. *J Neurochem* 1999; 72: 879—882.
42. Jacobson MD, Raff M. Programmed cell death and Bcl-2 protection in very low oxygen. *Nature* 1995; 374: 814—816.
43. Chen DF, Schmeider GE, Maetinou JC, Tonegawa S. Bcl-2 promotes regeneration of severed axons in mammalian CNS. *Nature* 1997; 385: 434—439.
44. Bruno V, Sortino MA, Scapagnini U, Nicoletti F, Canonico P. Antidegenerative effects of Mg²⁺-valproate in cultured cerebellar neurons. *Funct Neurol* 1995; 10: 121—130.
45. Chen G, Manji HK, Wright CB, Hawver DB, Potter WZ. Effects of valproic acid on β -adrenergic receptors, G-proteins, and adenylyl cyclase in rat C6 glioma cells. *Neuropsychopharmacology* 1996; 15: 271—280.
46. McLean MJ, Macdonald RL. Sodium valproate, but not ethosuximide, produces use- and voltage-dependent limitation of high frequency repetitive firing of action potentials of mouse central neurons in culture. *J Pharmacol Exp Ther* 1986; 237: 1001—1011.
47. Nosek TM. Depression of axonal excitability by valproate is antagonized by phenytoin. *Epilepsia* 1981; 22: 641—650.
48. Van Dongen AMJ, Van Erp MG, Voskuyl RA. Valproate reduces excitability by blockage of sodium and potassium conductance. *Epilepsia* 1986; 27: 177—182.
49. Zona C, Avoli M. Effects induced by the antiepileptic drug valproic acid upon the ionic currents recorded in rat neocortical neurons in cell culture. *Exp Brain Res* 1990; 81: 313—317.
50. Van der Berg RJ, Kok P, Voskuyl RA. Valproate and sodium currents in cultured hippocampal neurons. *Exp Brain Res* 1993; 93: 279—287.
51. Taverna S, Mantegazza M, Franceschetti S, Avanzini G. Valproate selectively reduces the persistent fraction of Na⁺ current in neocortical neurons. *Epilepsy Res* 1998; 32: 304—308.
52. Yamamoto R, Yanagita T, Kobayashi H, Yokoo H, Wada A. Up-regulation of sodium channel subunit mRNAs and their cell surface expression by antiepileptic valproic acid: Activation of calcium channel and catecholamine secretion in adrenal chromaffin cells. *J Neurochem* 1997; 68: 1655—1662.
53. Willow M, Catterall WA. Inhibition of binding of [³H]batrachotoxinin A 20- α benzoate to sodium channels by the anticonvulsant drugs diphenylhydantoin and carbamazepine. *Mol Pharmacol* 1982; 22: 627—635.
54. Willow M, Kuenzel EA, Catterall WA. Inhibition of voltage-sensitive sodium channels in neuroblastoma cells and synaptosomes by the anticonvulsant drugs diphenylhydantoin and carbamazepine. *Mol Pharmacol* 1984; 25: 228—234.
55. Macdonald RL, Bergey GK. Valproic acid augments GABA-mediated postsynaptic inhibition in cultured mammalian neurones. *Brain Res* 1979; 170: 558—562.
56. Harrison NL, Simmonds MA. Sodium valproate enhances responses to GABA receptor activation only at high concentrations. *Brain Res* 1982; 250: 201—204.
57. Gent JP, Phillips NI. Sodium di-*n*-propylacetate (valproate) potentiates responses to GABA and muscimol in single central neurones. *Brain Res* 1980; 197: 275—278.
58. Kerwin RW, Olpe HR, Schmutz M. The effect of sodium-*n*-dipropylacetate on γ -aminobutyric acid-dependent inhibition in the rat cortex and substantia nigra in relation to its anticonvulsant activity. *Br J Pharmacol* 1980; 71: 545—551.
59. Hayashi T, Negishi K. Suppression of retinal spike discharge by dipropylacetate (Depakene): a possible involvement of GABA. *Brain Res* 1979; 175: 271—278.

