

# Synthesis, physicochemical studies, fluorescence behavior, and anticancer properties of transition metal complexes with the pyridyl ligand

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A novel series of complexes with the formula  $[MLCl]$   $[M = Co(II) (1), Ni(II) (2), Cu(II) (3), Zn(II) (4)]$  arising from Pyridyl ligand,  $N,N'$ -bis(1-(2-pyridyl)ethylidene)-2,2-dimethylpropane-1,3-diamine), ligand, L, was synthesized and investigated by elemental analyses, FT-IR,  $^1H$  and  $^{13}C$  NMR, Powder XRD, and thermal analyses. TGA analysis indicated that all complexes degraded in three different steps, while the PXRD examination showed well-defined sharp crystalline peaks for the complexes, indicating significant crystallinity. The antiproliferative activity of the ligand and its complexes were also evaluated *in vitro* against the HeLa (Human Cervical Cancer Cells) and HCT116 (Colon Cancer Cells) cell lines. The findings suggested complex 4 to be potential anticancer agent against these cell lines. In addition, ligand and its complexes also exhibited considerable emission properties.

**Keywords:** Pyridyl ligand; metal complexes; anticancer activity.

## INTRODUCTION

Schiff base ligands are one of the most prominent nitrogen donor ligands in the field of inorganic chemistry due to their ability to chelate metal ions by donating lone pair of electrons. In addition, Schiff base ligands are easy to synthesize, stable, and adaptable<sup>1-5</sup>. Moreover, the Schiff base complexes are useful in a variety of contexts, including catalysis<sup>6</sup>, photochromism<sup>7, 8</sup>, nonlinear optics<sup>9</sup>, magnetism<sup>10</sup>, coordination chemistry<sup>11</sup>, and materials science<sup>12, 13</sup>.

Pyridyl ligands have gained a lot of attention among the various Schiff base ligands used in coordination chemistry because of their effective chelating properties and ability to develop distinct coordination architectures<sup>12, 13</sup>. The pyridyl functionality also improves electron delocalization and conformational rigidity after coordination to the metal ion, enhancing the luminescent, optical, catalytic, and medicinal properties of the complexes<sup>14-18</sup>. Over the years, cis-platin, a platinum-based drug, has been used to treat almost half of all cancer patients<sup>19</sup>. However, the use of cisplatin is restricted despite its therapeutic effectiveness in the treatment of cancer because it has serious toxic side effects such as nephrotoxicity, neurotoxicity, hematologic toxicity, and ototoxicity. In addition, cisplatin has poor water solubility and can cause acquired resistance in a variety of cancer types<sup>20</sup>.

Keeping in mind the benefits and downsides of platinum-based anticancer medications, researchers are therefore seeking for new potential anticancer medicines with high efficacy across a wide spectrum of tumours and low harmful side effects. Nitrogen-containing ligands and their metal complexes have drawn a lot of attention over the years due to their extensive biological applications, particularly when it comes to developing complexes with anticancer properties<sup>21</sup>, which are attributed to electronic interactions between the metal center and the  $\pi$ -electrons in rings<sup>22-23</sup>. Therefore, taking into account the properties of nitrogen-containing ligands and their complexes, we study here a novel series of complexes of the first series of transition metal ions obtained from pyridyl ligand<sup>12</sup> and investigate their anticancer activity against HeLa (Human Cervical Cancer Cells) and HCT116 (Colon

Cancer Cells) cancer cell lines. In addition, we have also reported their fluorescence behavior.

## EXPERIMENTAL

### Materials and methods

All of the chemicals, including 2,2-Dimethyl-1,3-propanediamine, 2-acetylpyridene, were procured from Sigma-Aldrich and utilized exactly as received. FT-IR spectra were collected on a Perkin Elmer 621 infrared spectrophotometer with KBr pallet.  $^1H$ - and  $^{13}C$ -NMR spectra in  $d_6$ -DMSO, as well as CHN, were obtained using a JEOL-400 spectrometer and an Elementar Varrio EL analyzer, respectively. The RF-6000 Spectro Fluorophotometer was used to obtain fluorescence spectra in methanol. The Powder XRD analysis with XRD pattern was performed using  $Cu K\alpha$  ( $\lambda = 1.54059 \text{ \AA}$ ) radiation on Bruker D2 Phaser X-ray diffractometer.

