

Antimicrobial materials properties based on ion-exchange 4A zeolite derivatives

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Zeolites are nanoporous alumina silicates in a framework with cations, exhibiting ion-exchange properties with metal ions making them possible antimicrobial materials. The aim of this study was to evaluate the antimicrobial activity of ion-exchanged zeolites and the toxic potential of these materials. Zeolite-Co²⁺ and Li⁺ exhibited the most effective inhibition on *Staphylococcus aureus* growth than in other microorganisms (*Escherichia coli* and *Pseudomonas aeruginosa*) in low concentrations. Zeolite-Cu²⁺ presented higher zone of inhibition when tested against *Candida albicans*, while Zeolite-Zn²⁺ showed similar effectiveness among all the microorganisms. When ion-exchanged zeolites were used in effective concentrations to achieve antimicrobial activity, no alterations against bioindicators organisms as *Artemia* sp. and *L. sativa* were found and, in addition, they have non-significant result in terms of DNA cleavage activity. Zeolites have advantage of releasing slowly the metals loaded and this characteristic can to be considered promising as potential antimicrobial materials in concentrations safe for use.

Keywords: antimicrobial, zeolite, ion-exchange, bacteria, yeast.

INTRODUCTION

The occurrence of adverse effects caused by the use of conventional antimicrobial agents, which ones have developed resistance to antibiotics and disinfectants, encourage the emergence and improvement of new materials with antimicrobial properties. New types of safe and cost-effective antimicrobial materials have been of great interest due to their several applications, especially in the area of medical devices and food packing and storage^{1,2}. For this reason, the development of antimicrobial packaging systems is a crucial strategy for improving the food shelf-life. An example is eugenol-bearing L malic acid-derived or carboxyanhydrides, which showed antimicrobial activity against pathogenic bacteria (*Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Escherichia coli*), making them promising for developments of bioactive and biodegradable polylactic acid based materials³. Moreover, previous studies have already showed that bulk or surface modifications with silver significantly improved the antimicrobial properties of polylactide films. These modified films were particularly efficient against *E. coli*, *Listeria monocytogenes* and *Salmonella typhimurium* bacteria, but were less efficient in the growth inhibition of *S. aureus*⁴. The use of materials that exhibit antimicrobial activity due to the oligodynamic effect can help to reduce, mitigate or even eradicate some of these contaminations and infections, resulting in improving public health⁵.

Zeolites are very important in the development of functional materials and are considered a promising substrate due to their unique features such as high ion exchange capacity, high surface area, hydrophilicity and easily tunable chemical properties⁶⁻⁸. Zeolites are alumina silicates composed of SiO₄ and AlO₄ tetrahedral structure, in which the anionic characteristic is counteracted by cations outside the molecule. It is possible to obtain synthetic zeolites, such as 4A, from kaolin; a mineral

clay characterized as one hydrated aluminium silicate with a composition similar to Al₂Si₂O₅(OH)₄⁹.

The characteristic composition of 4A zeolites is Na₁₂[Al₁₂Si₁₂O₄₈]27H₂O and it allows cationic exchange procedures. Synthetic and natural zeolites have also been targeted for antibacterial use by cation exchange process, especially after ion-exchange with metals such as silver, zinc, copper and others^{2,10-12}. Antibacterial metals loaded onto zeolites are released slowly and act as inorganic bactericide and disinfectants, which are excellent in terms of safety and thermal stability when compared to organic ones¹³.

Considering that antimicrobial metals show important advantages over traditionally used organic agents and that the zeolites have been considered a promising metal reservoir, the interest of this study was to evaluate the 4A zeolite exchanged with several ions as Cu²⁺, Zn²⁺, Co²⁺, Li⁺, La³⁺ and Ce³⁺ as antimicrobial agents against three bacteria including *S. aureus*, *E. coli* and *P. aeruginosa* and the yeast *Candida albicans*.

These microorganisms were selected due to the fact that they are existent in several environments and when present in food, water or medical materials, can compromise the quality of the product as well the human health. In addition, no study has examined the antimicrobial effect of 4A zeolite exchanged with ions such as Co²⁺, Li⁺, La³⁺ and Ce³⁺ as proposed by this study.

In addition, the study evaluated the toxic potential of these materials within the effective concentrations to antimicrobial activity against bioindicators organisms as *Artemia* sp. and *L. sativa* and also on plasmid DNA cleavage. Some metals used in the ion-exchanged zeolites are essential to life and play important role in the human metabolic system, but it can harm the living organisms in excessive level. Therefore, it is important to investigate the toxic effects of these compounds through toxicity assays.

MATERIALS AND METHODS

Chemicals

The NaNO_3 , CuCl_2 , ZnCl_2 , LaCl_3 , $\text{LiOH}\cdot\text{H}_2\text{O}$, $\text{CoSO}_4\cdot 7\text{H}_2\text{O}$, $\text{CeCl}_3\cdot 7\text{H}_2\text{O}$ used for ion-exchange process, were purchased from Vetec (Duque de Caxias, RJ, Brazil). Agarose and ethidium bromide (EB) from Sigma Aldrich Chemicals (St. Louis, MO, USA). The culture medium plate count agar (PCA), Luria Bertani (LB), Sabouraud dextrose agar (SDA) and brain heart infusion (BHI) were supplied by Himedia Laboratories (Bhaveshwar Plaza, Mumbai, India). The synthetic sea salt from Norsal (São Paulo, SP, Brazil). *Artemia* sp. and *L. sativa* were acquired from Bio Pet Food (Guarulhos, SP, Brazil) and Feltrin Sementes Ltda. (Farroupilha, RS, Brazil), respectively. All were purchased from commercial sources and were analytical grade. The pBSK-II plasmid DNA, used for cleavage assays, was purchased from Stratagene (San Diego, California, EUA).

S. aureus CCCD S007, *E. coli* CCCD E003, *P. aeruginosa* CCCD P003 and *C. albicans* CCCD CC001 strains were obtained from Cefar (São Paulo, SP, Brazil).

Ion-Exchange of 4A Zeolite

As a base material, previously synthesized 4A zeolite (100 g) was employed from commercial kaolin⁹. The cations used for ion-exchange (Cu^{2+} , Zn^{2+} , Co^{2+} , Li^+ , La^{3+} and Ce^{3+}) were chosen because of their antimicrobial characteristics^{14,15}. For each ion, the maximum exchange rate was 24% of the sodium present in the zeolite. Sodium nitrate (NaNO_3) was used as a carrier for the Cu^{2+} , Zn^{2+} , Li^+ and La^{3+} ions in the formulations presented in Table 1. The Co^{2+} and Ce^{3+} ions did not require a carrier once the amount of reactants exceeded the zeolite mass, and the fusion temperature of these ions is lower than that of NaNO_3 (581 K). The mixtures were homogenized and heated for 2 h at the temperatures listed in Table 1. After cooling to room temperature, the mixtures were washed with deionized water (1 L) and finally dried.

