

# Antagonism between lead and zinc ions in plants

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**Abstract:** The article presents the results of research which describes antagonism between Pb-Zn in selected plant species from the area of Czestochowa – Mirow district (north-western part of the Czestochowa Upland). There were analyzed changes in the ratio of Pb/Zn in different organs of the tested plants as a function of the Zn content changes. The content of metals in the plants and the soil was determined using atomic absorption spectrophotometry AAS. In all organs of the plants there was observed antagonistic decrease of Pb uptake and accumulation, resulting from the increase in the concentration Zn.

Antagonism between Zn and Pb in roots of the tested plants occurred at Zn content of 200–600 µg/g. In turn, antagonism in stems and flowers occurred at lower contents of zinc (100–180 µg/g). In leaves, antagonism between Pb and Zn occurred when Zn was present at the level of 300–800 µg/g.

Ex definition of the analyses confirm the presence of antagonism of lead with regard to high levels of Zn. The study also confirmed that the degree of antagonism depends on the plant species.

## Introduction

In recent times, plants contamination with various metal compounds related to the effects of human activity has been observed (Sas-Nowosielska 2009, Ociepa et al. 2008, Hławiczka 2008, Bednarek et al. 2005). Heavy metals of industrial pollutions prevent the full development of flora growing on spoil tips or the areas. The sources of plant pollution with heavy metals are also landfills, non-ferrous metal smelters, mines, plant protection chemicals, pesticides and fertilizers. The plants growing on the periphery of urban centers and busy roads contain 30–300 µg/g of Pb, and even > 1000 µg/g of Pb, wherein the average content of this element in the plant is in the range from 0.01 to 2.0 µg/g of Pb (Nagajyoti et al. 2010, Jadia and Fulekar 2009).

Ions of heavy metals may be a potential source of danger for the human and animals (Karczewska et al. 2008, Gruca-Królikowska and Waclawek 2006). Children and the elderly are particularly vulnerable to the toxic effects of heavy metals (especially cadmium and lead). Therefore there is an urgent need to minimize the negative effects of heavy metals on living organisms (Ociepa-Kubicka and Ociepa 2012, Kwiatkowska-Malina and Maciejewska 2011, Krzywy et al. 2010, Suna et al. 2010, Kabata-Pendias and Mukherjee 2007).

The uptake of toxic elements from contaminated soils by plants often exceeds their acceptable norms and leads to phytotoxicity (Niesiobędzka et al. 2005). The distribution of trace metal ions generally can be arranged as follows: roots

> stems > leaves > fruit > seeds (Jin et al. 2005). The uptake of metal ions by the plants depends not only on the content of the compounds of these elements in the soil, but also on the affinity of the plant in relation to the metal (Krzyszowska et al. 2010, Peralta-Videa et al. 2009, Vaillant et al. 2005, Lehmann and Rebele 2004).

In the mineral nutrition in plants there is often an antagonism between ions. The most significant antagonism exists between mono- and divalent cations, but it also occurs among cations of the same valency and anions. Each metal ion has a specific, characteristic manner of uptake, transport and accumulation in the plant, but in the presence of other ions in the soil there are also noted interactions between the ions (Eapen and D'souza 2005). These interactions can be of a neutral nature, wherein the occurrence of one element does not affect the uptake of others from the soil, or a synergistic one wherein there is observed an increase of one ion uptake in the presence of another and an antagonistic one – which is about stopping or reducing the uptake of one metal in the presence of another (Musielińska et al. 2014, Rout and Das 2003). As an example of the latter type of the interactions can serve antagonistic decrease in the uptake and accumulation of heavy metals in plants, e.g. ions of Pb or Cd with the increase of the concentration of trace elements in the soil, e.g. Zn (Aslan et al. 2003, Bolan et al. 2003, Sady 2001).

Concentration of heavy metals in soils, occurring especially in an easily bioavailable form to plants, may be the cause of an excessive uptake and negative influence on

plants (Siwek 2008, Terelak et al. 2000). Under the influence of lead and other heavy metals inhibition of growth and delay of germination processes were observed (Furmanek and Andrzejewska-Ponomarev 2006, Rao 2006, Usman et al. 2006). The simultaneous occurrence of both lead ions as well as other metal ions (e.g., zinc) during their accumulation in different organs of the plant may lead to the antagonism of lead. Zinc plays an important role in plant metabolism. Both the deficiency or excess of the metal limit the growth and development of plants. Zinc is taken by the plant proportionally to its content in the soil, although both soil properties and the choice of the species have a significant influence on its content in plants. Therefore, it seems important to base the discussion of the phytoaccumulation abilities of different plant species on metal speciation analysis and on determining the way of the co-occurrence of selected elements (Krzesłowska et al. 2010, Gruca-Królikowska and Waclawek 2006).

In this light, the research which is not only to confirm the occurrence of antagonism between the selected metal ions in plants with similar contents of bioavailable forms of these metals in the soil, but also to answer the question of whether a potential antagonism between lead and zinc ions depends on the plant species seems to be justifiable.

## Materials and methods

The study was conducted in the area of Czestochowa – Mirow district (240 ± 5 m a. s. l. N 50°49'20.05" E 19°13'00.20"; N 50°49'20.47" E 19°13'18.34" ± 0.50"). This area is a part of the Jurassic Landscape Parks and is directly adjacent to the "Gaszczyk" – the planned Wladyslaw Hyla forest reserve. It is located in the north-western part of the Czestochowa Upland (Poland) (Hereźniak 1984).

Plants were collected in the flowering period (60 species of 15 pieces) from 14 stands, each of an area of 5 m<sup>2</sup>. The plants washed with redistilled water and air dried in well ventilated, dry and clean rooms were crushed in an agate mortar, and ground using various types of mills, with special plastic blades. Samples of mass 1 g ± 0.00001 quantitatively transferred to conical flasks were treated with 10 cm<sup>3</sup> of spectrally pure concentrated (65%) HNO<sub>3</sub> Supra Pure Merck (Germany). The plant samples were digested in conical flasks of 100 cm<sup>3</sup> sand bath at a temperature of 70°C ± 2°C until the clear solution.

Soil samples were taken after the thaw from the root layer of the tested plant habitats, from the surface without undergrowth of size 20–50 cm<sup>2</sup>. The soil, (each sample of 1.5 kg) was dried using radiant tubes and ground in a mortar and sieved through a 1 mm mesh. In the next step the samples were ashed in a muffle furnace (temperature 400–600°C) until the constant weight. Next, the soil samples (1 g each) were treated with spectrally pure concentrated (65%) HNO<sub>3</sub> Supra Pure Merck (Germany) in an amount of 5 cm<sup>3</sup>. In order to isolate the various chemical forms of metal occurrence in the soil the samples were subjected to speciation analysis. The methodology of Rudd (Rudd et al. 1988), was applied, which is based on sequential extraction. Extractions of each of the samples (sample weight of the soil 1 g ± 0,00001) were carried out using a spectrally pure reagents in a ratio of 1:40: removable form – 1,0 M KNO<sub>3</sub>; adsorbed form – 0.5 M KF, (pH = 6.5); organic connections – 0.1 M Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>; carbonates

– 0.1 M EDTA (pH = 6.5); sulphides – 6.0 M HNO<sub>3</sub>. The tested soil samples were treated with the mentioned above solutions within 24 hours. The next step was centrifugation (for 10 minutes) of the analyzed samples (400 rev./min), and then the samples were washed with redistilled water (25 ml) and subjected to the centrifugation once more (for 5 min – 400 rev./min). The obtained extracts were acidified to a concentration of 1% HNO<sub>3</sub>. The recovery procedure ensured Pb and Zn in 99%. Each series of measurements was conducted with three blind tests, the values of which were taken into account in the results of the tested samples. The heavy metal content in the tested samples of soils and plants was determined by atomic absorption spectrophotometry using a Spectrophotometer Pye Unicam SP9 with the use of a hollow cathode lamp and acetylene-air flame, to the nearest 0.01 µg/g (Whiteside and Milner 1984). The detection of the studied metals was [µg/cm<sup>3</sup>]: Pb – 0.100; Zn – 0,010.