60. Baldino F, Geller HM. Sodium valproate enhancement of gamma-aminobutyric acid (GABA) inhibition: electrophysiological evidence for anticonvulsant activity. *J Pharmacol Exp Ther* 1981; 217: 445—450.
61. Baldino F, Geller HM. Effect of sodium valproate on hypothalamic neurons in vivo and in vitro. *Brain Res* 1981; 219: 231—237.
62. Olpe HR, Steinmann MW, Pozza MF, Brugger F, Schutz M. Valproate enhances GABA_A-mediated inhibition of locus coeruleus neurons in vitro. *Naunyn-Schmiedeberg's Arch Pharmacol* 1988; 338: 655—657.
63. Wolf R, Tscherne U, Emrich HM. Suppression of preoptic GABA release caused by push-pull-perfusion with sodium valproate. *Naunyn-Schmiedeberg's Arch Pharmacol* 1988; 338: 658—663.
64. Ko GY, Brown-Croyts LM, Teyler TJ. The effects of anticonvulsant drugs on NMDA-EPSP, AMPA-EPSP and GABA-IPSP in the rat hippocampus. *Brain Res Bull* 1997; 42: 297—302.
65. Hackmsn JC, Grayson V, Davidoff RA. The presynaptic effects of valproic acid in the isolated frog spinal cord. *Brain Res* 1981; 220: 269—285.
66. Rowley HL, Marsden CA, Martin KF. Differential effects of phenytoin and sodium valproate on seizure-induced changes in γ -aminobutyric acid and glutamate release in vivo. *Eur J Pharmacol* 1995; 294: 541—546.
67. Blume HW, Lamour Y, Arnauld E, Layton BS, Renaud LP. Sodium di-n-propylacetate (valproate) action on single neurons in rat cerebral cortex and hippocampus. *Brain Res* 1979; 171: 182—185.
68. Czuczwar SJ, Meldrum B. Protection against chemically induced seizures by 2-amino-7-phosphoheptanoic acid. *Eur J Pharmacol* 1982; 83: 335—3.
69. De Sarro GB, De Sarro A. Anticonvulsant properties of non-competitive antagonists of the N-methyl-D-aspartate receptor in genetically epilepsy-prone rats: comparison with CPPene. *Neuropharmacology* 1993; 32: 51—58.
70. Czuczwar SJ, Frey HH, Loscher W. Antagonism of N-methyl-D,L-aspartic acid-induced convulsions by antiepileptic drugs and other agents. *Eur J Pharmacol* 1985; 108: 273—2.
71. Brown LM, Lee YP, Teyler TJ. Antiepileptics inhibit cortical N-methyl-D-aspartate-evoked [³H]norepinephrine efflux. *Eur J Pharmacology* 1994; 254: 307—309.
72. Zeise ML, Kasparow S, Zieglgansverger W. Valproate suppresses N-methyl-D-aspartate-evoked, transient depolarizations in the rat neocortex in vitro. *Brain Res* 1991; 544: 345—348.
73. König G, Niedermeyer B, Deckert J, Gsell W, Ransmayr G, Riederer P. [³H] α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid [AMPA] binding by the anticonvulsant valproate in clinically relevant concentrations: an autoradiographic investigation in human hippocampus. *Epilepsy Res* 1998; 31: 153—157.
74. Kukino K, Deguchi T. Effects of sodium dipropylacetate on γ -aminobutyric acid and biogenic amines in rat brain. *Chem Pharm Bull* 1977; 25: 2257—2262.
75. Williams RJH, Patsalos PN, Lowe R, Lascelles PT. Changes in brain amino acid neurotransmitters induced by sodium valproate and their relevance to epilepsy and interactions with other anticonvulsants. In *The Place of Sodium Valproate in the Treatment of Epilepsy* London Academic Press, 1980, pp. 95—102.
76. Schechter PJ, Tranier Y, Grove J. Effect of n-dipropylacetate on amino acid concentrations in mouse brain: correlation with anti-convulsant activity. *J Neurochem* 1978; 31: 1325—1327.