The zeolites were previously characterized¹⁶ by X-ray fluorescence (XRF, APW 2400 Philips) and atomic absorption spectrophotometry (AAS, AA240FS Fast Sequential). For X-ray diffraction (XRD), a Shimadzu XRD 6000 equipped with a copper tube was used with a current of 30 mA and potential difference of 30 kV. The phases were identified by comparing the collected full-set diffraction patterns maintained by the *Joint Committee on Powder Diffraction Standards* (JCPDS). For the FTIR analyses, a Shimadzu IR Prestige 21 was used. The tablets were prepared in KBr with the zeolite at concentration of 1%, and spectra were obtained over the range of 4000 to 400 cm^{-1} . The scanning electron microscopy (SEM) image was obtained using Zeiss equipment, with a current of 30 mA and potential difference of 30 kV.

Microbiological assay

Agar diffusion test

The modified disc diffusion method was performed according to the recommendation of Clinical and Laboratory Standards Institute (CLSI) methodology, approved standards M02-A10^{17,18}. The inoculum for the agar diffusion method was prepared with growing *S. aureus*, *E. coli*, *P. aeruginosa* and *C. albicans*, separately, to the turbidity standards. Therefore, five well-isolated colonies of the same morphology type were selected from a PCA (used to grow bacteria) and SDA (used to grow yeast) plate culture grown by 24 to 48 h and transfer into a tube containing 5 mL of saline. The suspension was adjusted until reach the turbidity standard (measured by absorbance at 625 nm) from 0.08 to 0.13 (10^8 CFU/mL) for the microorganism, equivalent to a 0.5 MacFarland standard. Within 15 min after adjusting the turbidity of the inoculum suspension, a sterile cotton swab was dipped into the suspension and spread on the entire sterile agar surface using the same media culture for each microorganism. The medium surface was allowed to dry and the six materials (4A zeolite with Cu^{2+} , Zn^{2+} , Co^{2+} , Li^+ , La^{3+} , Ce^{3+}) were tested, as well as a negative control consisting of 4A zeolite without ion exchange (55.5 mg each). All plates were incubated 24 h at 310 K and 300 K for bacterial and yeast, respectively. After incubation, the presence of the growth inhibition zone around the antimicrobial materials samples was observed and its diameter was measured in millimeters. All experiments were performed in triplicate.

Microorganism cell survival assay

The survival of the microorganisms was observed in different concentrations of the materials tested through of growth inhibition. The survival rate was determined in broth culture containing the antimicrobial materials^{18,19}. The materials, 4A zeolite (negative control) and ion-exchanged ones (Cu^{2+} , Zn^{2+} , Co^{2+} , Li^+ , La^{3+} and Ce^{3+}) were then tested in triplicate at concentrations of 166.6, 96.2, 78.5, 55.5, 32.1, 26.2, 18.5, 10.7, 6.2, 3.6, 2.1, 1.2 and 0.7 mg/mL in a total volume of 3 mL. In the sequence, 300 μL of microorganisms at a concentration of 10^5 CFU/mL were inoculated in 2.7 mL of LB medium for bacteria growth and in BHI medium for yeast growth. Subsequently, the 4A zeolite and ion-exchanged zeolites were added, and the mixtures incubated for 24 h at 310 K and 300 K, respectively. After this, 50 μL aliquots of each sample were applied on PCA or SDA and incubated again for 24 h at 310 K and 300 K for bacterial and yeast growth, respectively. Finally, the number of colonies was counted and the average CFU per mL was determined.

Table 1. Composition of the mixtures used to ion exchange procedure in the 4A zeolite samples

Ion	Reagent	Mass, g	Vehicle NaNO_3 , g	Mass of zeolite, g	Temperature, K
Cu^{2+}	CuCl_2	35.0	70.0	100.0	603
Zn^{2+}	ZnCl_2	30.0	70.0	100.0	603
Co^{2+}	$\text{CoSO}_4\cdot 7\text{H}_2\text{O}$	119.0	–	100.0	383
Li^+	$\text{LiOH}\cdot\text{H}_2\text{O}$	63.2	50.0	100.0	773
La^{3+}	LaCl_3	53.0	50.0	100.0	603
Ce^{3+}	$\text{CeCl}_3\cdot 7\text{H}_2\text{O}$	124.0	–	100.0	373

Toxicological assays

Toxicity assay with *Artemia* sp.

The evaluation of cytotoxicity of the zeolites samples in *Artemia* sp. was performed according to previous methods^{20,21}. A solution of salt water was prepared with synthetic sea salt (30 g/L) and used as the incubation medium for the cysts of *Artemia* sp. Young individuals (n = 10) were exposed 4A zeolite (negative control) and ion-exchanged with Cu²⁺, Zn²⁺, Co²⁺ and Li⁺ in multi-well plates with 2 mL at concentrations of 166.6, 96.2, 78.5, 32.1, 18.5, 10.7, 6.2, 3.6, 2.1, 1.2 mg/mL in four replicates. After 24 h of exposure, the number of dead organisms was observed and noted and the results were expressed as mean ± standard deviation.

Lactuca sativa seed germination

The germination of *L. sativa* seeds assay was performed according to Charles et al. (2011)²² with minor modifications. Seeds (n = 10) were disposed on germination paper soaked with 2 mL of the 4A zeolite and ion-exchanged with Cu²⁺, Zn²⁺, Co²⁺ and Li⁺ in a Petri dish (90 mm) at 295 K in the dark. The treatments were arranged in a completely random design with three replications for each concentration (166.6, 96.2, 78.5, 32.1, 18.5, 10.7, 6.2, 3.6, 2.1, 1.2 mg/mL). Percentage of germinated seeds and the hypocotyl and radicle (root) growth were determined 72 h after initial exposure and then compared to the control group (without zeolite)²². Results were expressed as mean ± standard deviation.

Plasmid DNA cleavage assay

To assess the nuclease activity of the zeolites, plasmid DNA cleavage assay was performed according to Netto et al. (2013)²³ with adaptations. The concentrations of 0 (negative control), 12.5, 25, and 50% of the stock solutions of 4A zeolite (166 mg/mL) and ion-exchanged with Cu²⁺ (32 mg/mL), Zn²⁺ (166.6 mg/mL), Co²⁺ (18.5 mg/mL), Li⁺ (18.05 mg/mL) were incubated in 20 µL of a water solution containing pBSK-II plasmid DNA (300 ng) and 10 mM of HEPES buffer (pH 7.4) for 16 h at 310 K. Subsequently, these solutions were loaded onto 1% agarose gels containing ethidium bromide (EB), and after electrophoresis, bands corresponding to supercoiled form (uncleaved – FI), open circular form

(single strand cleaved – II) and the linear form (double strand cleaved – FIII) were photo documented when present. Fluorescence intensity of EB stained bands was measured with Gel Analyser® free version program. The experiments were performed in triplicate and the results were expressed as mean ± standard deviation.

Statistical analysis

Statistical analyses were performed using Analysis of Variance (ANOVA), followed by Bonferroni post-test in GraphPad Prism 5.0 software. The results are presented as the mean and the standard deviation (SD) and the *p* values <0.05 were considered statistically significant.

RESULTS AND DISCUSSION

4A Zeolite analysis

The results obtained from the XRF analysis are presented in Table 2. The sodium content (Na₂O) was 24.3% for the 4A zeolite (without ion-exchange), and this was used as a parameter for calculating the amounts of ions involved in the ion-exchange procedures. As shown in the same table, the substitution of the sodium content was almost complete for all ions used, with only two exceptions: Zn²⁺ and La³⁺.