The precision and accuracy of assays was monitored with the standard addition method by National Metrology Institute as well as with a certified reference material of a comparable content of the tested metal in relation to the analysed material: for soils-CRM277 (BCR); for plants-CTA-VTL-2 (ICH and TJ). The differences were 2–4%.

In addition, an intercalibration study (6 reference samples of the tested soil) was conducted in the Department of Environmental Monitoring, Central Mining Institute in Katowice. The results of analyses in comparison with the results of intercalibration study differ in the range of 3–5%.

In the case of some plant species there was no material representing a specific organ. Such cases are indicated in Table 2 and in Figs. 1–4 with the mark (\*), while in Figs. 5–8 there is included a number of samples, giving the value of n for each organ of the tested plants.

## Results and discussion

The content of lead and zinc [µg/g] in soil samples and the percentage of metal in different chemical forms are shown in Table 1. Based on the obtained results of the lead and zinc content in the soil, one can rank these metals as follows: Zn > Pb. Analysis of the results showed that the total lead content does not reflect the mobility and thus availability for plants. In the case of zinc one can indicate an inverse relation. The participation of zinc in two most mobile forms, exchangeable and adsorbed ones, shows a considerable availability for plants. According to the literature (Terelak et al. 2000), lead, due to the relatively limited solubility of minerals, is less mobile than zinc in the environment.

In the studied soils, there were small differences in the amount of lead (the standard error was 0.1 µg/g for n=14). The total content of it in the soil was 23.5 µg/g which was similar to the results obtained in the soils of the north-eastern Poland – 29 µg/g (Niesiobędzka 2000). The average content of lead in agricultural land in Poland is 14 µg/g (Kabata-Pendias and Pendias 2001-a), in the Kłodzko Valley – 10 µg/g (Waclawek and Maćko 2001). According to Czarnowska (Czarnowska 1996) "geochemical background" the content this metal is 3 to 18 µg/g soil. Terelak and co-authors (Terelak et al. 2000) indicate that the average lead content in soils of agricultural land in Poland is 13.8 µg/g. The level of lead in the tested soil did not exceed the limit value of 100 µg/g (Dz. U. No.

165/2002 item. 1359). IUNG (Sauve et al. 1997) assumes as a natural value of lead concentration: about 31 mg/kg of Pb in light acid soils, 50 mg/kg of Pb in light neutral soils, and 70 mg/kg in heavy soils with a pH slightly acidic or neutral. The results obtained allow to classify the tested soils as “clean” ones with natural content of this component.

Zinc content in soil is typically in the range of 10–200 µg/g (Dziubanek et al. 2012). In the tested soil the zinc content was at the level of 89.39 µg/g, and the result is within the accepted limits, but it exceeds the content suggested by other authors. Kabata-Pendias (Kabata-Pendias and Pendias 2001-a) provides the limit of Zn content of 80 µg/g and Terelak and co-authors (Terelak et al. 2000) believe that the content of zinc in soils in Poland does not exceed 40 µg/g. The literature data indicate that in the areas of urban and industrial emissions the zinc content in soils can exceed the normative values of 300 µg/g (Dziubanek et al. 2012), but in this case it does not. A comparison of the identified zinc content with the cited data (Niesiołędzka 2000) showed that the tested soils contain this element in geochemically average amounts.

Coexistence of Pb with Zn is described by the correlation coefficient: the positive one indicates directly proportional changes, and the negative one – changes which are inversely proportional. The interdependence of lead and zinc is described by the correlation coefficient, which in the exchangeable form in soil is 0.87 and in the absorbed form it is 0.38. In the both cases the positive coefficient indicates directly proportional relation. According to Zwonitzer and co-authors (Zwonitzer et al. 2003), zinc usually accompanies lead in the soil, which is reflected in the obtained results.

The contents of lead and zinc in different organs of the studied plant species are shown in Table 2.

The content of heavy metals in plants is generally a reflection of the amount of these bioavailable forms in soil. The results regarding the occurrence of heavy metals in plants are a valuable complement to the existing knowledge about the accumulation of metals in plants. In the analyzed plant species the lead content was in the range [µg/g]: 2.49–33.80 for roots; 2.49–8.32 for the stems; 2.48–5.26 for flowers; 2.49–13.65 for leaves. The zinc content in different organs of the plant was [µg/g]: root: 20.79–531.31; stem: 9.03–171.92; flower: 20.09–170.68; leaf: 16.51–754.94. One must note that the level of Pb and Zn in some species exceeded the amount of these

metals reported by other authors, e.g. in the species collected from southern Poland (Łaszewska et al. 2007). According to the literature, the average lead content in plants from contaminated areas is even up to 387 µg/g (Wybieralski and Maciejewska 2011), while from uncontaminated areas it is about 0.4 µg/g (Niesiołędzka 2000) and 0.35 µg/g (Wacławek and Maćko 2001). In grasses studied on permanent pastures near Warsaw (Wawkonowicz et al. 2011), lead content was in the range of 3.9 to 7.5 µg/g. These results do not indicate significant pollution of the species with the studied metal, however, they exceed the levels obtained by other cited authors. Wawkonowicz and co-authors (Wawkonowicz et al. 2011) indicate that in grasses the zinc content (Zn) ranged between 16 and 48 µg/g. However, according to Kabata-Pendias and co-author (Kabata-Pendias and Pendias 2001-a) the content of Zn in grasses in Poland is in the range from 12 to 72 µg/g, with an average of 30 µg/g. The results indicate that in the case of some plant species (Table 2) these values were significantly exceeded. The largest amounts of the metals were accumulated in the roots and leaves of the plants.

Lead is unevenly accumulated by the plant. Its concentration is greatest in those organs of the plants which are exposed to direct contact with its compounds. It is mainly accumulated (93–96%) in the roots of plants (Badora and Kozłowska-Strawska 2011, Siwek, 2008, Gruca-Królikowska and Wacławek 2006). Only a small part of the quantities drawn gets into the shoots. The uptake of the metal ions by the roots is a passive process and depends on the levels of bioavailable forms in soil (Janowska and Szymańska 2005). The high levels of lead content in leaves of some plant species may indicate that the lead compounds from the atmosphere went into leaves through the stomata, or after a prior cuticle damage, for example, as a result of acid rain or ozone. This problem does not apply to high zinc content in the leaves of some species, because the cuticle and the hair layer are not a barrier for it (Skołozdrzy et al. 2001, Terelak et al. 2000, Woźny 1995).

