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# INITIATING BIODEGRADATION OF POLYVINYLPYRROLIDONE IN AN AQUEOUS AEROBIC ENVIRONMENT

# ZAINICJOWANIE BIODEGRADACJI POLIWINYLOPIROLIDONU W ŚRODOWISKU WODNO-POWIETRZNYM

Abstract: A synthetic polymer, Polyvinylpyrrolidone (PVP - E 1201) primarily finds applications in the pharmaceutical and food industries due to its resistance and zero toxicity to organisms. After ingestion, the substance passes through the organism unchanged. Consequently, it enters the systems of municipal wastewater treatment plants (WWTP) without decomposing biologically during the waste treatment process, nor does it attach (through sorption) to particles of activated sludge to any significant extent, therefore, it passes through the system of a WWTP, which may cause the substance to accumulate in the natural environment. For this reason the paper investigates the potential to initiate aerobic biodegradation of PVP in the presence of activated sludge from a municipal wastewater treatment plant. The following agents were selected as the initiators of the biodegradation process - co-substrates: acrylamide, N-acethylphenylalanine and 1-methyl-2-pyrrolidone, a substance with a similar structure to PVP monomer. The biodegradability of PVP in the presence of co-substrates was evaluated on the basis of biological oxygen demand (BOD) as determined via a MicroOxymax O2/CO2/CH4 respirometer. The total substrate concentration in the suspension equaled 400 mg  $dm^{-3}$ , with the ratio between PVP and the co-substrate being 1:1, while the concentration of the dry activated sludge was 500 mg dm<sup>-3</sup>. Even though there was no occurrence of a significant increase in the biodegradation of PVP alone in the presence of a co-substrate, acrylamide appeared to be the most effective type of co-substrate. Nevertheless, a recorded decrease in the slope of biodegradation curves over time may indicate that a process of primary decomposition was underway, which involves the production of metabolites that inhibit activated sludge microorganisms. The resulting products are not identified at this stage of experimentation.

Keywords: polyvinylpyrrolidone, biodegradation, activated sludge, aqueous environment

*Polyvinylpyrrolidone* (PVP) is a synthetic polymer that dissolves very well in water. From a chemical perspective, it is a polymer lactam with an internal amide bond and ranks amongst poly-*N*-vinylamides (Fig. 1). The diverse forms produced commercially are differentiated through the K-number denoted (eg PVP K12, K17, K25 and K30), indicating average molecular weight. With regard to monomer unit structure, PVP exhibits an amphoteric characteristic, due to it containing an amide group of high polarity and non-polar methylene groups in its main chain and ring, which indicates its hydrophilic and polar properties and hydrophobic properties, respectively.

Due to its physical, chemical and biological properties (bio- and hemocompatible, physiologically inactive), PVP finds applications in a range of technological processes. Due to its resistance and zero toxicity to organisms, the compound is mainly used in the pharmaceutical and food industries (in the European Union soluble PVP is labeled E 1201,

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within the system of E-numbers, when utilized in food supplements, pills and sweeteners). After ingestion, the substance passes through the organism unchanged [1]; consequently, it enters the systems of municipal *wastewater treatment plants* (WWTP) without decomposing biologically during the waste treatment process, according to most recent studies [2-4]; nor does it attach (through sorption) to particles of activated sludge to any significant extent [5]. Therefore, it passes through the system of a WWTP, which may cause the substance to accumulate in the natural environment.

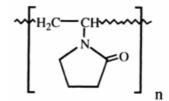


Fig. 1. The chemical structure of polyvinylpyrrolidone

However, some studies do not completely reject its biodegradability, pointing out the importance of experimental conditions and the microbial cultures used. Abd El-Mohdy and Ghan [6], describe biodegradation of PVP in a soil environment (soil burial test). Their findings of a decrease in mass does not evidence the complete resilience of pure PVP, as regards decomposition, with such mass dropping to about 10% after 6 months. Marusincova [7] also notes some positive findings on eliminating PVP (Mr ~ 10 kDa) from a municipal WWTP by using anaerobic sludge; increased production of  $CH_4$  was recorded for samples containing PVP after 24 days of testing compared with the endogenous production of  $CH_4$ .