The adsorption selectivity of cations is greatly influenced by pore size and specific hydrophilic capacity of the zeolites. If the volume of coordination sphere of hydrated cations to be added prevents the entry into the porous system, which may have occurred with the Zn²⁺ and La³⁺ ions, it is possible to adopt the ion exchange in solid state as an alternative.

In this case, the trituration of the transition metal salt with the dehydrated zeolite, followed by a heat treatment is sufficient for the occurrence of the ion-exchange process. Therefore, the ion-exchange in solid state when compared with the conventional method (i.e. in solution) is one alternative to introduce metal cations in zeolites, when the volume of the cation salvation sphere interferes in the ion-exchange process in solution²⁴.

However, this method may be less efficient when compared to the conventional method, resulting in a lower exchange between these cations in substitution for sodium.

In Table 2 can also be observed the loss to fire. This loss is related to the content of water in 4A zeolite,

Table 2. XRF and AAS of 4A zeolite (Z4A) before and after ion-exchange

Element, % mass	Z4A	Z4A+Cu ²⁺	Z4A+Zn ²⁺	Z4A+Co ²⁺	Z4A+Li ⁺	Z4A+La ³⁺	Z4A+Ce ³⁺
Al ₂ O ₃	28.2	23.38	22.3	25.53	32.54	21.97	28.60
CaO	0.07	<0.05	<0.05	<0.05	0.30	0.52	<0.05
Fe ₂ O ₃	<0.01	0.18	0.21	0.24	0.32	0.18	0.16
K ₂ O	0.01	0.25	0.27	0.27	0.19	0.28	0.25
MgO	<0.01	<0.05	<0.05	0.11	0.19	<0.05	<0.05
MnO	<0.01	<0.05	<0.05	0.07	<0.05	<0.05	<0.05
Na ₂ O	24.30	4.35	10.39	3.80	3.82	16.06	1.84
P ₂ O ₅	<0.01	<0.01	<0.05	<0.01	<0.05	<0.05	<0.05
SiO ₂	31.7	27.36	29.16	31.7	33.54	28.80	35.3
TiO ₂	<0.01	<0.01	<0.05	<0.01	<0.05	<0.05	<0.05
CuO	0.00	22.83	0.00	0.00	0.00	0.00	0.00
ZnO	0.00	0.00	17.61	0.00	0.00	0.00	0.00
LaO	0.00	0.00	0.00	0.00	0.00	10.47	0.00
Li ₂ O	0.00	0.00	0.00	0.00	15.80	0.00	0.00
Co ₂ O ₃	0.00	0.00	0.00	15.8	0.00	0.00	0.00
CeO	0.00	0.00	0.00	0.00	0.00	0.00	16.6
Loss fire	13.30	17.54	19.93	16.80	11.59	20.15	17.55

which in stoichiometric amount can reach up to 22% in mass. However, this amount depends on the $\text{SiO}_2/\text{Al}_2\text{O}_3$ ratio. Other factors can act in this loss of mass, since it did not occur uniformly, being possible that during the ion exchange the structure of the zeolites may have undergone alteration, resulting in different proportions in the $\text{SiO}_2/\text{Al}_2\text{O}_3$ ratio.

The morphology of 4A zeolite was showed in Figure 1, which presents cubic crystals approximately $2\ \mu\text{m}$, typical of zeolites 4A. The ion-exchanged zeolites were characterized in our previous study¹⁶.

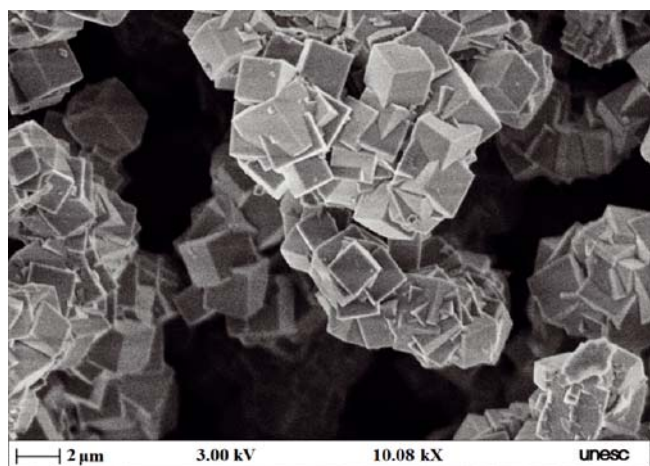


Figure 1. Morphology of 4A zeolite. Scale bar = $2\ \mu\text{m}$. Magnification = 10.08 kX

Antimicrobial activity

The ion-exchange zeolites were evaluated regarding their antimicrobial activity against a broad range of microorganisms including bacteria and yeast. In the present study, the results obtained in the agar diffusion assay are present as average diameter of the materials inhibition zone in Table 3 and Figure 2.

Table 3. Average diameter (mm) of the zone of inhibition promoted by 4A zeolite (Z4A) and exchanged zeolites with Cu^{2+} , Zn^{2+} , Co^{2+} , Li^+ , La^{3+} and Ce^{3+} against microorganisms

Microorganisms	Average diameter, mm						
	Z4A	Cu^{2+}	Zn^{2+}	Co^{2+}	Li^+	La^{3+}	Ce^{3+}
<i>Staphylococcus aureus</i>	0.0	6.0	8.5	12.0	14.5	0.0	0.0
<i>Escherichia coli</i>	0.0	5.0	8.5	9.0	10.0	0.0	0.0
<i>Pseudomonas aeruginosa</i>	0.0	7.0	9.5	11.5	12.5	0.0	0.0
<i>Candida albicans</i>	0.0	10.0	9.0	11.0	11.0	0.0	0.0

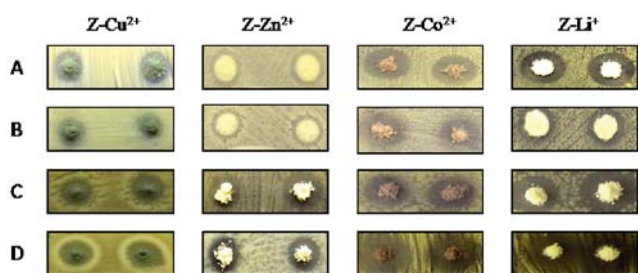


Figure 2. Zone of inhibition around zeolites ion-exchange with Cu^{2+} , Zn^{2+} , Co^{2+} and Li^+ against (A) *S. aureus*, (B) *E. coli*, (C) *P. aeruginosa* and (D) *C. albicans*

The standard control (4A zeolite without ion-exchange) has shown no effect against the microorganisms tested and similar results were findings for the zeolites exchange with La^{3+} and Ce^{3+} .

The zeolite- Cu^{2+} , Zn^{2+} , Co^{2+} and Li^+ showed the best results, where zeolite- Zn^{2+} inhibited similarly the bacteria and yeast tested in this study. Zeolite- Cu^{2+} presented higher zone of inhibition against *C. albicans*. In contrast, the zeolite- Co^{2+} and Li^+ inhibited the growth of gram positive *S. aureus* relatively higher than others microorganisms.