Incidentally high levels of these elements were also in flowers and stems of some species. This fact may suggest the contamination of the plants with these metals, as a result of atmospheric dust fall (Kabata-Pendias and Pendias 2001-a). In the case of zinc it can be a natural process comparable to ions metal accumulation in roots and shoots (Siwek 2008).

**Table 1.** Concentration of lead and zinc [µg/g] in soil samples and the percentage of the individual chemical forms in the soil

Concentration of lead and zinc in the individual chemicals forms in the soil (the standard error 0,1 µg/g; n=14)							
Metal	Total content	Removable form	Absorbed form	Organic connections	Carbonates	Sulphides	Residual
Zn	89.39	2.49	54.43	11.03	10.42	4.05	6.96
Pb	23.5	1.79	5.32	4.76	7.30	1.76	2.57
Percentage %							
Metal	Removable form	Absorbed form	Organic connections	Carbonates	Sulphides	Residual	
Zn	2.79	60.89	12.34	11.66	4.53	7.79	
Pb	7.62	22.64	20.25	31.06	7.49	10.94	

Table 2. The mean content of lead and zinc, and the ratio of Pb/Zn in different organs of the selected plant species [ $\mu\text{g/g}$ ]

PLANTS	STEMS			ROOTS			FLOWERS			LEAVES		
	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn
<i>Plantago lanceolata</i> L.	3.08	67.54	0.046	5.12	133.33	0.038	4.43	48.86	0.091	3.19	98.02	0.033
<i>Plantago major</i> L. s. str.	2.48	24.20	0.103	16.19	131.52	0.123	5.13	50.27	0.102	5.90	76.13	0.077
<i>Pimpinella major</i> (L.) Huds.	2.57	19.76	0.130	9.42	32.37	0.291	2.49	38.90	0.064	(*)	(*)	(*)
<i>Melandrium album</i> (Mill.) Garcke	2.69	93.85	0.029	4.67	106.39	0.044	2.72	73.54	0.037	3.54	247.08	0.014
<i>Artemisia vulgaris</i> L.	2.77	78.18	0.035	2.50	33.79	0.074	(*)	(*)	(*)	3.29	78.01	0.042
<i>Arnoseris minima</i> (L.) Schweigg. et Koerte	2.67	51.62	0.052	9.50	193.33	0.049	3.57	62.15	0.057	3.77	168.49	0.022
<i>Succisa pratensis</i> Moench	2.58	69.75	0.037	9.81	531.31	0.018	2.50	66.88	0.037	5.42	324.60	0.017
<i>Scabiosa ochroleuca</i> L.	2.63	24.55	0.107	6.26	54.00	0.116	2.70	66.31	0.041	4.29	74.70	0.057
<i>Verbascum lychnitis</i> L.	2.78	33.96	0.082	7.00	52.67	0.133	2.79	72.80	0.038	6.61	224.66	0.029
<i>Verbascum nigrum</i> L.	2.96	47.44	0.062	7.74	72.13	0.107	2.48	49.38	0.050	4.57	141.03	0.032
<i>Verbascum densiflorum</i> Bertol.	2.92	40.48	0.072	6.55	81.40	0.080	2.78	49.76	0.056	7.13	172.36	0.041
<i>Carlina vulgaris</i> L.	3.53	128.73	0.027	4.85	165.20	0.029	2.65	59.80	0.044	3.88	172.55	0.022
<i>Hypericum hirsutum</i> L.	2.48	133.59	0.019	4.77	212.36	0.022	2.48	48.93	0.051	2.49	65.77	0.038
<i>Hypericum perforatum</i> L.	5.27	30.31	0.174	9.50	25.83	0.368	2.50	86.79	0.029	4.07	222.45	0.018
<i>Prunella grandiflora</i> (L.) Scholler	2.96	70.31	0.042	2.50	144.82	0.017	5.33	43.59	0.122	3.46	30.31	0.114
<i>Peucedanum palustre</i> (L.) Moench	2.50	42.12	0.059	2.50	70.08	0.036	3.71	96.95	0.038	(*)	(*)	(*)
<i>Dianthus carthusianorum</i> L.	2.50	171.92	0.015	4.07	230.60	0.018	2.93	125.31	0.023	(*)	(*)	(*)
<i>Torilis japonica</i> (Houtt.) DC.	3.65	24.10	0.152	5.30	36.51	0.145	2.98	58.42	0.051	11.34	79.71	0.142
<i>Carum carvi</i> L.	8.49	35.17	0.241	23.36	25.46	0.918	3.00	63.00	0.048	13.65	104.16	0.131
<i>Chenopodium album</i> L.s.str.	2.50	23.77	0.105	3.36	23.64	0.142	2.50	76.26	0.033	2.50	103.08	0.024
<i>Trifolium pratense</i> L.	2.50	20.07	0.125	3.71	30.38	0.122	2.50	46.67	0.054	2.50	65.91	0.038
<i>Trifolium medium</i> L.	2.63	27.48	0.096	(*)	(*)	(*)	4.04	49.46	0.082	3.84	54.12	0.071
<i>Valeriana dioica</i> L. s. str.	2.50	36.69	0.068	5.44	67.58	0.080	2.50	49.31	0.051	4.07	68.42	0.059
<i>Oenanthe fistulosa</i> L.	3.18	67.89	0.047	3.70	80.14	0.046	2.97	92.55	0.032	(*)	(*)	(*)
<i>Achillea millefolium</i> L.	3.28	47.98	0.068	9.69	50.05	0.194	3.05	62.77	0.049	3.38	109.81	0.031
<i>Sanguisorba officinalis</i> L.	3.11	30.81	0.101	2.50	66.34	0.038	2.77	75.25	0.037	2.49	80.26	0.031
<i>Silene vulgaris</i> (Moench) Garcke	2.50	93.27	0.027	(*)	(*)	(*)	2.50	106.19	0.024	3.11	128.87	0.024
<i>Linaria vulgaris</i> Mill.	3.35	79.54	0.042	7.31	80.09	0.091	2.51	72.44	0.035	3.21	216.24	0.015
<i>Atriplex patula</i> L.	2.67	18.05	0.148	3.29	44.48	0.074	2.75	57.79	0.048	3.00	149.49	0.020
<i>Thymus serpyllum</i> L.	3.63	124.99	0.029	4.19	117.30	0.036	3.23	170.68	0.019	3.66	295.84	0.012
<i>Papaver rhoeas</i> L.	2.50	30.35	0.082	10.84	28.41	0.382	(*)	(*)	(*)	3.71	91.36	0.041

Table 2. The mean content of lead and zinc, and the ratio of Pb/Zn in different organs of the selected plant species [µg/g] – cont.