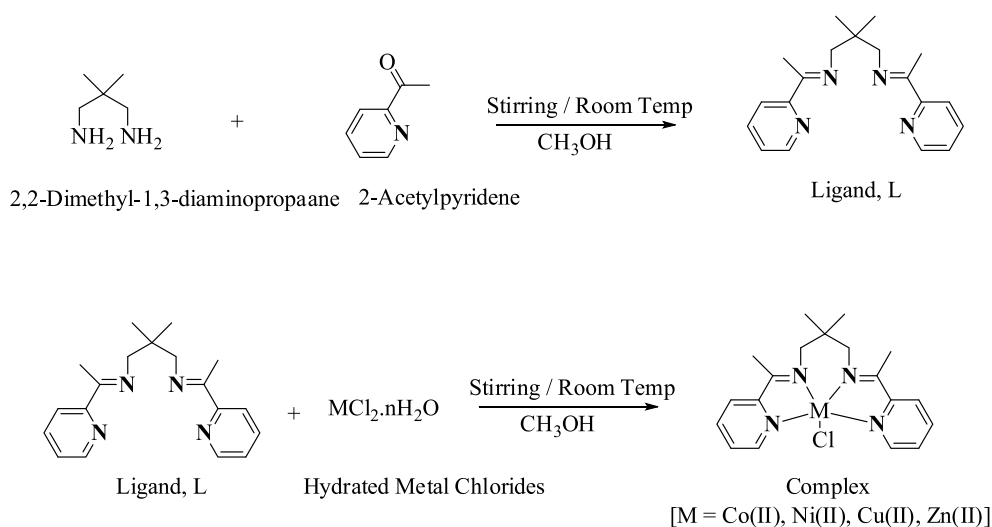
*Synthesis of Pyridyl ligand, (1E,1'E)-N,N'-(2,2-dimethylpropane-1,3-diyl)bis(1-(pyridin-2-yl)ethan-1-imine), L*

A methanolic solution of 2,2-Dimethyl-1,3-propanediamine (100 mg, 0.98 mmol) was mixed with 2-acetylpyridene in a 1:2 molar proportion in the same solvent for 10 hours at ambient conditions, yielding crystalline solid after evaporation (Scheme 1). Unfortunately, we were unable to find a suitable crystal for diffraction.

Color, Yellow; Chemical Formula:  $C_{19}H_{24}N_4$ , Anal. Calc.: C, 73.99; H, 7.84; N, 18.17; Found: C, 73.79; H, 7.79; N, 18.10;  $^1H$  NMR ( $d_6$ -DMSO, ppm): 8.58-7.38 (m, Ar-H, 8H), 3.39 (4H, N-CH<sub>2</sub>), 2.26 (N=C-CH<sub>3</sub>), 1.09 (6H, C-CH<sub>3</sub>),  $^{13}C$ -NMR ( $d_6$ -DMSO, ppm): 165.3 (-C=N), 148.1–157.0 (Ar-C), 36.5 (N-C-CH<sub>3</sub>-CH<sub>2</sub>-), 24.4 (-CH<sub>3</sub>-C-CH<sub>3</sub>), 60.1 (-CH<sub>3</sub>-C-CH<sub>3</sub>), IR (KBr,  $cm^{-1}$ ): 1612

### General procedure for the Synthesis of complexes

To the methanolic solution of ligand, L (50 mg, 0.162 mmol) was added metal salt in a 1:1 proportion followed by the vigorous stirring of the reaction mixture for 5 hours. Slight turbidity was obtained which was eliminated by filtration. The transparent and colored



**Scheme 1.** Schematic representation of the synthesis of ligand and its complexes

solution was dried under vacuum and recrystallized in methanol to obtain an analytically pure compound.