Only a few studies highlight antifungal effects of ion-exchanged zeolites, especially for toxigenic filamentous fungi. Therefore, in previous study performed by Savi et al.¹⁴ the ion-exchanged zeolites were evaluated against filamentous and toxigenic fungi: *Aspergillus flavus*, where zeolite with Li^+ and Cu^{2+} showed the best antifungal activity. In that study was also performed the antimycotoxin activity test where all zeolite samples (4A zeolite, zeolites with Li^+ , Cu^{2+} , Co^{2+} and Zn^{2+}) efficiently inhibited the aflatoxin B₁ (AFB₁) production by *A. flavus*. Even the zeolite that showed no significant antifungal effect, affected the secondary metabolites (AFB₁) production of *A. flavus*, suggesting some interaction with the fungi metabolism. Though the mechanisms of antifungal effects of metal ions in fungi are far from being understood in detail, these basic mechanisms enable us to consider the evaluation of antimicrobial effects on cell metabolism from different microorganism including bacterial and yeast. Their efficiency seems to depend on the sensitivity of each species of microorganisms and the interaction with the antimicrobial used.

Similarly to our results, Tekin et al. (2016) shown that the Zn^{2+} and Cu^{2+} loaded zeolite samples also exhibited antibacterial activity. The zeolite- Zn^{2+} and zeolite- Cu^{2+} showed greater inhibition zone of 13, 15, 10, 20 mm and 17, 20, 17, 14 mm for *E. coli*, *S. aureus*, *P. aeruginosa* and *C. albicans*, respectively¹¹.

Zeolites- Ag^+ were found to be the most effective as antibacterial when compared to zeolites exchanged with

Cu^{2+} and Zn^{2+} according to Demirci et al. (2014). On the other hand, the Cu^{2+} and Zn^{2+} ion-loaded zeolites were found to display more antifungal and anticandidal characteristics. Some results that are corroborating to ours, also showed zeolite- Cu^{2+} is the most effective against *C. albicans*²⁵.

It is important to emphasize that these metal ions can be considered as an alternative to safety and stability on antibacterial activity at low concentrations. Zinc is a vital antioxidant and anti-inflammatory agent in the human body, although a small quantity of Zn^{2+} ion is essential for numerous metabolic activities. Zeolite- Zn^{2+} have been used in biomedical applications due to their wound healing and bactericidal properties²⁶. Similar to zinc, copper is an essential metal for living organisms at low concentrations and are used inorganic antibacterial materials²⁷. In recent study, copper doped zeolite

composite adsorbents showed high efficiency to remove simultaneously heavy metals and total coliforms from wastewater of Akaki river, from central Ethiopia²⁸.

In contrast, the antimicrobial activity zeolite loaded of metals as Co^{2+} and Li^+ are scarce in the literature, although the Co^{2+} has already shown antibacterial activity against growth of the Gram-negative bacteria *Pseudomonas putida* and *E. coli* when evaluated as two cobalt imidazolate metal-organic frameworks²⁹.

The cell survival assays performed are in agreement with the findings of the agar diffusion assay. 4A zeolite had no effect against the microorganisms tested as well the zeolite- La^{3+} and Ce^{3+} . Zeolites exchanged with Cu^{2+} , Li^+ and Co^{2+} eliminated all living cells in some concentrations tested as shown in Figure 3.

The antimicrobial activity of the ion-exchanged zeolites was shown depending on the concentrations used to each microorganism individually. The zeolite- Cu^{2+} inhibited the *P. aeruginosa* in the concentration of 6.2 mg/mL or higher, but the zeolite- Zn^{2+} had no effect against *P. aeruginosa*. In addition, the action was not complete to other microorganism tested, which presented survival of small amounts of cells. The zeolites with the Li^+ and Co^{2+} can be considered the most effective among the six materials tested in the current study, although the ability of Cu^{2+} is also strong and can be considered effective. The zeolite- Li^+ shown antibacterial effect against *P. aeruginosa* and *C. albicans* in the concentration from 6.2 mg/mL while zeolite- Co^{2+} inhibited *S. aureus* from 0.7 mg/mL concentration.

The way ion-exchanged zeolites act as antimicrobial is not yet fully established, mainly by ions as Co^{2+} and Li^+ , which have not been yet examined in the literature. However, it is possible to affirm that the zeolite ion re-

lease observed in previous study³⁰ can get across the cell membrane causing damage to bacteria and fungi^{31,32}. This mechanism is well-known for Cu^{2+} and Zn^{2+} metal ions.

Toxicological studies

Artemia sp. is one of the most valuable test organisms available for several applications, including toxicology and ecotoxicology researches^{20, 33, 34}. The toxicity of chemical compounds with possible biological activity has been tested against *Artemia* sp. due to several advantages such as: well known biology and ecology, low cost of the organisms, as well, speed and convenience of the tests. According to the literature, there is no study involving the evaluation of the toxicity of modified zeolites using this model. However, this microcrustacean is often used as a model organism of toxicity assessment of nanoparticles^{20, 34}, iron metallo-drugs³³, and metal ions of copper and zinc²¹.

As observed in Figure 4, the zeolite ion-exchanged with Cu^{2+} , Zn^{2+} , Co^{2+} and Li^+ only shown toxic effect to *Artemia* sp. in concentrations above of 10.7 mg/mL, 96.2 mg/mL, 18.5 mg/mL and 6.2 mg/mL, respectively. Moreover, no mortality was observed within the ion-exchanged zeolites in effective concentrations used for antimicrobial activity.

In the same sense, the *L. sativa* was used to evaluate the toxicity of the materials. Several studies have reported successful use of *L. sativa* as bioindicator organisms for ecotoxicological evaluation of contaminated environments^{35, 36}. In this work, only the zeolite- Li^+ was capable of causing significant alteration on the germination seeds. The radicle and hypocotyl growth were affected after exposure to almost all the zeolites when used in high concentrations. In the antimicrobial effective con-