77. Chapman AG, Croucher MJ, Meldrum BS. Anticonvulsant activity of intracerebroventricularly administered valproate and valproate analogues. A dose-dependent correlation with changes in brain aspartate and GABA levels in DBA/2 mice. *Biochem Pharmacol* 1984; 33: 1459—1463.
78. Perry T, Hansen S. Biochemical effects in man and rat of three drugs which can increase brain GABA content. *J Neurochem* 1978; 30: 679—684.

79. Crowder JM, Bradford HF. Common anticonvulsants inhibit Ca^{2+} uptake and amino acid neurotransmitter release in vitro. *Epilepsia* 1987; 28: 378—382.
80. Zhang YF, Gibbs JW, Coulter DA. Anticonvulsant drug effects on spontaneous thalamocortical rhythms *in vitro*: valproic acid, clonazepam and alpha-methyl-alpha-phenylsuccinimide. *Epilepsy Res* 1996; 23: 37—53.
81. Schmutz M, Olpe HR, Koella WP. Central actions of valproate sodium. *J Pharm Pharmacol* 1979; 31: 1325—1327.
82. Salt TE, Tulloch IF, Walrwe DS. Anti-epileptic properties of sodium valproate in rat amygdaloid kindling. *Br J Pharmacol* 1980; 68: 34P.
83. Kelly KM, Gross RA, Macdonald RL. Valproic acid selectively reduces the low-threshold (T) calcium current in rat nodose neurons. *Neurosci Lett* 1990; 116: 233—238.
84. Agopyan N, Avoli H, Rieb L, Tancredi V. Depression of hippocampal low calcium field bursts by the antiepileptic drug valproic acid. *Neurosci Lett* 1985; 60: 57—62.
85. Franceschetti S, Hamon B, Heineman U. The action of valproate on spontaneous epileptiform activity in the absence of synaptic transmission and on evoked changes in $[\text{Ca}^{2+}]$ and $[\text{K}^{+}]$ in the hippocampal slice. *Brain Res* 1986; 386: 1—11.
86. Slater GE, Johnston GD. Sodium valproate increases potassium conductance in *Aplysia* neurones. *Epilepsia* 1978; 19: 379—384.
87. Chen G, Manji HK, Wright CB, Hawver DB, Potter WZ. Effects of valproic acid on beta-adrenergic receptors, G-proteins, and adenylyl cyclase in rat C6 glioma cells. *Neuropsychopharmacology* 1996; 15: 271—280.
88. Sorensen KV. Valproate: a new drug in migraine prophylaxis. *Acta Neurol Scand* 1988; 78: 346—348.
89. Hering R, Kuritzky A. Sodium valproate in the prophylactic treatment of migraine: a double-blind study versus placebo. *Cephalalgia* 12: 81—84.
90. Jensen R, Brinck T, Olesen J. Sodium valproate has a prophylactic effect in migraine without aura: a triple-blind, placebo-controlled crossover study. *Neurology* 1994; 44: 647—651.
91. Lenaerts M, Bastings E, Sianard J, Schoenen J. Sodium valproate in severe migraine and tension-type headache: an open study of long-term efficacy and correlation with blood levels. *Acta Neurol Belg* 1996; 96: 126—129.
92. Mathew N, Saper J, Silberstein S, Rankin L, Markley HG, Solomon S *et al*. Migraine prophylaxis with divalproex. *Arch Neurol* 1995; 52: 281—286.
93. Klapper JA. Divalproex sodium in migraine prophylaxis study group: A dose -controlled study. *Cephalalgia* 1997; 17: 103—108.
94. Weber RB, Reinmuth OM. The treatment of migraine with propranolol. *Neurology* 1972; 22: 336—369.
95. Kaniecki R. A comparison of divalproex with propranolol and placebo for the prophylaxis of migraine without aura. *Arch Neurol* 1997; 54: 1141—1145.