Complex 1: Chemical Formula:  $C_{19}H_{24}ClCoN_4$ ; Color: Brown, Anal. Cal: C, 56.65; H, 6.01; N, 13.91, Found: C, 56.61; H, 5.95; N, 13.84, IR (KBr,  $cm^{-1}$ ): 1618

Complex 2: Chemical Formula:  $C_{19}H_{24}ClNiN_4$ ; Color: Greenish brown; Anal. Cal: C, 56.69; H, 6.01; N, 13.92; Found: C, 56.63; H, 5.96; N, 13.88; IR (KBr,  $cm^{-1}$ ): 1613

Complex 3: Chemical Formula:  $C_{19}H_{24}ClCuN_4$ ; Color: Blue; Anal. Cal: C, 56.01; H, 5.94; N, 13.75, Found: C, 55.97; H, 5.89; N, 13.71; IR (KBr,  $cm^{-1}$ ): 1615

Complex 4: Chemical Formula:  $C_{19}H_{24}ClZnN_4$ ; Color: Yellow; Anal. Cal: C, 55.76; H, 5.91; N, 13.69; Found: C, 55.71; H, 5.85; N, 13.61;  $^1H$  NMR ( $d_6$ -DMSO, ppm): 9.0-7.65 (m, Ar-H, 8H), 3.85 (4H, N-CH<sub>2</sub>), 2.60 (N=C-CH<sub>3</sub>), 1.20 (6H, C-CH<sub>3</sub>),  $^{13}C$ -NMR ( $d_6$ -DMSO, ppm): 165.3 (-C=N), 156.7, 147.9, 136.2, 119.9 (Ar-C), 24.2 (N-C-CH<sub>3</sub>-CH<sub>2</sub>-), 59.9 (-CH<sub>2</sub>-C-CH<sub>2</sub>), 13.3 (-CH<sub>3</sub>-C-CH<sub>3</sub>), 35.2 (-CH<sub>2</sub>-C-CH<sub>2</sub>), IR (KBr,  $cm^{-1}$ ): 1620

#### *In vitro* Cytotoxicity assay

The MTT assay was used to determine the cytotoxic activity of the ligand and its studied complexes. The MTT assay [(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide), a yellow-colored water-soluble tetrazolium salt that cleaves the tetrazolium ring and converts it to a water-insoluble purple colored formazan when it interacts with living cells that contain a mitochondrial enzyme (succinate-dehydrogenase). The MTT [3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide] assay was used to assess the anticancer activity of the investigated compounds in HeLa (Human Cervical Cancer Cells) and HCT116 (Colon Cancer Cells) cancer cell lines under different treatment conditions to detect mitochondrial dehydrogenase activity in living cells.

The pyridyl ligand and its complexes were tested *in vitro* against HeLa (Human Cervical Cancer Cells) and HCT116 (Colon Cancer Cells) cancer cell lines. Before the experiment, all the cancer cell lines were grown in Dulbecco's Modified Eagle Medium (DMEM) with 2 mM/L glutamine, 10% fetal bovine serum (FBS), 50 unit/ml penicillin, and 50 g/ml streptomycin in a 5% CO<sub>2</sub> and 95% air incubator at 37 °C<sup>24-25</sup>. The cells were seeded at a density of  $1 \times 10^4$  cells/well in 96-well plates and cultured for 24 hours in 100  $\mu$ L of DMEM

at 37 °C in a 5% CO<sub>2</sub> atmosphere. Cells were treated in triplicate with increasing concentrations of the tested compounds dissolved in DMSO, ranging from 1–100  $\mu$ M for 24 h. DMSO-treated cells were used as a control. As previously stated<sup>19, 26-27</sup>, cell viability was determined using the tetrazolium dye, MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide. After 24 hours of incubation with the tested compound, the cells were given 10  $\mu$ L liters of freshly prepared MTT (5 mM) solution and incubated for another 2 hours at 37 °C in 5% CO<sub>2</sub>. The supernatants were then extracted, and 100  $\mu$ L DMSO was pipetted into each well to solubilize the blue formazan products produced during MTT incubation<sup>19, 26-27</sup>. To assess cell viability, the absorbance of each well was measured using a UV-visible spectrophotometer at 630–550 nm, and the percent cell viability was calculated as follows:

$$\text{percent viability} = \text{Test OD} / \text{Control OD} \times 100$$

The formula  $y = mx + b$  was used to measure IC<sub>50</sub> values, where m and b indicate slope and intercept, respectively, using Microsoft Excel 2010.

## RESULTS AND DISCUSSION

The Schiff base ligand L, which was made by condensing 2,2-dimethyl-1,3-diaminopropane and 2-acetylpyridine in a 1:2 molar ratio in methanolic medium is shown in Scheme 1. The isolated complexes [M(L)Cl] were produced in methanol by reacting the ligand, L, with metal ions [M = Co(II) (1), Ni(II) (2), Cu(II) (3), and Zn(II) (4)] in 1:1 molar ratio. All of the complexes investigated are non-ionic and stable at room temperature. Unfortunately, all of our efforts to get an appropriate crystal for single crystal X-ray crystallography remained unsuccessful.

The FT-IR spectra of the ligand and its studied complexes were analyzed, and the positions of various characteristic bands confirm the proposed structure for the ligand and its complexes. The most distinctive peak, 1612  $cm^{-1}$ , owing to  $\nu(C=N)$ , confirms the formation of the studied ligand, L, which moved to 1613–1620  $cm^{-1}$  after coordination to metal ions<sup>12, 27-28</sup>. The vibrations of the free ligand due to  $\nu(Ar-CH)$  were detected at 3091  $cm^{-1}$ <sup>28, 29</sup>. However, their location shifts to 3102–3110

$\text{cm}^{-1}$ , once the metal is coordinated to the synthesized ligand<sup>29,30</sup>. Furthermore, the aliphatic vibrations caused by the  $\nu(\text{C-H})$  stretching vibrations of  $-\text{CH}_2/\text{CH}_3$  appeared at  $2918\text{--}2960\text{ cm}^{-1}$ <sup>12</sup> [Supplementary Information Figure S1].

The  $^1\text{H-NMR}$  spectral findings of the pyridyl ligand and its zinc complex are mentioned in the Experimental Section, which support the recommended structure. Signals at  $8.58\text{--}7.38\text{ ppm}$  are assigned to the pyridyl protons in the ligand, L. A proton adjacent to the nitrogen of the pyridyl moiety has a double doublet (1H,  $8.08\text{ ppm}$ ,  $J = 7.6$  and  $5.6\text{ Hz}$ ), while the proton close to the side chain of the pyridyl moiety resonated at  $8.1\text{ ppm}$  with coupling constant  $8.0\text{ Hz}$ . The proton at the para position of the nitrogen of the pyridyl moiety appears at  $7.78\text{ ppm}$  as a multiplier due to coupling and cross-coupling with

neighboring protons. The proton linked by para proton and the proton next to the nitrogen of the pyridyl moiety appeared as a multiplet at  $7.76\text{ ppm}$ . In addition, the chemical shifts due to the aliphatic protons corresponding to  $\text{N}=\text{C}-\text{CH}_3$ ,  $\text{N}-\text{CH}_2$  and  $\text{C}-\text{CH}_3$  ( $6\text{H}$ , s) appear at  $3.39\text{ ppm}$ ,  $2.26\text{ ppm}$  and  $1.09\text{ ppm}$ , respectively. However, these positions are moved to  $9.0\text{--}7.65\text{ ppm}$  (Ar-H, m,  $8\text{H}$ ) when coordinated to zinc ion, whereas the chemical shifts for  $-\text{H}_2\text{C}-\text{CH}_3$ ,  $\text{N}-\text{CH}_2$  at  $1.20\text{ ppm}$  and  $3.85\text{ ppm}$ , respectively [Figure 1 and 2].