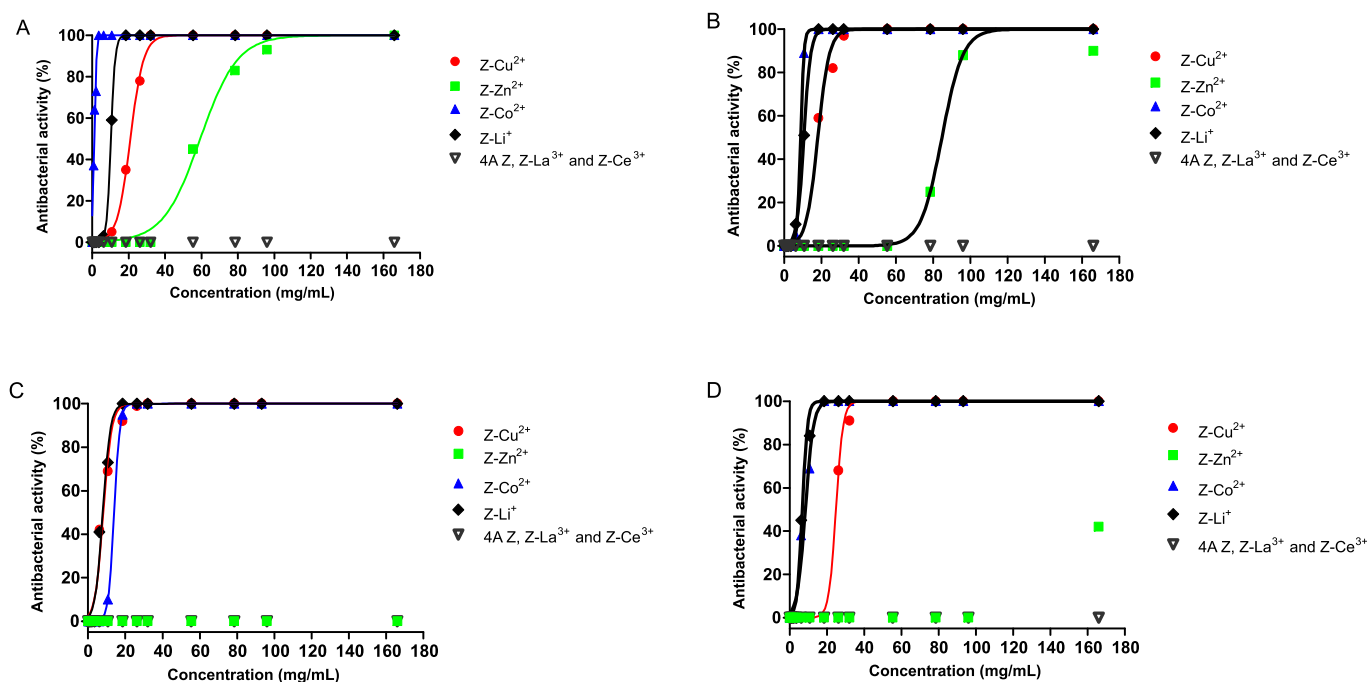


Figure 3. Antibacterial activity (%) of 4A zeolite and zeolites ion-exchange on the bacterial growth: (A) *S. aureus*, (B) *E. coli* (C) *P. aeruginosa* and (D) *C. albicans*

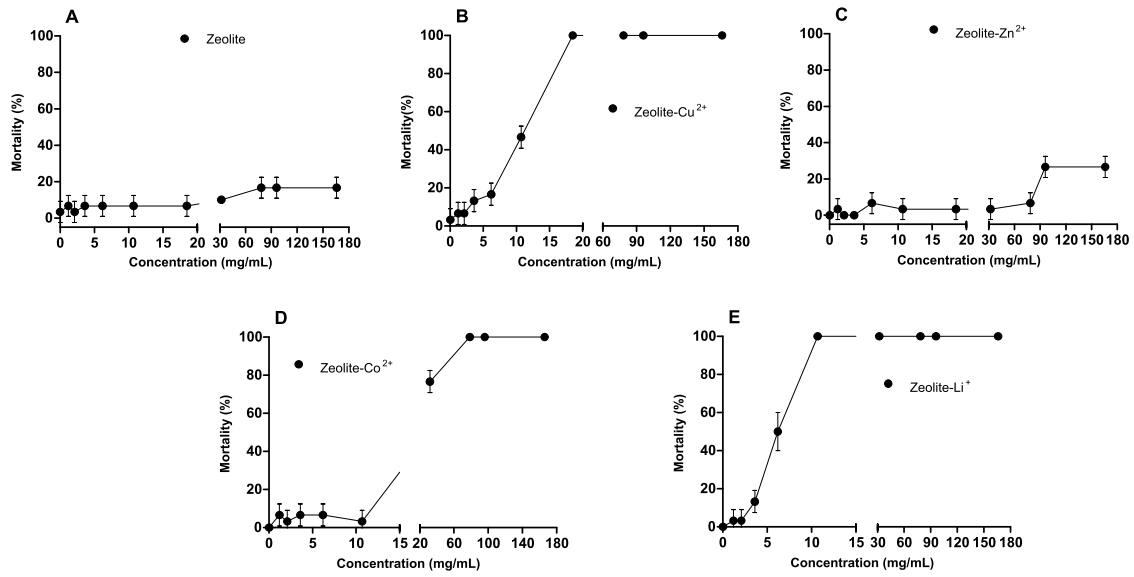
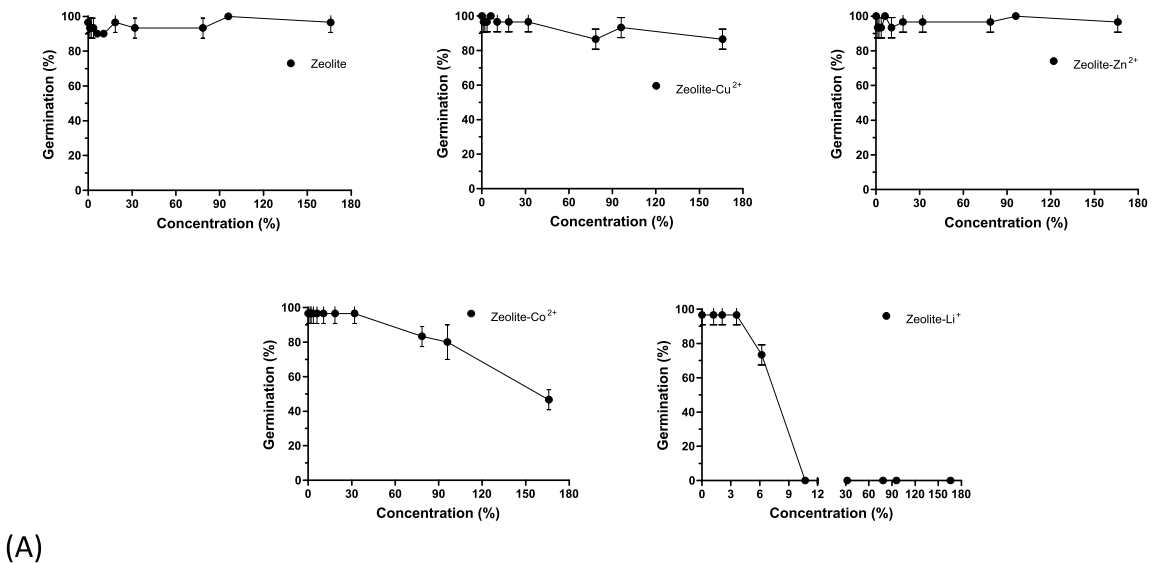
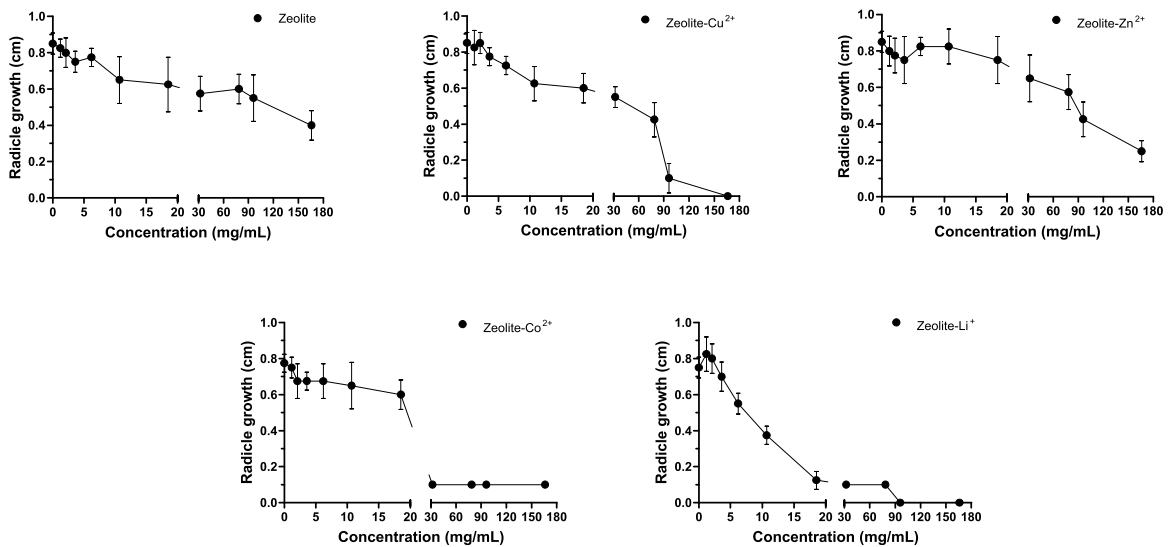