96. Harrow M, Goldberg JF, Grossman LS, Meltzer HY. Outcome in manic disorders. *Arch Gen Psychiatry* 1990; 47: 665—671.
97. Maj M, Priozi R, Kemali D. Long-term outcome of lithium prophylaxis in patients initially classified as complete responders. *Psychopharmacology* 1989; 98: 535—538.
98. Lambert PA, Cavaz G, Borselli S, Carrel S. Action neuropsychotrope d'un nouvel antiépileptique: Le Dépamide. *Ann Med Psychol* 1966; 1: 707—710.
99. Emrich HM, Von Zerssen D, Kissling W, Moller HJ, Windorfer A. Effect of sodium valproate in mania. The GABA hypothesis of affective disorders. *Arch Psychiatr Nervenkr* 1980; 229: 1—16.
100. Bowden C, Brugger AM, Swann AC, Calabrese JR, Janicak PG, Petty F *et al*. Efficacy of divalproex vs. lithium and placebo in the treatment of mania. *J Am Med Assoc* 1994; 271: 918—924.

101. Calabrese JR, Delucchi GA. Spectrum of efficacy of valproate in 55 patients with rapid-cycling bipolar disorder. *Am J Psychiat* 1990; 147: 431—434.
102. Lambert PA, Venaud G. Comparative study of valpromide versus lithium as prophylactic treatment in affective disorders. *Nervure J Psychiat* 1995; 7: 1—9.
103. Denicoff KD, Smith-Jackson EE, Bryan AL, Ali SO, Post RM. Valproate prophylaxis in a prospective clinical trial of refractory bipolar disorder. *Am J Psychiatry* 1997; 154: 1456—1458.
104. Prien RF, Himmelhoch JM, Kupfer DJ. Treatment of mixed mania. *J Affect Disord* 1988; 15: 9—15.
105. Swann AC, Bowden CL, Morris D, Calabrese JR, Petty F, Small J *et al.* Depression during mania: Treatment response to lithium or divalproex. *Arch Gen Psychiatry* 1997; 54: 37—42.
106. Dixon JF, Hokin LE. The antibipolar drug valproate mimics lithium in stimulating glutamate release and inositol 1,4,5-triphosphate accumulation in brain cortex slices but not accumulation of inositol monophosphates and biphosphates. *Proc Natl Acad Sci USA* 1997; 94: 4757—4760.
107. Kessler RC, McGonagle KA, Zhao S, Nelson CB, Hughes M, Eshelman S, *et al.* Lifetime and 12 month prevalence of DSM-III-R psychiatric disorders in the United States. Results from the National Comorbidity Survey. *Arch Gen Psychiatr* 1994; 51: 8—19.
108. Primeau F, Fontaine R, Beauclair L. Valproic acid and panic disorder. *Can J OPsychiatr* 1990; 35: 248—250.
109. Keck PE, Taylor VE, Tugrul KC, McElroy SL, Bennett JA. Valproate treatment of panic disorder and lactate-induced panic attacks. *Biol Psychiatr* 1993; 33: 542—546.
110. Woodman C, Noyes R. Panic Disorder: treatment with valproate. *J Clin Psychiatr* 1994; 55: 134—136.
111. Baetz M, Bowen RC. Efficacy of divalproex sodium in patients with panic disorder and mood instability who have not responded to conventional therapy. *Can J Psychiatr* 1998; 43: 73—77.
112. Shoulson I, Kartzinel R, Chase TN. Huntingtons's disease: Treatment with dipropylacetic acid and gamma- aminobutyric acid. *Neurology* 1976; 26: 61—63.
113. Dreifuss FE, Santili N, Langer DH, Sweeney KP, Moline KA, Menander KB. Valproic acid hepatic fatalities: A retrospective review. *Neurology* 1987; 37: 379—385.
114. Zimmerman HJ, Ishak KG. Valproate-induced hepatic injury: Analyses of 23 fatal cases. *Hepatology* 1982; 2: 591—597.
115. Mortensen PB, Gregersen N, Kolvraa S, Christensen E. The occurrence of C6—C10-dicarboxylic acids in urine from patients and rats treated with dipropylacetate. *Biochem Med* 1980; 24: 153—161.