The  $^{13}\text{C-NMR}$  findings [Fig. 3] confirm the  $^1\text{H-NMR}$  observations, revealing the presence of a significant azomethine carbon signal at  $165.3\text{ ppm}$ , as well as aromatic carbon signals at  $148.1\text{--}157.0\text{ ppm}$ . Furthermore, the carbon signal owing to  $\text{N}-\text{C}-\text{CH}_3-\text{CH}_2-\text{C}-\text{CH}_2-$  and  $-\text{CH}_3-\text{C}-\text{CH}_3$ ,  $-\text{CH}_3-\text{C}-\text{CH}_3$  appear at  $36.5$ ,  $70.5$ ,  $24.4$

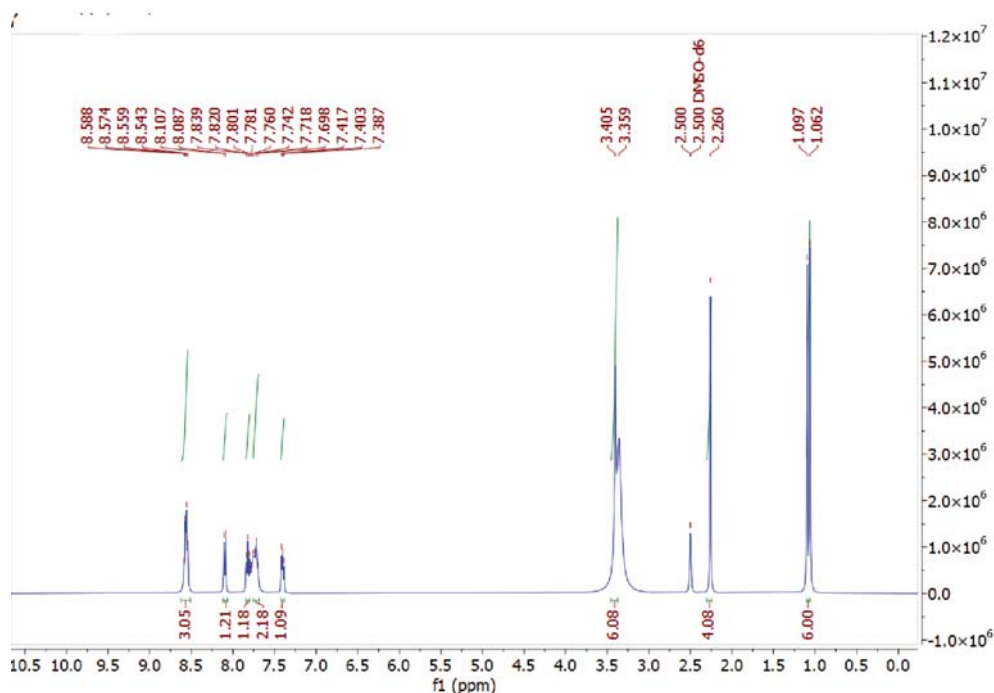


Figure 1.  $^1\text{H-NMR}$  spectrum of ligand, L

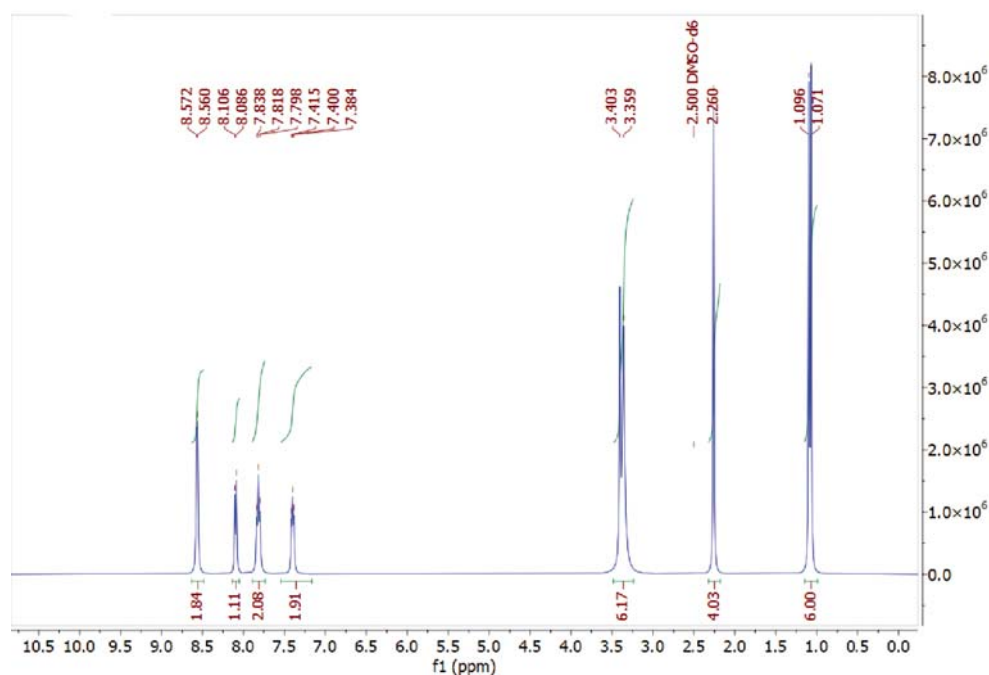
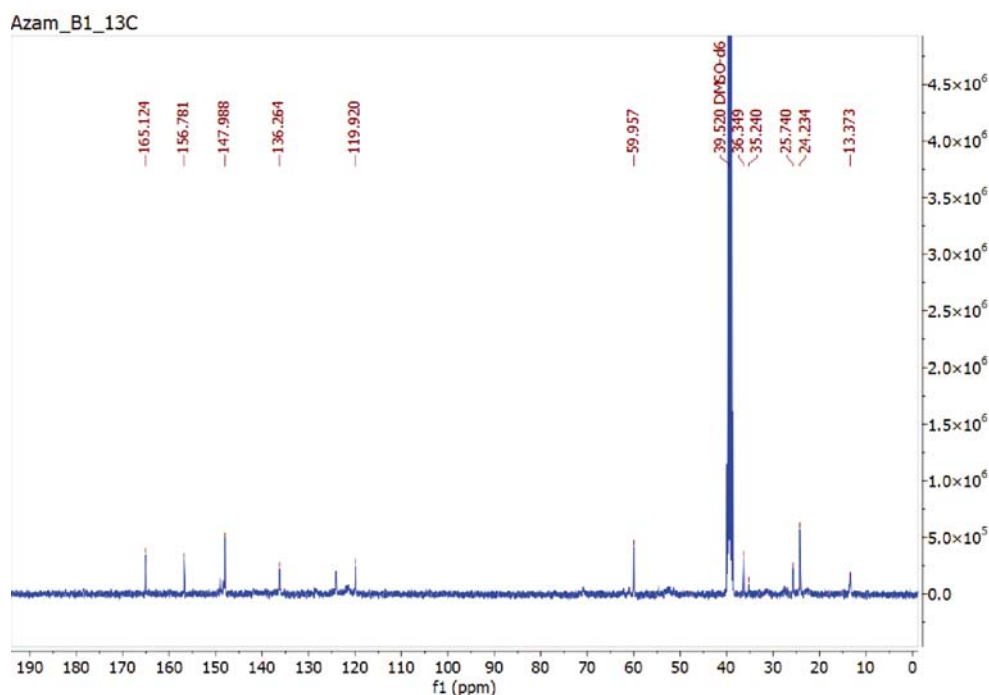