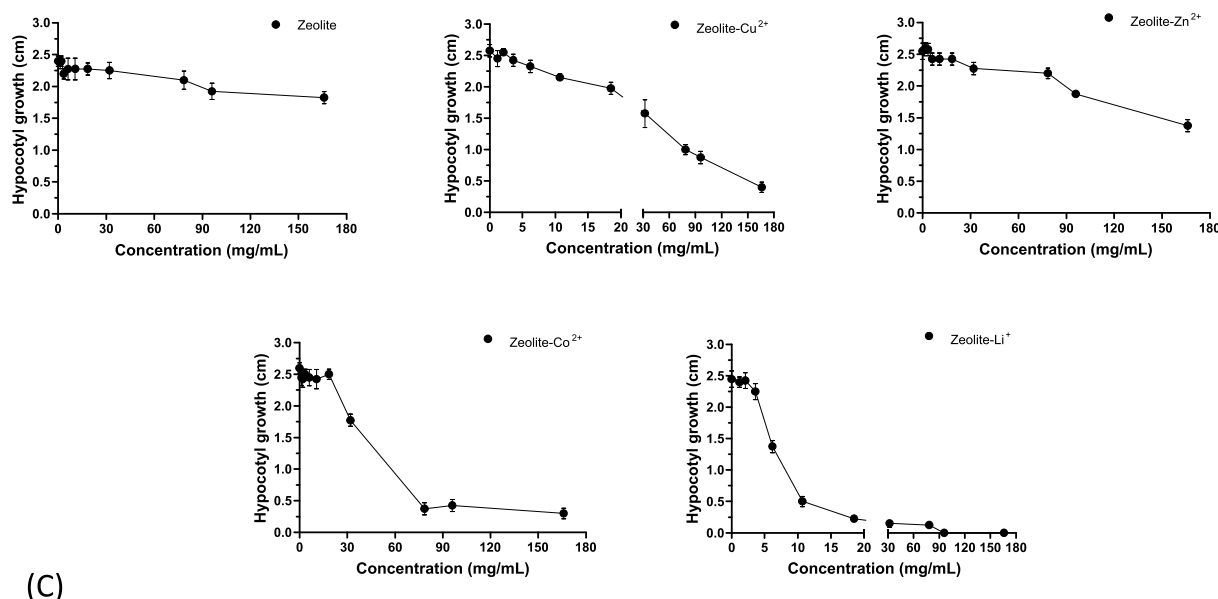
Figure 4. Mortality (%) of *Artemia* sp. after 24 h exposure at 166.6, 96.2, 78.5, 32.1, 18.5, 10.7, 6.2, 3.6, 2.1 and 1.2 mg/mL concentrations of 4A zeolite (A) and its ion-exchanged zeolites with Cu²⁺ (B), Zn²⁺ (C), Co²⁺ (D) and Li⁺ (E)



(A)



(B)



(C)

Figure 5. Sub-chronic toxic effects of the 4A zeolite and ion-exchange with Cu²⁺, Zn²⁺, Co²⁺ and Li⁺ on seeds of *L. sativa* exposed to different concentration in terms of germination (A), radicle growth (B) and hypocotyl growth (C). Data expressed as mean ± SD of three replicates

centrations, the zeolites had presented no toxic effects, with the exception of the zeolite-Li⁺.

Considering to *L. sativa* seeds germination, only zeolite-Co²⁺ and Li⁺ showed reduction in the concentrations from 166.6 and 6.2 mg/mL, respectively (Figure 5A). Even after germination of *L. sativa*, the ions Cu²⁺, Zn²⁺, Co²⁺ and Li⁺ caused radicle (Figure 5B) and hypocotyls (Figure 5C) growth reduction in concentrations equal or higher than 32.1 mg/mL, 96.2 mg/mL, 32.1 mg/mL and 6.2 mg/mL, respectively.

The zeolites presented different result in the plasmid DNA cleavage assay depending on the metal used in their structure as exchange ion. The Figure 6 shows that the 4A zeolite (1A), zeolite-Li⁺ (4A) and Co²⁺

(5A) presented no result or cleavage activity that can be observed. The zeolite-Zn²⁺ (2A) presented a slight activity that was visually observed with the increase of FII amount but not statistically significant. On the other hand, the zeolite-Cu²⁺ (3A) presented a strong activity with statistical significance, but probably not by its own feature but due to the highly redox properties of the copper(II) ion³⁷ in the presence of HEPES buffer which is well-known to act as reducing agent³⁸ leading to the generation of several oxidative species that damages plasmid DNA^{39, 40}. This hypothesis also explains the lack of DNA cleaving activity of the others zeolites, since those ion metals has none or much lower redox properties compared to copper(II).

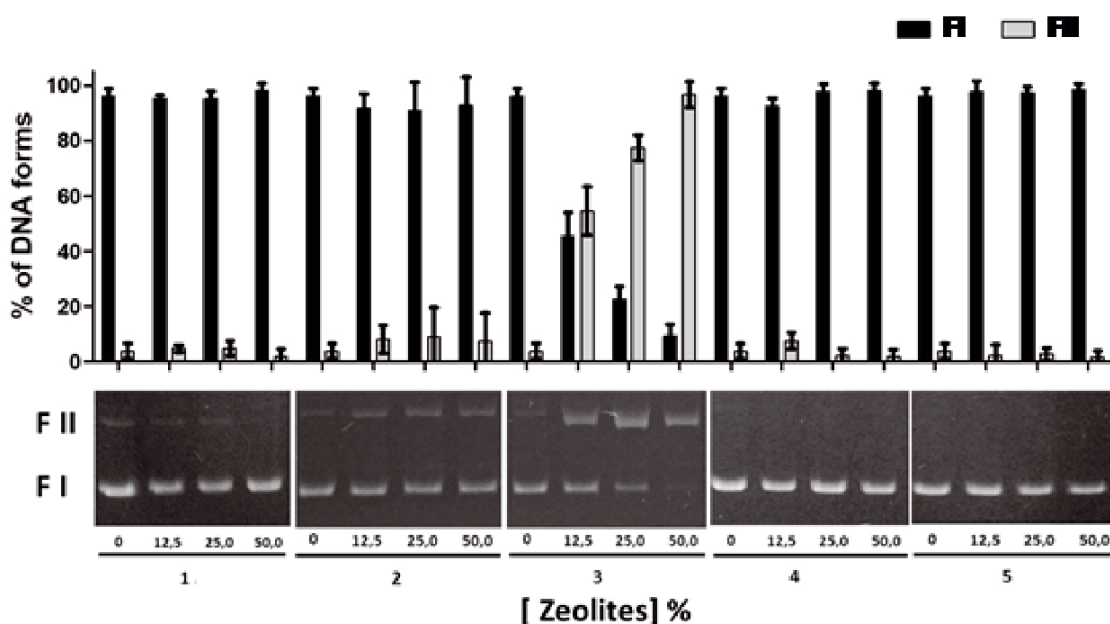


Figure 6. Percentage of DNA forms (FI – uncleaved and FII – single strand cleaved) observed after exposure to different concentrations of (1) 4A zeolite and ion-exchange with (2) Zn²⁺, (3) Cu²⁺, (4) Li⁺ and (5) Co²⁺