116. Coulter DL, Allen RJ. Secondary hyperammonaemia: A possible mechanism for valproate encephalopathy. *Lancet* 1980; 1: 1310—1311.
117. Rawat S, Borkowski Wj, Swick HM. Valproic acid and secondary hyperammonemia. *Neurology* 1981; 31: 1173—1174.
118. Zaccara G, Campostrini R, Paganini M, Moroni F, Valenza T, Targioni G, *et al.* Acute changes of blood ammonia may predict short-term adverse effects of valproic acid. *Neurology* 1984; 34: 1519—1521.
119. Becker CM, Harris RA. Influence of valproic acid on hepatic carbohydrate and lipid metabolism. *Arch Biochem Biophys* 1983; 223: 381—392.
120. Turnbull DM, Bone AJ, Bartlett K, Koundakjian PP, Sherratt HSA. The effects of valproate on intermediary metabolism in isolated rat hepatocytes and intact rats. *Biochem Pharmacol* 1983; 32: 1887—1892.
121. Bjorge SM, Baille TA. Inhibition of medium-chain fatty acid β -oxidation *in vitro* by valproic acid and its unsaturated metabolite, 2-*n*-propyl-4-pentenoic acid. *Biochem Biophys Res Commun* 1985; 132: 245—252.

122. Draye JP, Vamecq J. The inhibition by valproic acid of the mitochondrial oxidation of monocarboxylic and ω -hydroxymonocarboxylic acids: Possible implications for the metabolism of gamma-aminobutyric acid. *J Biochem* 1987; 102: 235—242.
123. Veitch K, Van Hoof F. *In Vitro* effects of eight-carbon fatty acids on oxidations in rat liver mitochondria. *Biochem Pharmacol* 1990; 40: 2153—2159.
124. Rogies V, Vandenberghe Y, Vercruysse A. Inhibition of gluconeogenesis by sodium valproate and its metabolites in isolated rat hepatocytes. *Xenobiotica* 1985; 15: 759—765.
125. Cloudé FX, Grimber G, Parvy P, Rabier D, Petit F. Inhibition of ureagenesis by valproate in rat hepatocytes. *Biochem J* 1983; 216: 233—236.
126. Li J, Norwood DL, Mao LF, Schultz H. Mitochondrial metabolism of valproic acid. *Biochemistry* 1991; 30: 388—394.
127. Bjorge SM, Baille TA. Studies of the β -oxidation of valproic acid in rat liver mitochondrial preparations. *Drug Metab Dispos* 1991; 19: 823—829.
128. Kesterson JW, Granneman GR, Machinist JM. The hepatotoxicity of valproic acid and its metabolites in rats. I. Toxicologic, biochemical and histopathologic studies. *Hepatology* 1984; 4: 1143—1152.
129. Granneman GR, Wang SI, Kesterson JW, Machinist JM. The hepatotoxicity of valproic acid and its metabolites in rats. II. Intermediary and valproic acid metabolism. *Hepatology* 1984; 4: 1153—1158.
130. Geber N, Dickinson RG, Hartland RC, Lynn RK, Houghton D, Antonias JI. *et al.* Reye-like syndrome associated with valproic acid therapy. *J Pediatr* 1979; 95: 142—144.
131. Rumbach L, Warter Jm, Rendon A, Marescaux C, Micheletti G, Waksman A. Inhibition of oxidative phosphorylation in hepatic and cerebral mitochondria of sodium valproate-treated rats. *J Neurol* 1983; 61: 417—423.
132. Ponchaut S, Van Hoof F, Veitch K. Cytochrome aa_3 depletion is the cause of the deficient intramitochondrial respiration induced by chronic valproate administration. *Biochem Pharmacol* 1992; 43: 644—647.

Received: April 30, 1999

Accepted: June 30, 1999

Author's address: G. Tunncliff, Department of Biochemistry and Molecular Biology, Indiana University School of Medicine, 8600 University Boulevard, Evansville, IN 47712, USA.