Figure 2.  $^1\text{H-NMR}$  spectrum of complex 4

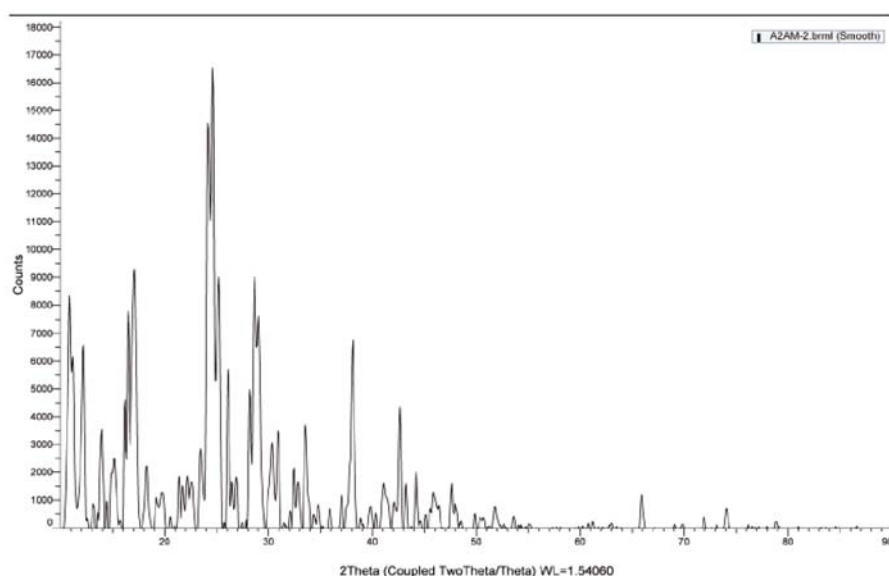


**Figure 3.**  $^{13}\text{C}$ -NMR spectrum of complex 4

and 60.1 ppm, respectively [Supplementary Information Fig. S2]. However, the sites for carbon signals are shifted when metal ion is coordinated.

The Powder XRD analysis of the zinc complex was performed using X- Cu  $K\alpha$  ( $\lambda = 1.54059 \text{ \AA}$ ) radiation powder X-ray powder diffraction for the studied complex was scanned at a wavelength of  $1.5406 \text{ \AA}$  and  $2\theta$  range of  $0^\circ$ – $90^\circ$ . The results showed that the studied complexes had well-defined sharp crystalline peaks, indicating their strong crystallinity. Since all of the complexes have a similar structure, we only display the diffractogram of zinc complex [Fig 4]. The complex shows maximum reflection at  $2\theta = 23.45$  and inter-planar distance at  $d = 3.79 \text{ \AA}$ , which was calculated by Bragg's equation  $n\lambda = 2d\sin\theta$ . Unit cell dimensions values  $a \neq b \neq c$  suggests Orthorhombic crystal structure with monoclinic space group. In addition, Scherrer's formula<sup>21</sup> was also used to measure the average crystallite size for the complex.

TGA was performed in an inert environment of nitrogen at a heating rate of  $10 \text{ }^\circ\text{C min}^{-1}$  and a temperature range of  $53$ – $709 \text{ }^\circ\text{C}$  [Supplementary Information Fig. S3]. The degradation pattern was consistent across all complexes. Therefore, we've included only the TGA degradation of complex 4. The zinc complex causes a total weight loss of 68.41% in three stages. The first step occurs at about  $53$ – $70 \text{ }^\circ\text{C}$ , resulting in a slight weight loss of 7.42% due to one methanol molecule. The second stage of thermal decomposition, which occurred at temperatures between  $150$ – $200 \text{ }^\circ\text{C}$ , accounted for 21.58% of the total mass of the complex caused by the release of one acetyl moiety ( $\text{C}_7\text{H}_7\text{NO}$ ). The third step, the largest weight loss stage, which happens at roughly  $325 \text{ }^\circ\text{C}$ , accounts for 39.41% of the overall mass of the complex, equivalent to the whole remaining organic moiety. The production of ZnO as a final product (31.55%) is then indicated by a straight line.