CONCLUSION

The current study showed that zeolite-Cu²⁺, Li⁺ and Co²⁺ eliminated all living cells in some concentrations tested and were considered the most effective among six materials tested. In the toxicity assays, they affected the *Artemia* sp. mortality and *L. sativa* germination when tested in high concentrations. In contrast, it is important to emphasize that the ion-exchanged zeolites showed no toxic effect against *Artemia* sp. and *L. sativa* in effective concentrations used for antimicrobial activity. For example, zeolite-Co²⁺ showed strong antibacterial activity against *S. aureus* in low concentration (0.7 mg/mL), which showed no alterations against the bioindicators organism and no significant result on plasmid DNA cleavage activity. The ion-exchanged zeolites can be considered promising in terms of potential antimicrobial materials since they presented antibacterial activity in concentrations that are safe for use. Finally, it is important to highlight the efficiency of some ion-exchanged zeolites used to eliminate specific microorganism and therefore, deserve special attention as antimicrobial agent against resistant microorganisms that cause strong impact to public health.

DISCLOSURE STATEMENT

No potential conflict of interest was reported by the authors.

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LITERATURE CITED

- Boschetto, D.L., Lerin, L., Cansian, R., Pergher, S.B.C. & Di Luccio, M. (2012). Preparation and antimicrobial activity of polyethylene composite films with silver exchanged zeolite-Y. *Chem. Eng. J.* 204, 210–216. <http://dx.doi.org/10.1016/j.cej.2012.07.111>
- Sánchez, M.J., Mauricio, J.E., Paredes, A.R., Gamero, P. & Cortés, D. (2017). Antimicrobial properties of ZSM-5 type zeolite functionalized with silver. *Mater. Lett.* 191, 65–68. <http://dx.doi.org/10.1016/j.matlet.2017.01.039>
- Gazzotti, S., Todisco, S.A., Picozzi, C., Ortenzi, M.A., Farina, H., Lesma, G. & Silvani, A. (2019). Eugenol-grafted aliphatic polyesters: Towards inherently antimicrobial PLA-based materials exploiting OCAs chemistry. *Eur. Polym. J.* 114, 369–379. <https://doi.org/10.1016/j.eurpolymj.2019.03.001>
- Turalija, M., Bischof, S., Budimir, A. & Gaan, S. (2016). Antimicrobial PLA films from environment friendly additives. *Compos. Part B: Engin.* 102, 94–99. <https://doi.org/10.1016/j.compositesb.2016.07.017>
- Braunwarth, H. & Brill, F.H.H. (2014). Antimicrobial efficacy of modern wound dressings: Oligodynamic bactericidal versus hydrophobic adsorption effect. *Wound Medicine.* 5, 16–20. <http://dx.doi.org/10.1016/j.wndm.2014.04.003>
- Breck, D.W. (1984). *Zeolite molecular sieves: structure, chemistry, and use* ed (Universidade de Michigan), pp. 771.
- McCusker, L.B., Olson, D.H. & Baerlocher, C. (2007). *Atlas of Zeolite Framework Types 6^a* ed (Elsevier Science). ISBN: 978-0-444-53064-6.
- Kulprathipanja, S. (2010). *Zeolites in Industrial Separation and Catalysis.* pp. 618. Wiley.
- Melo, C.R., Riella, H.G., Kuhnen, N.C., Angioletto, E., Melo, A.R., Bernardin, A.M., da Rocha, M.R. & da Silva, L. (2012). Synthesis of 4A zeolites from kaolin for obtaining 5A zeolites through ionic exchange for adsorption of arsenic. *Mater. Sci. Engin., B.* 177(4), 345–349. <http://dx.doi.org/10.1016/j.mseb.2012.01.015>
- Rivera-Garza, M., Olguín, M.T., García-Sosa, I., Alcántara, D. & Rodríguez-Fuentes, G. (2000). Silver supported on natural Mexican zeolite as an antibacterial material. *Micropor. Mesopor. Mat.* 39(3), 431–444. [http://dx.doi.org/10.1016/S1387-1811\(00\)00217-1](http://dx.doi.org/10.1016/S1387-1811(00)00217-1)
- Tekin, R. & Bac, N. (2016). Antimicrobial behavior of ion-exchanged zeolite X containing fragrance. *Micropor. Mesopor. Mat.* 234, 55–60. <http://dx.doi.org/10.1016/j.micromeso.2016.07.006>
- Ferreira, L., Fonseca, A.M., Botelho, G., Aguiar, C.A. & Neves, I.C. (2012). Antimicrobial activity of faujasite zeolites doped with silver. *Micropor. Mesopor. Mat.* 160, 126–132. <http://dx.doi.org/10.1016/j.micromeso.2012.05.006>
- Fang, M., Chen, J.H., Xu, X.L., Yang, P.H. & Hildebrand, H.F. (2006). Antibacterial activities of inorganic agents on six bacteria associated with oral infections by two susceptibility tests. *Int. J. Antimicrob. Agents.* 27(6), 513–517. <http://dx.doi.org/10.1016/j.ijantimicag.2006.01.008>
- Kalinowska, M., Piekut, J., Bruss, A., Follet, C., Sienkiewicz-Gromiuk, J., Świsłocka, R., Rzączyńska, Z. & Lewandowski, W. (2014). Spectroscopic (FT-IR, FT-Raman, ¹H, ¹³C NMR, UV/VIS), thermogravimetric and antimicrobial studies of Ca(II), Mn(II), Cu(II), Zn(II) and Cd(II) complexes of ferulic acid. *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* 122, 631–638. <http://dx.doi.org/10.1016/j.saa.2013.11.089>
- Zhang, B., Lin, Y., Tang, X., He, S. & Xie, G. (2010). Synthesis, characterization, and antimicrobial properties of Cu-inorganic antibacterial material containing lanthanum. *J. Rare Earths.* 28, 451–455. [http://dx.doi.org/10.1016/S1002-0721\(10\)60346-8](http://dx.doi.org/10.1016/S1002-0721(10)60346-8)
- Savi, G.D., Cardoso, W.A., Furtado, B.G., Bortolotto, T., Da Agostin, L.O.V., Nones, J., Zanoni, E.T., Montedo, O.R.K. & Angioletto, E. (2017). New ion-exchanged zeolite derivatives: Antifungal and antimycotoxin properties against *Aspergillus flavus* and aflatoxin B₁. *Mat. Res. Exp.*, 4, 085401. <http://dx.doi.org/10.1088/2053-1591/aa84a5>
- CLSI (2009). Clinical and Laboratory Standards Institute. Performance standards for antimicrobial disk susceptibility tests. Pennsylvania, USA., Wayne.
- Balouiri, M., Sadiki, M. & Ibsouda, S.K. (2016). Methods for in vitro evaluating antimicrobial activity: A review. *J. Pharmaceut. Anal.* 6(2), 71–79. <http://dx.doi.org/10.1016/j.jpha.2015.11.005>
- Santos, M.F., Oliveira, C.M., Tachinski, C.T., Fernandes, M.P., Pich, C.T., Angioletto, E., Riella, H.G. & Fiori, M.A. (2011). Bactericidal properties of bentonite treated with Ag⁺ and acid. *Int. J. Miner. Process.* 100(1), 51–53. <http://dx.doi.org/10.1016/j.minpro.2011.04.012>
- Rajabi, S., Ramazani, A., Hamidi, M. & Naji, T. (2015). *Artemia salina* as a model organism in toxicity assessment of nanoparticles. *DARU J. Pharm. Sci.* 23(1), 20. <http://dx.doi.org/10.1186/s40199-015-0105-x>
- Brix, K.V., Gerdes, R.M., Adams, W.J. & Grosell, M. (2006). Effects of copper, cadmium, and zinc on the hatching success of brine shrimp (*Artemia franciscana*). *Arch. Environ. Contam. Toxicol.* 51(4), 580–583. <http://dx.doi.org/10.1007/s00244-005-0244-z>
- Charles, J., Sancey, B., Morin-Crini, N., Badot, P.-M., Degiorgi, F., Trunfio, G. & Crini, G. (2011). Evaluation of the phytotoxicity of polycontaminated industrial effluents using the lettuce plant (*Lactuca sativa*) as a bioindicator. *Ecotoxicol. Environ. Saf.* 74(7), 2057–2064. <https://doi.org/10.1016/j.ecoenv.2011.07.025>
- Netto, E., Madeira, R.A., Silveira, F.Z., Fiori, M.A., Angioletto, E., Pich, C.T. & Geremias, R. (2013). Evaluation of the toxic and genotoxic potential of acid mine drainage using physicochemical parameters and bioassays. *Environ.*