As mentioned in the introduction, PVP contains a lactam ring in its structure, this being  $\gamma$ -lactam, a substance that may be subject to attack by  $\gamma$ -lactamase (an enzyme). Therefore, from both a theoretical and practical perspective, it would be of interest to use microorganisms producing the aforementioned  $\gamma$ -lactamase as a cornerstone for biologically decomposing PVP. Indeed, a number of microorganisms have been identified very recently, such as *Pseudomonas fluorescens* [8], *Microbacterium hydrocarbonoxydans* [9], *Sulfolobus solfataricus MT4*, *Rhodococcus* sp. *ENZA1*, *Aureobacterium sp. ENZA25*, *Pseudomonas solanacearum ENZA20*, *P. cepaecia* and *P. fluorescens ENZA22*, and *Commonas acidovorans* [10]. Among other things, the work of Line et al [11] also found that some soil bacteria producing  $\gamma$ -lactamase are able to utilize *N*-acyl as a sole source of carbon and energy [11], while the study of Hickey et al [12] revealed increased activity by  $\gamma$ -lactamase in the presence of acrylamide substrate. This theoretically suggests that *N*-acethylphenylalanine or acrylamide might initiate the production of  $\gamma$ -lactamase in some types of microorganisms.

The literature study shows that the ultimate fate of PVP within a WWTP and subsequently in the environment has not been widely explored, so reiterating presumptions that PVP does not pose an environmental problem is not really an option. This paper investigates the potential to initiate aerobic biodegradation of PVP in the presence of mixed microbial cultures, in the form of activated sludge from a municipal wastewater treatment plant, with the following agents selected as the initiators of the biodegradation process - co-substrates: 1-methyl-2-pyrrolidone, a substance with a similar structure to PVP monomer, as well as acrylamide [12] with *N*-acethylphenylalanine [11], this based on literature research.

### Materials and methods

The materials used for tests were commercial products that are currently available: polyvinylpyrrolidone K15 (Mr ~ 10 kDa), the Aldrich Chemical Company; 1-methyl-2-pyrrolidone (1M-2P), acrylamide (AC) and N-acethylphenylalanine (APhA), at a purity of 98%, the Aldrich Chemical Company. Others chemicals employed were of analytical purity, produced or delivered by the Lachema Co., Brno, the Czech Republic.

A mixed microbial culture in the form of activated sludge from a municipal *wastewater treatment plant* (WWTP Zlin-Malenovice, the Czech Republic) was applied as the inoculum (biological material). Prior to such testing, the aerobic sludge was first centrifuged at 4,500 rpm<sup>-1</sup> for 10 minutes at 20°C. After removing the supernatant, the biomass was suspended in the biomedium [13] and then aerated for 24 hours.

Biodegradability was determined using a respirometer, a MicroOxymax  $O_2/CO_2/CH_4$ . The sample tested (PVP or PVP + co-substrate) was the sole source of organic carbon and energy in the environment. The conditions of the test were as follows: suspension volume 50 mL, dried sludge 500 mg·dm<sup>-3</sup>, concentration of PVP 200 mg·dm<sup>-3</sup>, concentration of the co-substrate 200 mg·dm<sup>-3</sup>, temperature  $25\pm1^{\circ}C$  and pH = 7. At the beginning and end of the experiment, sludge solids, pH and reference DOC (*Dissolved Organic Carbon*) were determined for all samples. Allylthiourea as an inhibitor of nitrifying processes was not dosed when studying the biodegradation of PVP alone, even though the substance studied contains organically bound nitrogen. This procedure was chosen with regard to the work of Marusincova [7], where aerobic biodegradation of PVP *via* ordinary wastewater treatment plant culture was not underway and, furthermore, the addition of allylthiourea was causing the process to become inhibited. All measurements were taken three times in parallel.

The course of biodegradation was evaluated on the basis of determining *Biochemical Oxygen Demand* (BOD) [13], expressed as the ratio of the biological oxygen demand measured vs *Theoretical Oxygen Demand* - BOD/TOD [%], according to equation:

Biodegradation of PVP = 
$$\frac{\left(\frac{BOD_{PVP/COSUBSTRATE} - BOD_{COSUBSTRATE}}{c_{PVP}}\right)}{TOD_{PVP}} \times 100$$
(1)

where BOD<sub>PVP/COSUBSTRATE</sub> and BOD<sub>PVP/COSUBSTRATE</sub> are experimentally found values pertaining to biological oxygen demand of PVP biodegradation in the presence of the co-substrate, respectively merely the co-substrate, (all in [mg·dm<sup>-3</sup>]),TOD is theoretical oxygen demand (for PVP = 2020 mg·g<sup>-1</sup>), and c<sub>PVP</sub> is the concentration of PVP in the suspension tested in [g·dm<sup>-3</sup>].

## **Results and discussion**

The first part of the work focused on observing the biodegradability of PVP and each of the co-substrates. The results of these measurements were then used to evaluate the

impact of the co-substrates on the biodegradation of PVP. Due to the fact that the retention time of wastewater in WWTP aeration tanks is in the order of several hours (depending on the type of WWTP facility), testing was carried out only for 30 days.