PLANTS	STEMS			ROOTS			FLOWERS			LEAVES		
	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn	Pb	Zn	Pb/Zn
<i>Taraxacum officinale</i> F. H. Wigg.	2.64	50.47	0.052	(*)	(*)	(*)	2.63	30.19	0.087	2.50	52.95	0.047
<i>Saponaria officinalis</i> L.	2.50	15.72	0.159	2.93	24.20	0.121	2.50	30.57	0.082	3.11	42.96	0.072
<i>Solidago canadensis</i> L.	2.49	41.96	0.059	(*)	(*)	(*)	2.49	21.73	0.115	2.49	40.88	0.061
<i>Melilotus alba</i> Medik.	2.50	16.18	0.155	2.75	28.65	0.096	2.50	36.81	0.068	2.50	49.30	0.051
<i>Avena sativa</i> L.	3.24	9.03	0.359	12.62	49.30	0.256	2.50	20.09	0.124	3.00	16.51	0.182
<i>Typha angustifolia</i> L.	2.50	38.93	0.064	(*)	(*)	(*)	2.50	50.06	0.050	2.50	22.59	0.111
<i>Urtica dioica</i> L.	2.75	24.69	0.111	3.12	25.64	0.122	2.50	34.17	0.073	3.10	45.03	0.069
<i>Anthyllis vulneraria</i> L.	2.50	12.57	0.199	(*)	(*)	(*)	2.50	27.35	0.091	2.50	26.21	0.095
<i>Erigeron ramosus</i> (Walters) Britton. Sterns et Poggenb.	2.49	71.99	0.035	2.49	94.57	0.026	2.64	39.01	0.068	4.41	283.19	0.016
<i>Galium saxatile</i> L.	2.72	155.28	0.017	9.32	177.66	0.052	2.63	61.10	0.043	2.73	94.78	0.029
<i>Berteroa incana</i> (L.) DC.	2.52	152.77	0.017	6.61	184.64	0.036	2.50	62.32	0.040	2.50	262.60	0.010
<i>Thalictrum minus</i> L. ssp. minus	2.50	38.21	0.065	(*)	(*)	(*)	(*)	(*)	(*)	2.50	90.71	0.028
<i>Rorippa sylvestris</i> (L.) Besser	2.79	64.90	0.043	2.50	23.29	0.107	3.11	107.76	0.029	2.82	173.70	0.016
<i>Falcaria vulgaris</i> Bernh.	2.50	58.69	0.043	(*)	(*)	(*)	2.50	52.17	0.048	(*)	(*)	(*)
<i>Senecio paludosus</i> L.	2.50	57.61	0.043	2.50	20.79	0.120	2.50	67.58	0.037	2.50	226.53	0.011
<i>Senecio jacobaea</i> L.	3.06	57.57	0.053	8.19	71.59	0.114	2.62	78.92	0.033	4.36	236.60	0.018
<i>Senecio sylvaticus</i> L.	8.32	156.99	0.053	33.80	215.29	0.157	2.50	145.83	0.017	7.52	754.94	0.010
<i>Senecio vernalis</i> Waldst. et Kit.	2.95	50.19	0.059	4.40	62.65	0.070	5.26	60.36	0.087	6.18	93.95	0.066
<i>Senecio aquaticus</i> Hill	2.49	91.01	0.027	3.33	139.05	0.024	2.49	103.38	0.024	(*)	(*)	(*)
<i>Descurainia sophia</i> (L.) Webb ex Prantl	2.50	23.66	0.106	7.70	138.00	0.056	3.65	55.13	0.066	2.93	118.02	0.025
<i>Cicuta virosa</i> L.	2.87	94.07	0.031	6.06	106.10	0.057	3.71	75.85	0.049	3.00	424.97	0.007
<i>Rumex hydrolapathum</i> Huds.	2.86	30.34	0.094	5.71	65.39	0.087	3.17	48.15	0.066	5.59	126.50	0.044
<i>Malva moschata</i> L.	3.28	38.27	0.086	(*)	(*)	(*)	2.48	85.66	0.029	2.49	130.94	0.019
<i>Anthriscus sylvestris</i> (L.) Hoffm.	3.22	45.02	0.072	3.13	42.66	0.073	2.50	27.74	0.090	2.49	55.39	0.045
<i>Filipendula ulmaria</i> (L.) Maxim.	2.48	79.07	0.031	2.50	115.55	0.022	(*)	(*)	(*)	(*)	(*)	(*)
<i>Euphorbia cyparissias</i> L.	2.50	100.01	0.025	6.17	107.77	0.057	(*)	(*)	(*)	2.49	146.13	0.017
<i>Tanacetum vulgare</i> L.	3.29	25.45	0.129	2.49	26.60	0.094	2.66	36.60	0.073	2.59	138.40	0.019
<i>Vicia sepium</i> L.	2.50	119.34	0.021	(*)	(*)	(*)	(*)	(*)	(*)	2.64	93.06	0.028
<i>Leucanthemum vulgare</i> Lam.	2.50	27.74	0.090	(*)	(*)	(*)	3.46	32.69	0.106	(*)	(*)	(*)

(\*) brak danych

The results of analyzes have also shown a relation between the total lead content and its level in plants. The lead content in some plants (Table 2) did not correlate with a lead content in bioavailable forms (Table 1), which may be due to the fact that 73 to 95% of the metal compounds in plants is of atmospheric origin, not geochemical one from the soil (Kabata-Pendias and Pendias 2001-a).

Lead is one of the elements strongly accumulated in the plant. The observed high correlation coefficient between the lead and zinc in the exchangeable form in soil could suggest that in different plant organs one can also notice apparent positive correlation between these metals. However, in the literature (Aslan et al. 2003, Bolan et al. 2003) there is emphasized the presence of antagonism between metals and macro- or microelements, therefore, this issue is illustrated by a series of engravings, that show the changes to the quotient of the contents of Pb/Zn in the tested plants as a function of Zn content changes Fig. 1–4.

In all the analyzed organs of the plants, together with the increase of zinc content there is observed the antagonism of lead, the content of which is maintained at the level characteristic for the species (Figs. 1–4). In the roots of the tested species, with the zinc content of 200 to 600 µg/g, antagonism of lead is found in the following plants: St. John's wort (*Hypericum hirsutum* L.), Carthusian Pink (*Dianthus carthusianorum* L.), and Devil's-bit Scabious (*Succisa pratensis* Moench.) – Fig. 1.

Antagonism of lead in the stems of the tested plants due to large content of zinc (range 100–180 µg/g) refers to the species: cypress spurge (*Euphorbia cyparissias* L.), bush vetch (*Vicia sepium* L.), wild thyme (*Thymus serpyllum* L. emend. Fr.), Carlina thistle (*Carlina vulgaris* L.), hairy St. John's wort (*Hypericum hirsutum* L.), hoary alyssum (*Berteroa incana* (L.) DC.), Heath Bedstraw (*Galium saxatile* L.), Woodland ragwort (*Senecio sylvaticus* L.) and Carthusian Pink (*Dianthus carthusianorum* L.) – Fig. 2.