- Toxicol. Pharmacol.* 35(3), 511–516. <http://dx.doi.org/10.1016/j.etap.2013.02.007>
24. Luna, F.J. & Schuchardt, U. (2001). Modificação de zeólitas para uso em catálise. *Quim. Nova.* 24, 885–892.
25. Demirci, S., Ustaoglu, Z., Yilmazer, G.A., Sahin, F. & Bac, N. (2014). Antimicrobial properties of zeolite-X and zeolite-A ion-exchanged with silver, copper, and zinc against a broad range of microorganisms. *Appl. Biochem. Biotechnol.* 172(3), 1652–1662. <http://dx.doi.org/10.1007/s12010-013-0647-7>
26. Tapiero, H. & Tew, K.D. (2003). Trace elements in human physiology and pathology: zinc and metallothioneins. *Biomed. Pharmacother.* 57(9), 399–411.
27. Alswat, A.A., Ahmad, M.B., Hussein, M.Z., Ibrahim, N.A. & Saleh, T.A. (2017). Copper oxide nanoparticles-loaded zeolite and its characteristics and antibacterial activities. *J. Mater. Sci. Technol.* 33(8), 889–96. <http://dx.doi.org/10.1016/j.jmst.2017.03.015>
28. Fanta, F.T., Dubale, A.A., Bebizuh, D.F. & Atlabachew, M. (2019). Copper doped zeolite composite for antimicrobial activity and heavy metal removal from waste water. *BMC Chemistry.* 13(1), 44. <http://dx.doi.org/10.1186/s13065-019-0563-1>
29. Aguado, S., Quirós, J., Canivet, J., Farrusseng, D., Boltes, K. & Rosal, R. (2014). Antimicrobial activity of cobalt imidazole metal-organic frameworks. *Chemosphere.* 113, 188–192. <http://dx.doi.org/10.1016/j.chemosphere.2014.05.029>
30. Savi, G.D., Cardoso, W.A., Furtado, B.G., Bortolotto, T., Zanoni, E.T., Scussel, R., Rezende, L.F., Avila, R.A.M., Montedo, O.R.K. & Angioletto, E. (2018). Antifungal activities against toxigenic *Fusarium* specie and deoxynivalenol adsorption capacity of ion-exchanged zeolites. *J. Environ. Sci. Health, Part B.* 53(3): 184–190. <http://dx.doi.org/10.1080/03601234.2017.1405639>.
31. Tamayo, L., Azócar, M., Kogan, M., Riveros, A. & Páez, M. (2016). Copper-polymer nanocomposites: An excellent and cost-effective biocide for use on antibacterial surfaces. *Mater. Sci. Engin.: C.* 69, 1391–1409. <http://dx.doi.org/10.1016/j.msec.2016.08.041>
32. Savi, G.D., Bortoluzzi, A.J. & Scussel, V.M. (2013). Antifungal properties of Zinc-compounds against toxigenic fungi and mycotoxin. *Int. J. Food Sci. Technol.* 48(9), 1834–1840. <http://dx.doi.org/10.1111/ijfs.12158>
33. Vitorino, H.A., Mantovanelli, L., Zanotto, F.P. & Esposito, B.P. (2015). Iron metalodrugs: stability, redox activity and toxicity against *Artemia salina*. *PLoS One.* 10(4), e0121997. <http://dx.doi.org/10.1371/journal.pone.0121997>
34. Arulvasu, C., Jennifer, S.M., Prabhu, D. & Chandhirasekar, D. (2014). Toxicity effect of silver nanoparticles in brine shrimp *Artemia*. *Sci. World J.* 2014, 256919. <http://dx.doi.org/10.1155/2014/256919>
35. Bortolotto, T., Bertoldo, J.B., da Silveira, F.Z., Defaveri, T.M., Silvano, J. & Pich, C.T. (2009). Evaluation of the toxic and genotoxic potential of landfill leachates using bioassays. *Environ. Toxicol. Pharmacol.* 28(2), 288–293. <http://dx.doi.org/10.1016/j.etap.2009.05.007>
36. Rodrigues, L.C.d.A., Barbosa, S., Pazin, M., Maselli, B.d.S., Beijo, L.A. & Kummrow, F. (2013). Fitotoxicidade e citogenotoxicidade da água e sedimento de córrego urbano em bioensaio com *Lactuca sativa*. *Rev. Bras. Eng. Agríc.* 17, 1099–1108.
37. Angele-Martinez, C., Nguyen, K.V., Ameer, F.S., Anker, J.N. & Brumaghim, J.L. (2017). Reactive oxygen species generation by copper(II) oxide nanoparticles determined by DNA damage assays and EPR spectroscopy. *Nanotoxicology.* 11(2), 278–288. <http://dx.doi.org/10.1080/17435390.2017.1293750>
38. Wang, F. & Sayre, L.M. (1989). Oxidation of tertiary amine buffers by copper(II). *Inorg. Chem.* 28(2), 169–170. <http://dx.doi.org/10.1021/ic00301a001>
39. Tachon, P. (1989). Ferric and cupric ions requirement for DNA single-strand breakage by H₂O₂. *Free Radic. Res. Commun.* 7(1), 1–10.
40. Burrows, C.J. & Muller, J.G. (1998). Oxidative Nucleobase Modifications Leading to Strand Scission. *Chem. Rev.* 98(3), 1109–1152.