**Figure 4.** Powder X-Ray Diffractogram for complex 4



### In-vitro Anticancer activity

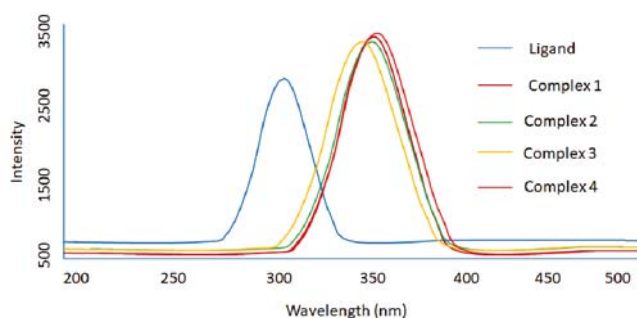
MTT test was used to assess the antiproliferative activity of the ligand and its investigated compounds against HeLa (Human Cervical Cancer Cells) and HCT116 (Colon Cancer Cells). The cytotoxic behavior of the cancerous cell lines was investigated at various concentrations ranging from 1.0  $\mu\text{M}$  to 50  $\mu\text{M}$  for 24 h. However, at maximum concentration of 50  $\mu\text{M}$  for cancer cell line HeLa, the ligand, complex 1, and 2 displayed 27.5%, 30.2%, and 31.7% cell viability, whereas the cell viability for cancer cell lines HCT116 for the ligand and complex 2 and 4 was observed 14.5%, 21.3%, and 25.8%. Table 1 shows the estimated IC<sub>50</sub> values for both cell lines. Complex 4 With IC<sub>50</sub> values of 20.9  $\mu\text{M}$  and 17.5  $\mu\text{M}$  showed strong anticancer activity against HeLa and HCT116 cell lines, respectively. Complex 3 also had significant anticancer activity against HeLa and HCT116 cell lines, with IC<sub>50</sub> values of 22.7  $\mu\text{M}$  and 21.5  $\mu\text{M}$ , respectively. However, complex 2 was discovered to be moderately active against HeLa and HCT116, with an IC<sub>50</sub> value of 23.2  $\mu\text{M}$  and 23.8  $\mu\text{M}$ , respectively. In contrast to complexes 2, 3, and 4, ligand and complex 1 displayed low anticancer activity, with IC<sub>50</sub> values of 31.5  $\mu\text{M}$  and 28.5  $\mu\text{M}$ , and 25.7  $\mu\text{M}$  and 24.1  $\mu\text{M}$ , respectively against the HeLa and HCT116 cell lines. Overall, the findings indicate that all of the complexes have greater anticancer activity than the parent ligand. Complex 4 has, on the other hand, demonstrated potential anticancer activity in both cell lines.

**Table 1.** IC<sub>50</sub> values for the cell lines

Compound	IC <sub>50</sub> ( $\mu\text{M}$ )	
	HeLa	HCT116
L	31.5	28.5
1	25.7	24.1
2	23.2	23.8
3	22.7	21.5
4	20.9	17.5

### Fluorescence properties

The fluorescence properties of the Schiff base ligand, L, and its complexes were reported in methanol at room temperature [Fig 5]. The emission peak appeared at 344 nm when the ligand was excited at 320 nm. The fluorescence activity of ligand is due to the intraligand  $\pi$ - $\pi^*$  transitions. All the complexes emitted a strong fluorescence emission band in the range of 346–351 nm when excited at 320 nm. However, complex 4 emits a broad emission peak. Thus, the emission peak of the free ligand and its metal complexes differed significantly, showing ligand-metal ion coordination.



**Figure 5.** Fluorescence spectra of ligand and its complexes

### CONCLUSION

A novel series of complexes with metal ions from the first transition series were produced from a pyridyl ligand and investigated using a variety of different spectroscopic techniques.

The investigated compounds showed significant ability to inhibit the growth of tumors when tested on Human cervical (HeLa) and colon cancer cells (HCT116). However, the findings revealed that complex 4 with the lowest IC<sub>50</sub> has potential anticancer activity against all these cell lines.

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