When the zinc content is from 100 to 180 µg/g, antagonism of lead is in flowers of the following plant species (Fig. 3): Marsh Ragwort (*Senecio aquaticus* Hill), Bladder campion (*Silene vulgaris* (Moench) Garcke), Creeping Yellowcress (*Rorippa sylvestris* (L.) Besser), Carthusian Pink (*Dianthus carthusianorum* L.), Woodland Ragwort (*Senecio sylvaticus* L.) and wild thyme (*Thymus serpyllum* L. emend. Fr.).

Antagonism between lead and zinc were also observed in the leaves of individual plant species (Fig. 4). When zinc content is 300 to 800 µg/g, antagonism of lead refers to the following species: Devil's-bit Scabious (*Succisa pratensis* Moench.), Cowbane (*Cicuta virosa* L.) and Woodland Ragwort (*Senecio sylvaticus* L.).

One of the important factors affecting the uptake of the metals from soil is a plant species and root structure. The literature widely describes the differences in the uptake and accumulation of metals depending on the element, plant species and population (Krzyszowska et al. 2010, Peralta-Videa et al. 2009, Korzeniowska and Stanisławska-Głubiak 2007, Vaillant et al. 2005, Tyksiński and Kurdubska 2004, Lehmann and Rebele 2004, Shu et al. 2002, Mengel et al. 2001, Kabata-Pendias and Pendias 2001-b, Marschner 1995). So far, there has also been proved that an adequate content of microelements can lead to a reduction in accumulation of heavy metals in plants (Gruca-Królikowska and Waclawek 2006). According to Kabata-Pendias, zinc impairs the transport of lead to the aerial

parts of plants (Kabata-Pendias and Pendias 2001-a), which could confirm the observed phenomenon of discrimination of lead by zinc in the case of flowers, stems and leaves of some tested species.

In conclusion, the above analyses show that the potential antagonism between Pb and Zn occurs only in the case of some plants (and within the selected organs of the plants). Therefore, in order to illustrate better the degree and way of coexistence of the analyzed metals in different organs of the tested plants the linear regression equation was used (Figs. 5–8). The interpretation of the regression equations presented below is as follows: the coefficient before the metal is a measure of the increase (or decrease) of the contents of one element while changing the content of the other element; the intercept is the value above which one can observe a certain relation, which is described by the correlation coefficient.

The change of Pb content in the function of Zn content changes in roots of the tested plant species is described by regression equation (Fig. 5):  $Pb = 0,0143 * Zn + 5,26$ ;  $R^2 = 0,0485$ . According to the above interpretation, the change in Zn content by 100 units will result in the increase of Pb in the root by 1.43 µg, but only above the level of Zn equal to 5.26. The above conclusion is justified for 4.85% of the cases. Analysing the equation describing the change of Pb and Zn in stems of the tested plants (Fig. 6):  $Pb = 0.0037 * Zn + 2.78$ ;  $R^2 = 0.018$ , we note that the change in Zn content by 100 units causes an increase of Pb content in the stems by 0.9 µg (when the Zn content is higher than 2.61). This relation describes less than 2% of cases. Considering the next two regression equations describing the change of Pb and Zn content in flowers (Fig. 7.) and leaves (Fig. 8.) we can see that they describe a number of cases which are insignificant from the statistical point of view: flowers:  $Pb = - 6E - 05 * Zn + 2.95$ ;  $R^2 = 5E - 06$ ; leaves:  $Pb = 0.0038 * Zn + 3.37$ ;  $R^2 = 0.047$ .

The calculated Pearson correlation coefficients (root: 0.220; stalk: 0.134; flower: -0.002; leaf: 0.217) indicate a low level of relation between the considered variables. The considerations therefore confirm that the competition between Pb and Zn is not a permanent phenomenon, but is likely to be observed in some morphological parts of the selected plant species. Confrontations of intercepts including the regression equations for the root, stem, flowers and leaves are considered helpful in explaining physiological levels of a given element and the various interactions between metals.

The above considerations are confirmed by Aery and Rana (Aery and Rana 2007), who while studying the interactions between Zn and Pb in barley also observed antagonistic reactions between these metals, but only at a specific (in this case higher), range of concentrations of these metals. A similar relation has been noted by Symeonidis and Karataglis (Symeonidis and Karataglis 1992) in the studies of interactions between the metals in *Holcus lanatus* L. The antagonistic effect of Zn on Pb was also observed in *Faba vulgaris* Moench (Bystrická et al. 2013). Reduction of lead content to  $0.61 \pm 0.17$  µg/g was due to the presence of zinc in an amount of 80 µg/g. According to the authors of the cited works (Khan et al. 2013), the response of plants in such a situation is sometimes individual, and the decrease in lead content in the presence of zinc (with a different range of concentrations of these metals), should be treated as a feature specific to a given species.