The BOD<sub>max</sub>/TOD values for each co-substrate (data not presented) revealed that the substances are highly biologically degradable. Acrylamide decomposed from 100% within as little as 170 hours of testing, the lag phase being 60 hours. Similarly, 1M-2P was fully decomposed after 300 hours, with the lag phase equaling 30 hours. As regards APhA, 83% biodegradation was achieved after approximately 200 hours, with the lag phase being 30 hours. BOD attributable to the decomposition of PVP alone, without any additional co-substrate, progressed almost at the level of endogenous respiration. Despite the above, about 3.19  $\pm$  2.91% of biodegradation was achieved. Nonetheless, this value cannot be considered significant due to its high standard deviation, and it may be assumed that under the given conditions decomposition occurs only through the production of unreacted *N*-vinylpyrrolidone, a monomer contained in commercial PVP.

Figures 2 and 3 show the impact of the co-substrate on the biodegradation of PVP alone. It should be emphasized that each curve (Fig. 2) and column (Fig. 3), as shown in the diagrams, are based on the BOD value for merely PVP, ie after deducting oxygen consumption attributable to the decomposition of the co-substrate itself and endogenous respiration of activated sludge.

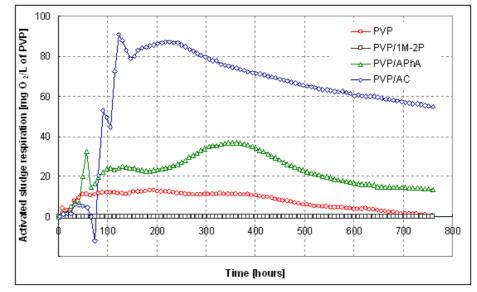


Fig. 2. The course of PVP biodegradation in the presence of co-substrates expressed as the respiration of activated sludge per instance of decomposition of PVP alone; that is after deduction of endogenous respiration and respiration attributable to the decomposition of merely the co-substrate

The most significant influence on the degradation of PVP was that of acrylamide. The biodegradation curve showing the combination of PVP and acrylamide was located above the curve of merely the co-substrate. The difference between both curves pertains to the

biodegradation of PVP alone (about 20% of biodegradation). Nevertheless, the sample containing PVP + the co-substrate showed a stagnation of BOD values over time compared with endogenous respiration, which resulted in a drop in the biodegradation curves (Fig. 2). The results displayed in Figure 2 thus indicate that there might be primary decomposition of PVP accompanied by the production of metabolites inhibiting microorganisms in the activated sludge.

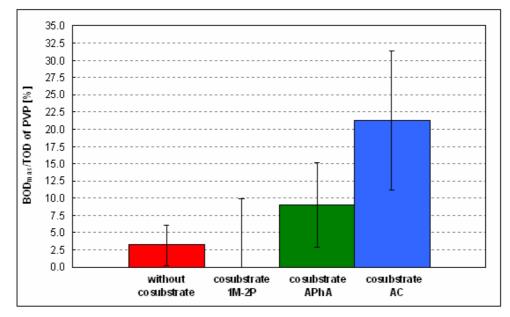


Fig. 3. The percentage of PVP biodegradation expressed as BOD<sub>max</sub>/TOD; ± standard deviation

Adding APhA to the PVP sample did not induce the presumed support of decomposition of the latter (Fig. 2). The values of biological oxygen demand attributable to the decomposition of PVP in the presence of APhA varied only slightly to those of BOD attributable to the decomposition of the co-substrate alone. As with acrylamide, there was also a reduction in the biodegradation curves over time in this case.

Although 1M-2P is a substance with a structure greatly resembling that of PVP monomer, there was clearly no formation of  $\gamma$ -lactamase or other enzymes capable of decomposing PVP throughout the biodegradation process. Concurrently, adding 1M-2P suppressed the decomposition of merely PVP (Fig. 2).

The results imply (Fig. 3) that the aerobic environment is probably one in which partial PVP lactam ring cleavage has the potential to occur through the activities of microorganisms and influence of the AC and APhA co-substrates, and which may not be accompanied by polymer chain decomposition; it was not studied whether this is caused by  $\gamma$ -lactamase production or not. The declining trend of the curves indicates the aforementioned production of metabolites inhibiting microorganisms in the activated sludge. Any resulting intermediates were not identified at this stage of experimentation.

## Conclusions

- The results of this preliminary research need to be viewed with reservation (due to the high values of standard deviations). However, in light of the knowledge obtained, it can be concluded that the biodegradation of PVP is influenced by the presence of other substances. Toxic metabolites may form during the PVP decomposition process. This fact will require verification in future research using advanced instrumental techniques (GC or HPLC with MS detection). At the same time it would be appropriate to extend the testing period.
- None of the selected co-substrates proved a significant accelerator of PVP biodegradation. The efficacy per co-substrate may be arranged as follows: AC > APhA > 1M-2P, with an astonishing 20% of biodegradation achieved in the case of acrylamide.

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