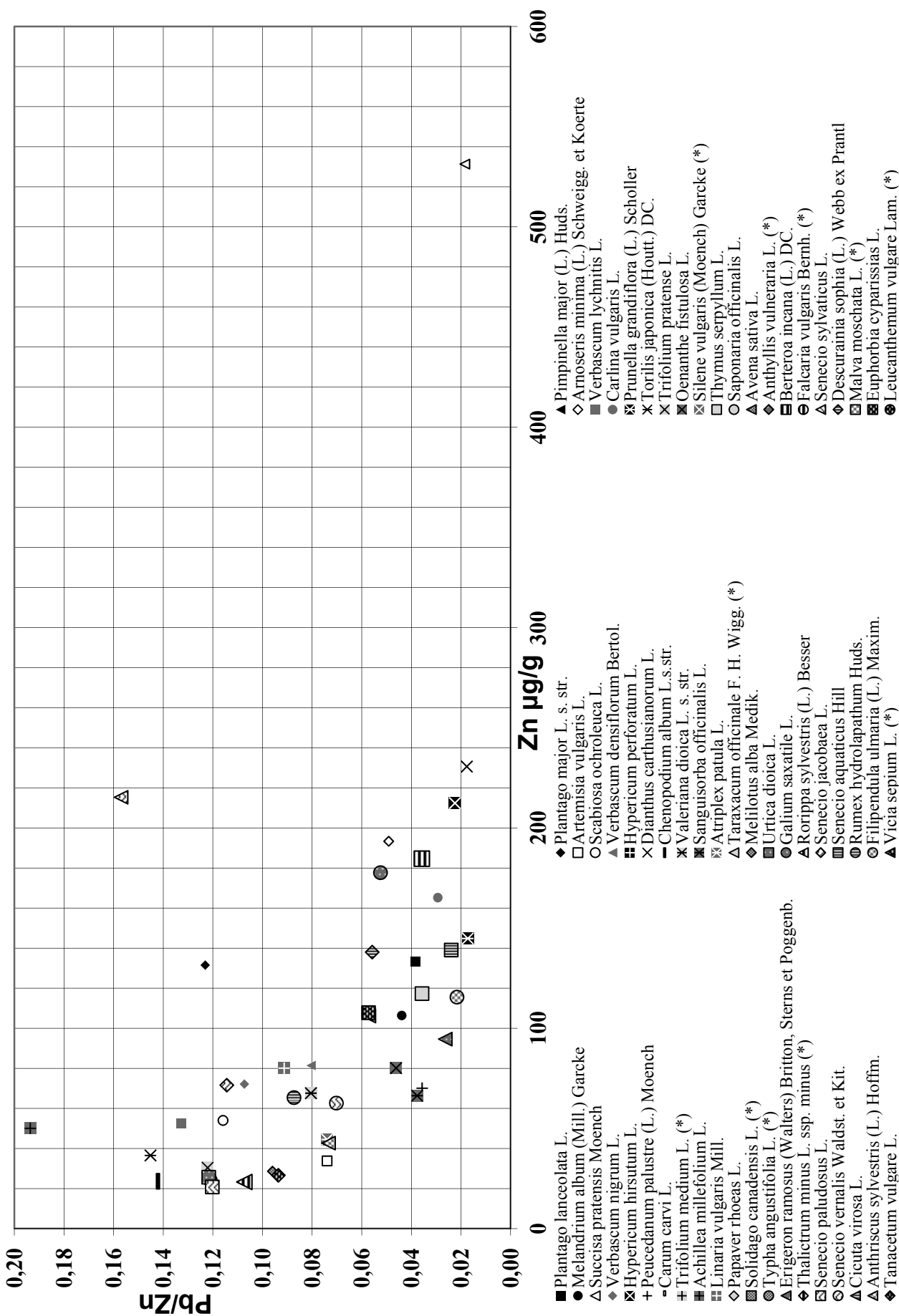


Fig. 1. Antagonism between Pb-Zn ions in the roots of selected plant species. (\*) – no data

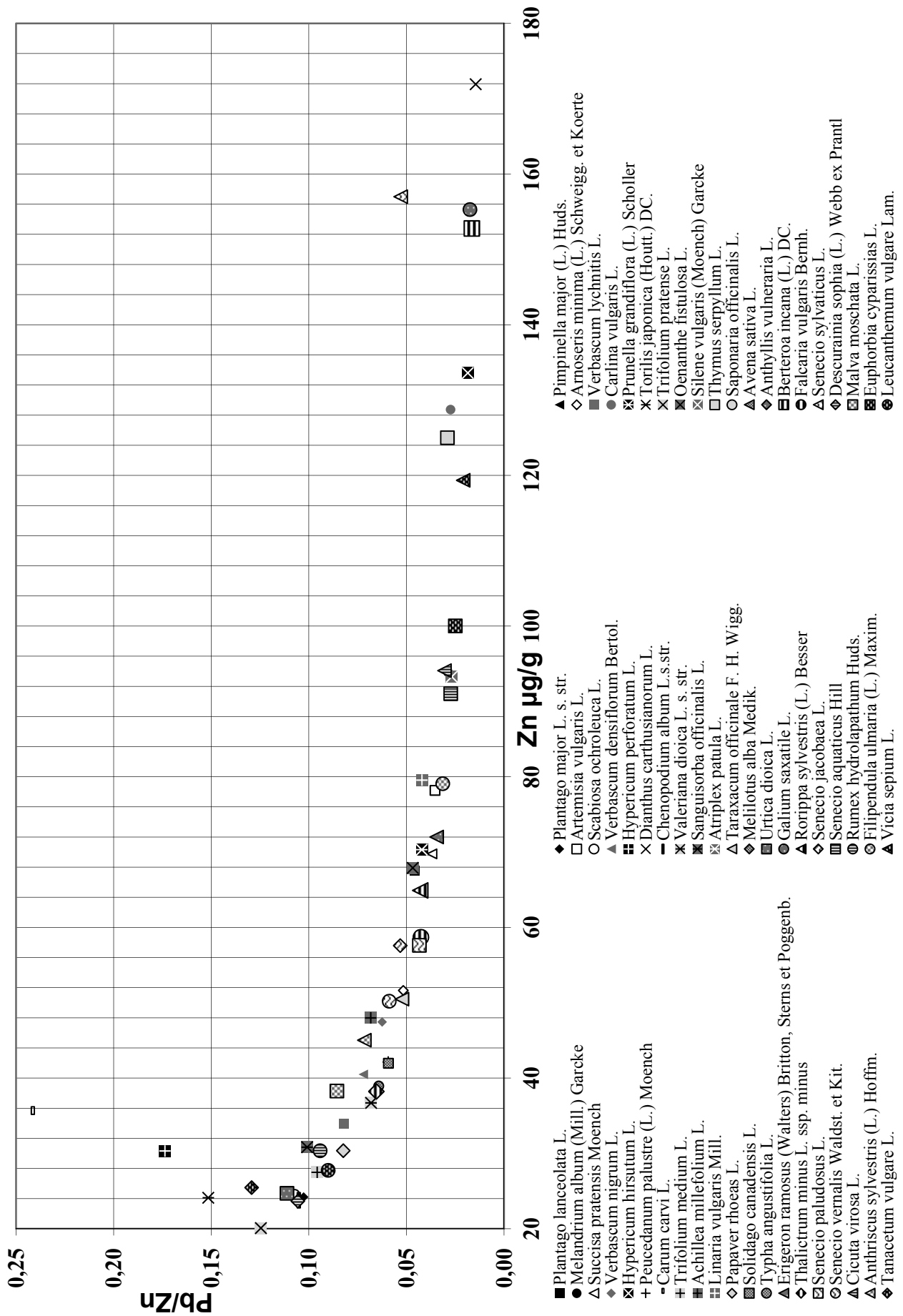


Fig. 2. Antagonism between Pb-Zn ions in the stems of selected plant species. (\*) – no data



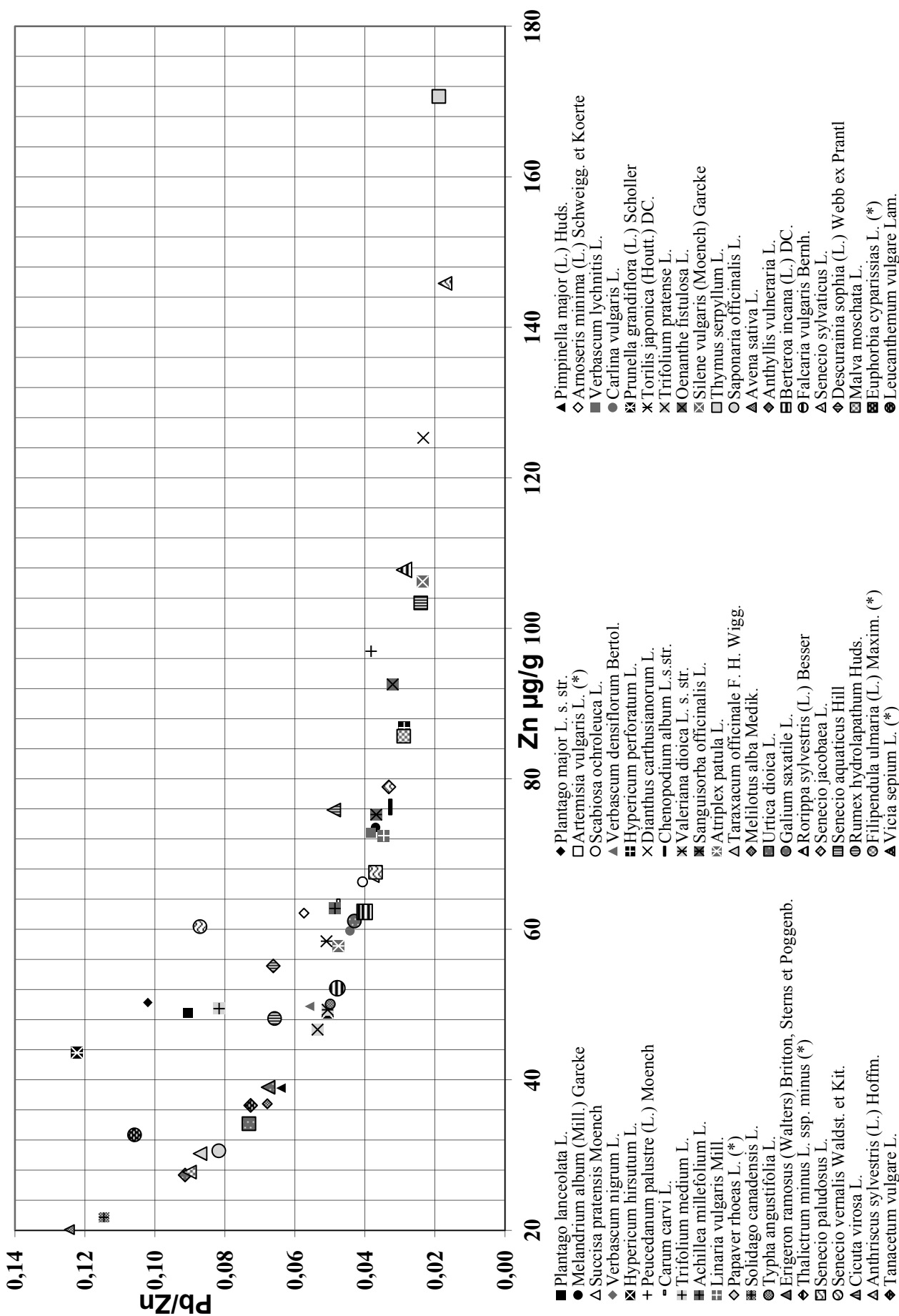


Fig. 3. Antagonism between Pb-Zn ions in the flowers of selected plant species. (\*) – no data

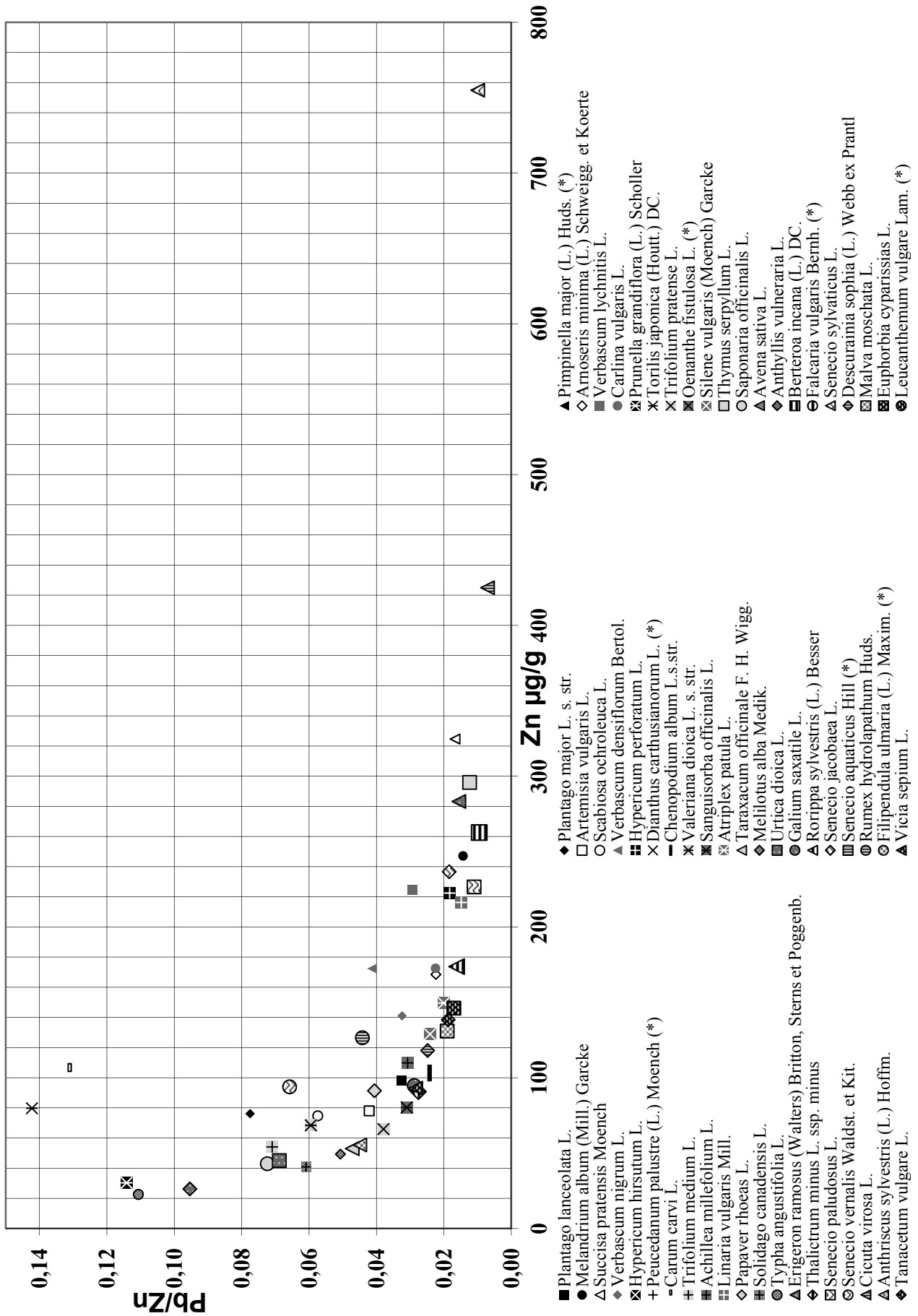
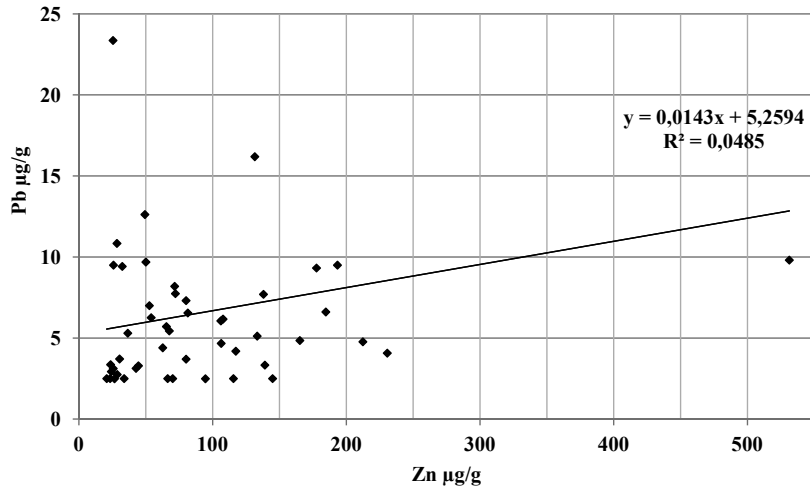
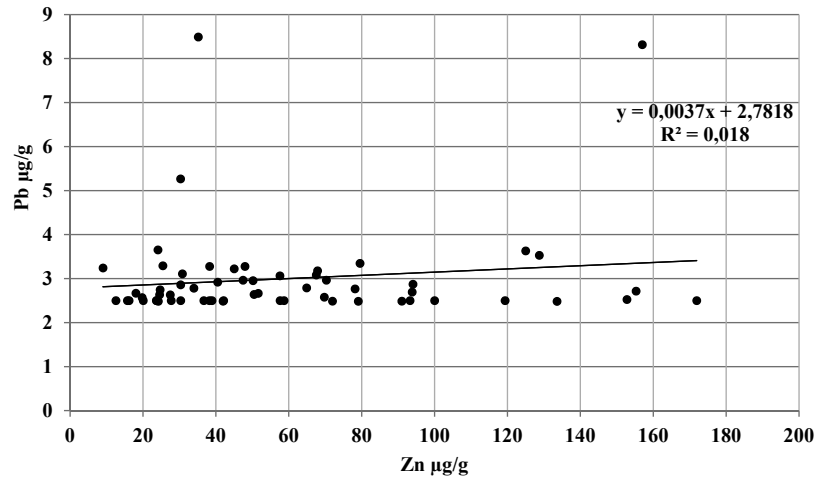


Fig. 4. Antagonism between Pb-Zn ions in the leaves of selected plant species. (\*) – no data



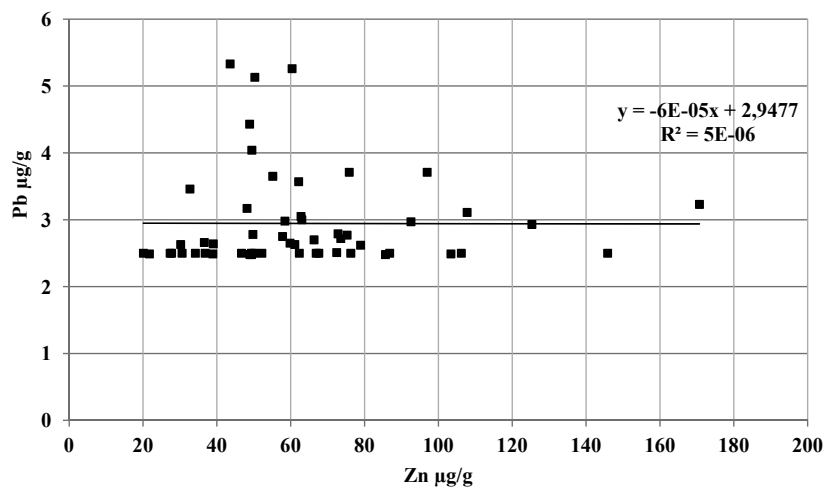
◆ ROOTS

Fig. 5. Simple linear regression equation showing the relationships between Pb and Zn in roots (n=49)



● STEMS

Fig. 6. Simple linear regression equation showing the relationships between Pb and Zn in stems (n=60)



■ FLOWERS

Fig. 7. Simple linear regression equation showing the relationships between Pb and Zn in flowers (n=54)

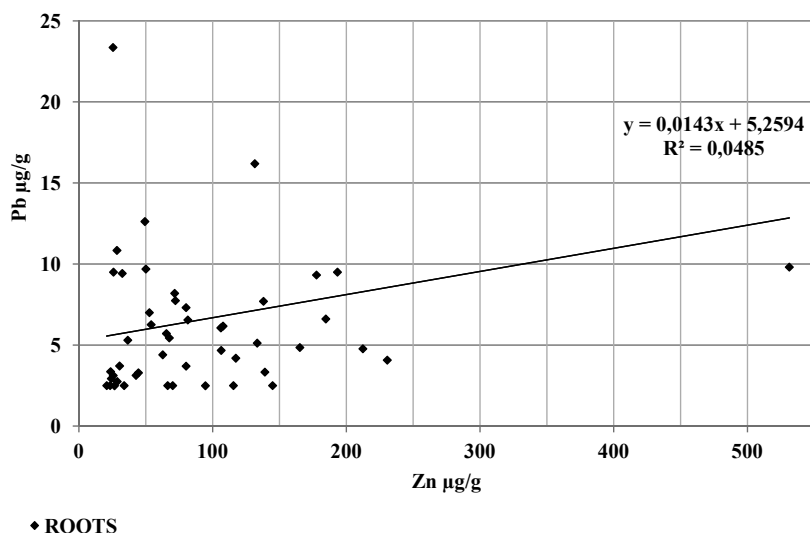


Fig. 8. Simple linear regression equation showing the relationships between Pb and Zn in leaves (n=52)

## Conclusions

1. Antagonism between lead and zinc occurs in all organs of most investigated plant species (60).
2. It was confirmed that the antagonism between lead and zinc in plants occurs in some range of concentrations of the metals, characteristic for a given species.
3. The so-called Zn limits above which the antagonism of lead occurs were determined for each plant species.
4. The on-going observations show that Carthusian Pink (*Dianthus carthusianorum* L.), wild thyme (*Thymus serpyllum* L. emend. Fr.), Woodland Ragwort (*Senecio sylvaticus* L.), Devil's-bit Scabious (*Succisa pratensis* Moench.) and hairy St John's wort (*Hypericum hirsutum* L.) are the species in which the strongest antagonism between lead and zinc occurs.

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## Antagonizm pomiędzy jonami ołowiu i cynku w roślinach

**Streszczenie:** W artykule przedstawiono wyniki badań opisujące występowanie antagonizmu pomiędzy Pb-Zn w wybranych gatunkach roślin z terenu Częstochowy – dzielnica Mirów (północno-zachodnia część Wyżyny Częstochowskiej). Analizowano zmiany ilorazu zawartości Pb/Zn w poszczególnych organach badanych roślin jako funkcję zmian zawartości Zn. Zawartość metali w roślinach i w glebie została określona metodą atomowej spektrofotometrii absorpcyjnej AAS. We wszystkich organach roślin obserwowano antagonistyczny spadek pobierania i kumulacji Pb, będący skutkiem wzrostu koncentracji Zn.

Antagonizm między Zn i Pb w korzeniach badanych roślin występował przy zawartości Zn 200–600  $\mu\text{g/g}$ . Z kolei w łodygach i w kwiatach, antagonizm występował przy niższych zawartościach cynku (100–180  $\mu\text{g/g}$ ). W liściach antagonizm pomiędzy Pb i Zn pojawiał się w przypadku obecności Zn na poziomie zawartości 300–800  $\mu\text{g/g}$ .

Szczegółowe wyniki analiz potwierdzają obecność antagonizmu ołowiu w niektórych roślinach, w odniesieniu do dużych zawartości cynku. Badania potwierdziły także, że stopień antagonizmu zależy od gatunku rośliny.